

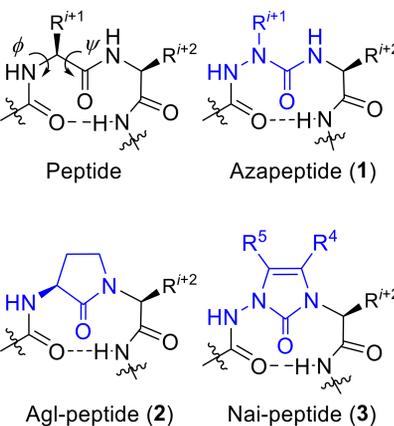
Abstract

N-Aminoimidazol-2-one (Nai) residues adopt peptide β - and γ -turns. 4-, 5- and 4,5-Substituted Nai-peptides were synthesized using a common route featuring proline-catalyzed condensations of aldehydes and ketones onto azopeptides. The bioactive conformer of a cluster of differentiation 36 receptor (CD36) modulator has been identified using Nai-peptide ligands.

Introduction: Nai peptide conception

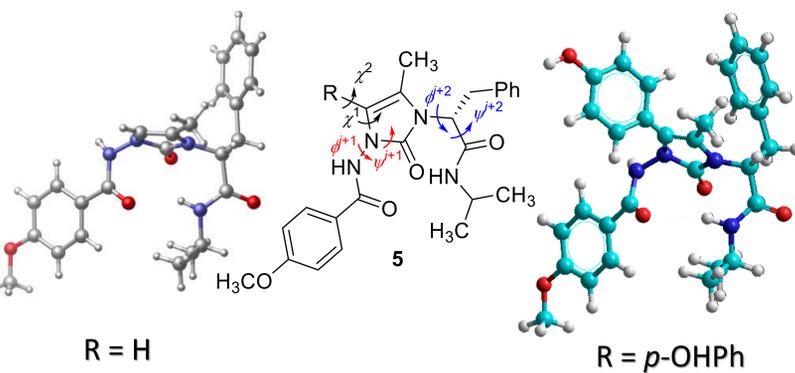
Peptide turns warrant mimicry to study implications in molecular recognition. **Aza-** and **lactam-peptides 1** and **2** favor turn geometry by way of stereo-electronic and covalent effects [1, 2]. Combining such properties, substituted *N*-aminoimidazol-2-one (**Nai, 3**) residues offer potential to mimic turn backbone and side chain geometry and function [3].

Approaches for β -Turn mimicry



Nai residues induce turn geometry

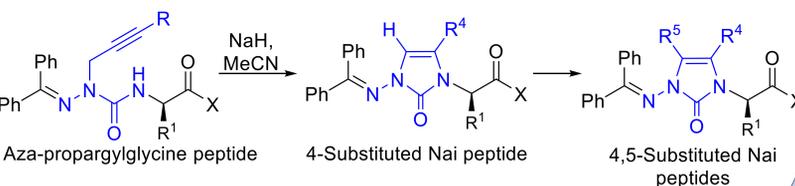
In model peptides, 4- and 4,5-substituted Nai residues induced turn geometry as observed by NMR spectroscopy, X-ray crystallography, and computational analysis [5,6,10]. 4-Methyl, 5-aryl Nai residues replicate natural side chain orientation on β -turn conformations [5,6,10].



Peptide	ϕ^{i+1}	ψ^{i+1}	ϕ^{i+2}	ψ^{i+2}	χ^1	χ^2
Ideal type II' β -turn	60	-120	-80	0	-	-
<i>p</i> -MeOBz-4-Me-Nai-D-Phe-NH <i>i</i> Pr (4)	58.9	-153.3	-69.1	-4.6	-	-
<i>p</i> -MeOBz-4-Me,5- <i>p</i> -HOPh-Nai-D-Phe-NH <i>i</i> Pr (5)	48.6	-143.7	-62.4	33.6	-41.1	76.0

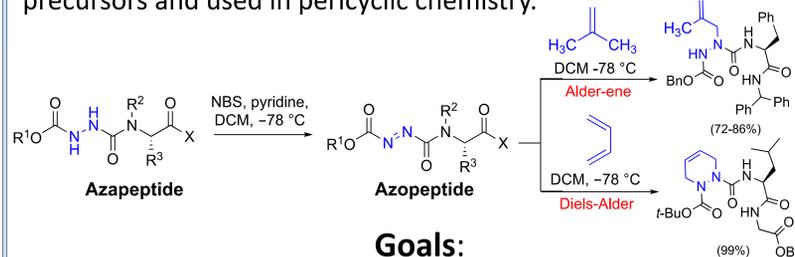
Earlier Nai syntheses

4-Methyl Nai residues were made from azaPra peptides by base-promoted 5-*endo-dig* cyclization and olefin isomerization. Embellishment by Sonogashira ($R^4 = CH_2Ar$), Pd-catalyzed arylation ($R^5 = Ar$) and Vilsmeier-Haack formylation ($R^5 = CHO$) chemistry gave various 4- and 4,5-substituted Nai analogs [4-7,10]. Ester epimerization during alkaline-mediated cyclization was suppressed using amide and acid counterparts [6,7,10].



Azopeptide synthesis and pericyclic chemistry [8]

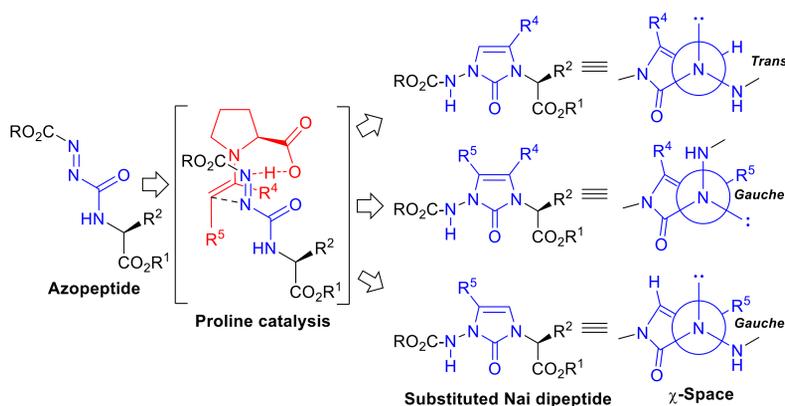
Azopeptides have been synthesized by oxidation of aza-glycine precursors and used in pericyclic chemistry.



Goals:

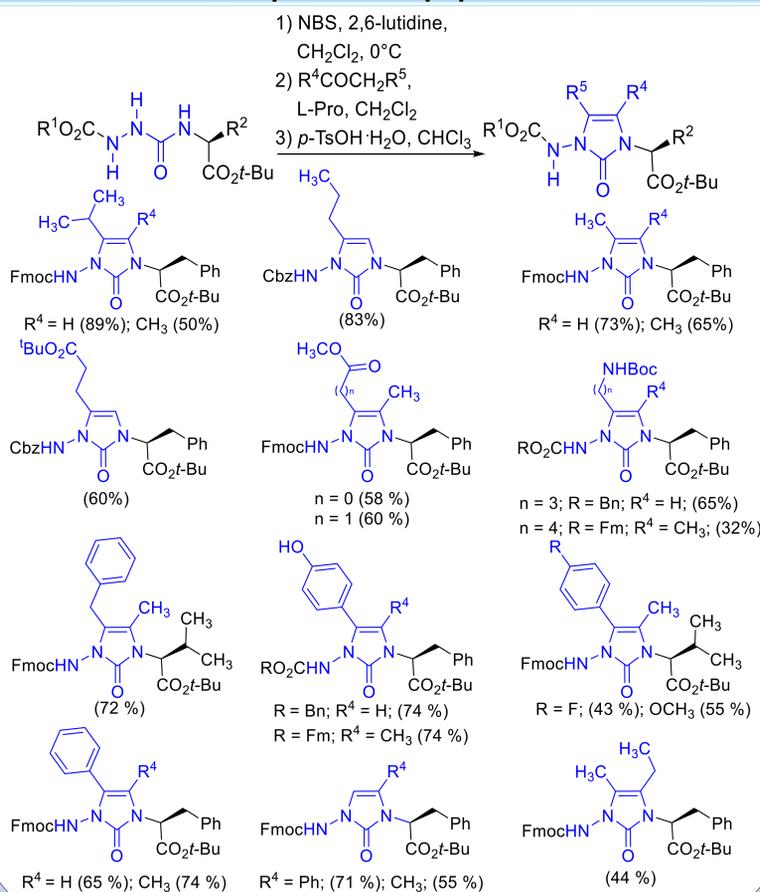
- Synthesis of 4-, 4,5- and 5-substituted Nai residues from a common azopeptide approach
- Biomedical application of Nai-peptides

Azopeptide approach to substituted Nai residues [9-11]



A common azopeptide approach features organocatalyzed reactions with carbonyl components. Proline-catalyzed enamine addition onto azopeptides gives selectively α -nitrogen alkylation to provide γ -oxo aza-amino amides. Subsequent, intramolecular cyclative acid-mediated dehydration gives diverse 4-, 5- and 4,5-substituted Nai-dipeptides with potential to respectively explore *gauche* and *trans* side chain orientations [9-11].

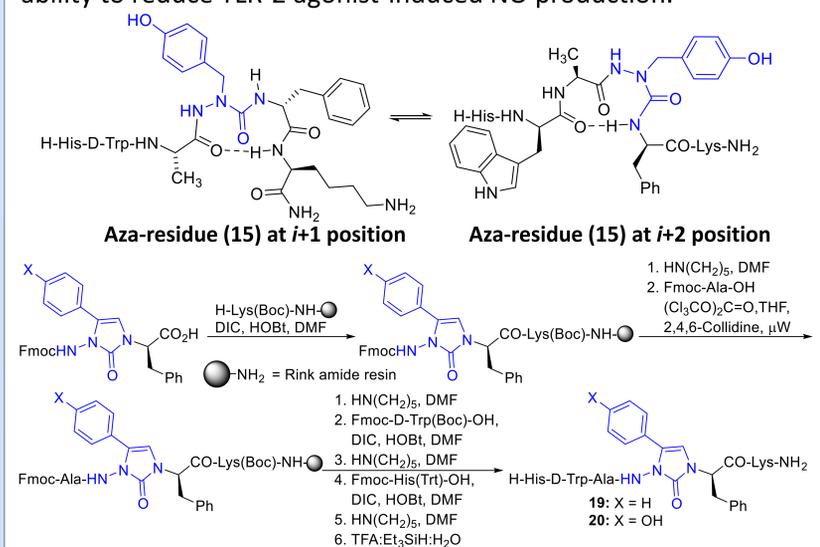
Scope of Nai dipeptides



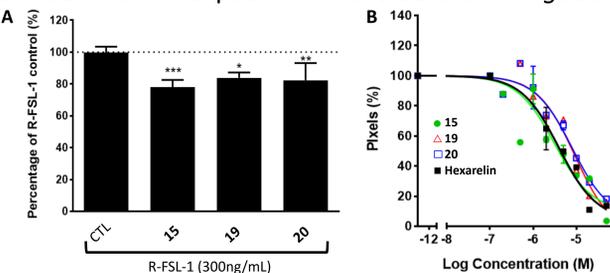
[5-Aryl-Nai⁴]GHRP-6 Analogs CD36 modulation

H-His-D-Trp-Ala-Trp-D-Phe-Lys-NH₂; **GHRP-6**
 H-His-D-Trp-Ala-**azaTyr**-D-Phe-Lys-NH₂; [aza-Tyr⁴]-GHRP-6 (**15**)

Azapeptide analogs of GHRP-6 have been pursued as selective CD36 modulators [9,10,12]. For example, azapeptide **15** exhibits relatively high CD36 binding affinity, curbs macrophage-driven inflammation and mitigates angiogenic and atherosclerotic pathology [9,10,12]. The bioactive conformer of azapeptide **15** was studied by the solid-phase synthesis of [5-Aryl-Nai⁴]GHRP-6 analogs **19** and **20** and examination of CD36 binding affinity and ability to reduce TLR-2 agonist-induced NO production.



5-Aryl-Nai analogs **19** and **20** had similar effects on NO production (**A**) and bound CD36 with 2.6- to 3.2-fold lower affinity (**B**) as azapeptide **15** indicating a likely common β -turn conformer with the aza-residue in the *i*+1 position and side chain in *gauche* chi-space.



Conclusions

- 4-, 4,5- and 5-Substituted Nai β -turn mimics were synthesized from azopeptides by a common proline-catalyzed route.
- 5-Aryl-Nai residue incorporation into CD36 modulator **15** indicates bioactive β -turn backbone and side chain geometry.

References

- Chingle, R.; Proulx, C.; Lubell, W.D. *Acc. Chem. Res.* **2017**, *50*, 1541–1556.
- Freidinger, R. M.; Veber, D. F.; Perlow, D. S.; Brooks, J. R.; Saperstein, R. *Science*, **1980**, *210*, 656–658.
- St-Cyr, D. J.; García-Ramos, Y.; Doan, N. D.; Lubell, W. D. *Peptidomimetics – I*, Springer, **2017**, 125–175.
- Proulx, C.; Lubell, W. D. *Org. Lett.*, **2012**, *14*, 4552.
- Proulx, C.; Lubell, W. D. *Peptide Science*, **2014**, *102*, 7-15.
- Poupart, J.; Doan-Ngoc, D.; Bérubé, D.; Hamdane, Y.; Medena, C.; Lubell, W. D. *Heterocycles*, **2019**, *99*, 279-293.
- Poupart, J.; Hamdane, Y.; Lubell, W. D. *Can. J. Chem.* **2020**, *98*, 278-284.
- Chingle, R.; Lubell, W.D. *Org. Lett.*, **2015**, *17*, 5400–5403.
- Hamdane, Y.; Chauhan, S. P.; Vutla, S.; Mulumba, M.; Ong, H.; Lubell, W. D. *Org. Lett.* **2021**, *23* (9), 3491–3495.
- Hamdane, Y.; Poupart, J.; Lubell, W. D., *Synthesis*, **2022**, *54*, A-1.
- Hamdane, Y.; Truong, D.; Lubell, W. D. **2024** [in preparation]
- Mellal, K.; Omri, S.; Mulumba, M.; Tahiri, H.; Fortin, C.; Dorion, M.-F.; Pham, H.; Ramos, Y.G.; Zhang, J.; Pundir, S.; Joyal, J.-S.; Bouchard, J.-F.; Sennlaub, F.; Febbraio, M.; Hardy, P.; Gravel, S.-P.; Marleau, S.; Lubell, W.D.; Chemtob, S.; Ong, H. *Sci. Rep.* **2019**, *9*, 12903.

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