

Laetiporus sulphureus mushroom enhances cytotoxic effect of *Bifidobacterium animalis* spp. *lactis* on HCT-116 cells in a co-culture system

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Abstract: The study aimed to test the effect of probiotic *Bifidobacterium animalis* spp. *lactis* (BAL) on the HCT-116 cell line viability, and to compare its effect with co-treatment BAL/*Laetiporus sulphureus* (EALS). The trypan blue staining method was used to estimate HCT-116 viability. The levels of NO₂⁻ were determined using 0.1% N-(1-naphthyl) ethylenediamine, as well as 1% sulfanilic acid. The determination of H₂O₂ was based on the oxidation of phenol red. Our results showed the significant cytotoxicity of the BAL on the HCT-116 cells in a co-culture system, while the BAL/EALS co-treatment further enhanced the cytotoxicity on the HCT-116 cells. We detected higher H₂O₂ and NO₂⁻ values in treatment with BAL, especially in co-treatment BAL/EALS. The death of the HCT-116 cells may be due to elevated levels of H₂O₂ and NO₂⁻ and their products (peroxynitrites).

Keywords: Edible mushroom; probiotics; *Bifidobacterium*; cancer cells; extract; H₂O₂; peroxynitrites

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1. Introduction

Colon cancer is one of the main health problems today. It is the second most common cancer diagnosed in women and the third in the male population [1].

Laetiporus sulphureus (Bull.) Murrill is an edible mushroom with sulfuric-yellow-coloured fruiting bodies [2]. *L. sulphureus* is an important source of various bioactive components such as; vitamins, carbohydrates, phenolic compounds, essential amino and fatty acids, and minerals. These components enable its biological activity [3]. It is known that certain components of mushrooms (such as polyphenols) interact with probiotics in the human colon due to their indigestion, and their mutual reaction is two-way.

Since the *L. sulphureus* mushroom has no cytotoxic effect on CRC cells [4, 2], it is necessary to examine the combined anticancer effect of *L. sulphureus* and probiotics. *Bifidobacterium animalis* spp. *lactis* (BAL) is a normal inhabitant of the human colon and a widely used probiotic. Many studies show that certain species of the *Bifidobacterium* genus can induce anticancer impact on colon cancer cells (Caco-2 and HT-29) [5].

The main aim of our study was to investigate the cytotoxic effect of BAL probiotics on the HCT-116 cell line and compare its effects with the BAL/EALS co-treatment. Also, we examined the levels of reactive species and compared them in both treatments. All experiments were performed in the modified HoxBan co-culture system.

2. Materials and Methods

Laetiporus sulphureus was gathered in the Adžine Livade at a height of 629 m (Šumadija region, Kragujevac, Serbia). The gathered mushroom material was dried in the air at room temperature. The 100 g of the mushroom fruiting body was soaked in 500 mL of ethyl acetate solvent. The finely dried ground mushroom material was macerated three times at room temperature every 48 h. Finally, the samples were filtered through sterile gauze and the filtrates were evaporated to dryness using the evaporator (IKA, Sindelfingen, Germany). The dry ethyl acetate extracts (EALS) were stored at -18°C before use [6].

Bifidobacterium animalis spp. *lactis* (BB-12) was obtained from the Microbiology Laboratory, Institute for Information Technologies, Serbia. The preparation of probiotic suspension was previously described in detail in the paper by Andrews et al. [8]. The turbidity of the initial suspension was measured by using the McFarland densitometer (Biosan, Latvia).

The HCT-116 cell line was obtained from ATCC (Manassas, VA, USA). The cells were cultured in Dulbecco's modified Eagle's minimal essential medium (DMEM) supplemented with fetal bovine serum and antibiotics. The HCT-116 cell viability was determined with trypan blue staining 12 and 24 h after incubation in the co-culture system [7].

The modified HoxBan co-culture system was formed in test tubes. The initial bacterial suspension contained about 108 colony-forming units (CFU/mL). Then, 1:4 dilutions of start suspension were additionally prepared in sterile 0.85% saline. To finish, 40 μL of BAL suspension was inoculated into 40 mL of sterile Muellere-Hinton soft medium. The coverslips with untreated cells were turned towards the agar surface without BAL–negative control (C_0); or with BAL in agar–positive control (C). In both negative and positive control, 10 mL of DMEM with 10% FBS and without antibiotics were added. HCT-116 cells on the surface of the co-culture system and BAL in the bacterial medium in the coculture overlaid with 10 mL of EARS treatment. The detailed procedure was described in the manuscript by Arsenijevic et al. [9].

The levels of NO_2^- were determined using 0.1% N-(1-naphthyl) ethylenediamine, and 1% sulfanilic acid according to protocol Šeklić et al. [6]. On the other hand, determination of H_2O_2 was performed using horseradish peroxidase [10].

Statistical analyses were performed using the ANOVA method in statistical software SPSS (Windows, ver. 17, 2008, Chicago, IL, USA). $p < 0.05$ is considered a statistically significant difference (*#).

3. Results and Discussion

The significant cytotoxicity effect of the BAL on HCT-116 cell viability in a co-culture system was shown (Figure 1). Our results are in a positive correlation with studies by Nowak et al. [11], and Faghfoori et al. [5]. These authors showed the cytotoxic effect of BAL on both HT-29 and Caco-2 cells. The Viability of HCT-116 cells was additionally decreased after incubation with the BAL/EALS co-treatment. It is especially noted at the EALS concentration of 50 $\mu\text{g}/\text{mL}$ (Figure 1). Since the EALS extract had no cytotoxic activity, as shown by the results of Jovanovic et al. [4] and Younis et al. [2], we can conclude that noncytotoxic EALS extract enhances the cytotoxic effect on HCT-116 cells in co-culture with the BAL. The combined mechanism of action of BAL/EALS will be additionally investigated in future studies.

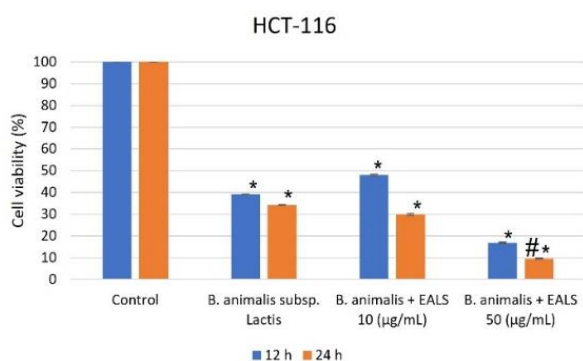


Figure 1. The percentage of HCT-116 viable cells in the co-culture system after incubation with BAL and BAL/EALS (conc. 10 and 50 µg/mL). * A stat. significance difference between Control and treatments at 12 and 24 h of incubation, and; # A stat. significance difference between BAL/EALS 10 µg/mL and BAL/EALS 50 µg/mL after 24 h.

Cancer cells survive high levels of ROS and RNS by increasing antioxidant status to optimize ROS/RNS-induced proliferation, and at the same time avoid thresholds of these parameters that would trigger cell death [12, 13]. However, disproportionately increased RNS and ROS levels can induce death in cancer cells. Our study demonstrated that the probiotic BAL caused an increase of H₂O₂ and NO₂⁻ levels in the HCT-116 after 24 h of incubation (Figure 2A). By the incubation of the HCT-116 with BAL/EALS (10 and 50 µg/mL), tested parameters were significantly increased especially after 12 h (Figure 2B). These results indicate that BAL/EALS treatments caused acute oxidative stress in the HCT-116 cells. H₂O₂ and nitrites generate peroxy-nitrites. Then, the contact between peroxy-nitrites and residual H₂O₂ generates primary singlet oxygen. These molecules can cause local inactivation of protective catalase in cancer cells, and H₂O₂ and peroxy-nitrites survive at the site of local inactivation. This leads to the generation of secondary singlet oxygen through the interaction between H₂O₂ and peroxy-nitrites and catalase inactivation. Finally, the deactivation of catalases allows the influx of H₂O₂ through aquaporins, leading to intracellular glutathione depletion and sensitization of the cancer cells for cell death induction through lipid peroxidation [14].

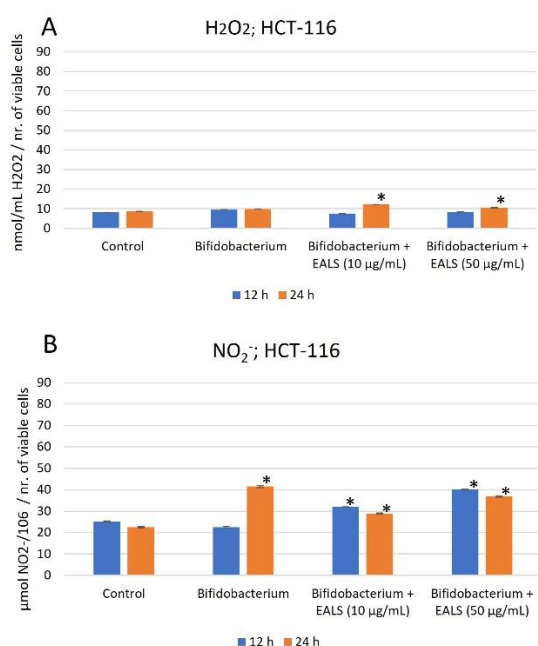


Figure 2. Levels of H₂O₂ (A) and NO₂⁻ (B) parameters in HCT-116 cell line after treated by BAL, and BAL/EALS (conc. 10 and 50 µg/mL). * A statistical significance difference between control and treatments at 12 and 24 h.

4. Conclusions

Our study demonstrated the viability of colorectal cancer cells (HCT-116), after incubation with the probiotic BAL and compared its effects with co-treatment BAL/EALS. Our study demonstrated a significant reduction of HCT-116 viability in the co-culture system with BAL after 12 h of incubation. However, we observed that in the co-cultures BAL and EALS extract the viability of HCT-116 was further reduced. The strongest cytotoxic effect was noticed after incubation with BAL and EALS in a concentration of 50 µg/mL. Death of the HCT-116 cells is due to elevated levels of H₂O₂, NO₂⁻ and their co-products (peroxynitrites).

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References

1. Ferlay, J.; Colombet, M.; Soerjomataram, I.; Dyba, T.; Randi, G.; Bettio, M.; Gavin, A.; Visser, O.; Bray, F. Cancer incidence and mortality patterns in Europe: Estimates for 40 countries and 25 major cancers in 2018. *European Journal of Cancer*. **2018**, 1 – 32.
2. Younis, A.M.; Yosric, M.; Stewarte, J.K. In vitro evaluation of pleiotropic properties of wild mushroom *Laetiporus sulphureus*. *Ann. Agric. Sci.* **2019**, 64, 79–87.
3. Adamska, I. The possibility of using sulphur shelf fungus (*Laetiporus sulphureus*) in the food industry and in medicine - A review. *Foods*. **2023**, 12, 1539.
4. Jovanović, M. M.; Marković, K. G.; Grujović, M. Ž.; Pavić, J.; Mitić, M.; Nikolić, J.; Šeklić, D. Anticancer assessment and antibiofilm potential of *Laetiporus sulphureus* mushroom originated from Serbia. *Food Science and Nutrition*. **2023**, fsn3.3577.
5. Faghfoori, Z.; Faghfoori, M. H.; Saber, A.; Izadi, A.; Yari Khosroushahi, A. Anticancer effects of bifidobacteria on colon cancer cell lines. *Cancer Cell International*. **2021**, 21, 258.
6. Šeklić, D.S.; Stanković, M.S.; Milutinović, M.G.; Topuzović, M.D.; Štajn, A.Š.; Marković, S.D. Cytotoxic, antimigratory, pro-and antioxidative activities of extracts from medicinal mushrooms on colon cancer cell lines. *Arch. Biol. Sci.* **2016**, 68, 93–105.
7. Sadabad, S.M.; Martels, Z.Y.; Khan, T.M.; Blokzijl, T.; Paglia, G.; Dijkstra, G.; Harmsen, M.H.; Faber, N.K. A simple coculture system shows mutualism between anaerobic faecalibacteria and epithelial cells. *Scientific Reports*. **2015**, 5, 1–9.
8. Andrews, J. M. BSAC standardized disc susceptibility testing method (version 4). *Journal of Antimicrobial Chemotherapy*. **2005**, 56, 60-76.
9. Arsenijević, D.; Jovanović, M.; Pecić, K.; Grujović, M.; Marković, K.; Šeklić, D. Effects of *Laetiporus sulphureus* on Viability of HeLa Cells in Co-Culture System with *Saccharomyces boulardii*. *Biol. Life Sci. Forum* **2022**, 18, 69.
10. Pick, E.; Keisari, Y. A simple colorimetric method for the measurement of hydrogen peroxide produced by cells in culture. *Journal of Immunological Methods*. **1980**, 38, 161 – 170.
11. Nowak, A.; Paliwoda, A.; Blasiak, J. Anti-proliferative, pro-apoptotic and anti-oxidative activity of Lactobacillus and Bifidobacterium strains: A review of mechanisms and therapeutic perspectives. *Critical Reviews in Food Science and Nutrition*. **2019**, 59, 3456–3467.

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12. Dodson, M.; Portuguese, C. R.; Zhang, D.D. NRF2 plays a critical role in mitigating lipid peroxidation and ferroptosis. *Redox Biology*. **2019**, *23*, 101107.
 13. Hayes, D. J.; Kostova, D.A.; Tew, D.K. Oxidative Stress in Cancer. *Cancer Cell*. **2020**, *38*, 167–197.
 14. Bauer, G. The synergistic effect between hydrogen peroxide and nitrite, two long-lived molecular species from cold atmospheric plasma, triggers tumor cells to induce their own cell death. *Redox Biology*. **2019**, *26*, 101291.