

The 1st International Electronic **Conference on Toxics**

20-22 March 2024 | Online



🗖 2D

🗖 3D

Exploring marine-derived fungal preussin toxicity on MDA-MB-231 cells cultured in 2D and 3D

Rosária Seabra^{1,2,*}, Fernanda Malhão^{1,2}, Alexandra Correia^{1,3}, Carla Costa^{4,5}, Anake Kijjoa^{1,2}, Eduardo Rocha^{1,2}

¹ICBAS – Instituto de Ciências Biomédicas Abel Salazar, Universidade do Porto, Porto, Portugal; ²CIIMAR – Centro Interdisciplinar de Investigação Marinha e Ambiental, Universidade do Porto, Matosinhos, Portugal; ³I3S – Instituto de Investigação e Inovação em Saúde, Universidade do Porto, Porto, Portogal; ⁴INSA – National Institute of Health Dr. Ricardo Jorge, Porto, Portugal; ⁵ITR – Laboratório para a Investigação Integrativa e Translacional em Saúde Populacional, EPIUnit – Instituto de Saúde Pública da Universidade do Porto, Porto, Portugal; *rcseabra@icbas.up.pt

Introduction & aim

Triple-negative breast cancer (TNBC) has a poor prognosis, and limited therapeutic options¹, which allied with chemotherapy-associated toxicity and drug resistance call for the search for new molecules that may serve as new drugs, drug adjuvants, or scaffolds for drug development.

Preussin (Pr) is a natural compound shown to decline cell viability and proliferation and induce cell death and cell cycle arrest, mainly in bidimensional (2D) cell cultures of various cell lines ^{2–4}.

Three-dimensional (3D) cell cultures are described to better mimic the tumor behaviour in vivo⁵.

This work aimed to explore better the effects of Pr on cell viability, proliferation, death induction, genotoxicity, and migration of a TNBC cell line, comparing its effects in 2D and 3D cell cultures.



Results & discussion



Fig. 1. Cell viability of MDA-MB-231 cell line after 24 and 72 h (2D cultures) (A) or 96 h (3D cultures) (B) of exposure to Pr, C or positive control of doxorrubicin (Dx).





Statistical analysis

- n = 5 independent experiments
- Normality and homogeneity of variance : Shapiro-Wilk's and Levene's tests
- Differences from the control group (C): One-way ANOVA followed by Holm-Šídák test
- Data are presented as mean and standard deviation. Significant differences are shown with asterisks (* p < 0.05; ** p < 0.01; p < 0.001; **** p < 0.001)

Acknowledgments

Strategic funding UIDB/04423/2020 and UIDP/04423/2020 through national funds provided by FCT—Fundação para a Ciência e a Tecnologia to CIIMAR and the Master in Oncology of the ICBAS—U.Porto

References

1. Giaquinto, A.N; et al. Breast Cancer Statistics, 2022. CA Cancer J Clin 2022, 72, 524–541, doi:10.3322/caac.21754. 2. Buttachon, S.; et al. Bis-Indolyl Benzenoids, Hydroxypyrrolidine Derivatives and Other Constituents from Cultures of the Marine Sponge-Associated Fungus Aspergillus Candidus KUFA0062. Mar Drugs 2018, 16, 119, doi:10.3390/md16040119. 3. Malhão, F.; et al. Cytotoxic and Antiproliferative Effects of Preussin, a Hydroxypyrrolidine Derivative from the Marine Sponge-Associated Fungus Aspergillus Candidus KUFA 0062, in a Panel of Breast Cancer Cell Lines and Using 2D and 3D Cultures. Mar Drugs 2019, 17, 448, doi:10.3390/md17080448.

4. Achenbach, T; et al. Inhibition of Cyclin-Dependent Kinase Activity and Induction of Apoptosis by Preussin in Human Tumor Cells. Antimicrob Agents Chemother 2000, 44, 2794–2801, doi:10.1128/AAC.44.10.2794-2801.2000.

5. Sant, S.; Johnston, P.A. The Production of 3D Tumor Spheroids for Cancer Drug Discovery. Drug Discov Today Technol 2017, 23, 27–36, doi:10.1016/j.ddtec.2017.03.002.

to C (A) or, 25 (B) or, 35 µM (C) of Pr. Blue arrow: cell debris; Red arrow: cells with morphology compatible with apoptosis; Yellow arrow: multivesicular dense bodies.

With the increase in concentrations of Pr, there was a rise in cell debris, intracellular multivesicular dense bodies, and cells with morphology compatible with apoptosis in both culture models (data from the 2D culture was not shown).

Conclusion

