





# Identification of Lactobacillus reuteri P2M1 from Swine and **Evaluation of its Probiotic Function in Vitro**

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# INTRODUCTION

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The post-antibiotic era, characterized by rising antimicrobial resistance, demands innovative strategies to enhance livestock health. Probiotics, particularly *lactic acid bacteria* (LAB), offer multifaceted benefits, including pathogen inhibition, immune modulation, and oxidative stress mitigation. Limosilactobacillus reuteri (L. reuteri), a gutadapted LAB, has shown promise in improving feed efficiency and intestinal barrier function.

### AIM

▲ Identified the swine-derived *L. reuteri* P2M1 strain through biochemical and molecular methods.

# **METHOD**

#### **A**Strain Isolation and Characterization.

The strain isolated from healthy pig feces was biochemically identified, and molecular biological identification was performed using bacterial 16S rDNA sequence PCR amplification and sequencing techniques.

#### **Functional Assays.**

The growth curve and acid production curve of the strain were drawn to evaluate the tolerance of the strain to acid and bile salts, and the sensitivity of the strain to antibiotics was detected. The free radicalscavenging ability and extracellular polysaccharide production of the strain were determined

▲ Evaluated its probiotic potential, including acid/bile tolerance, antibiotic sensitivity, antioxidant activity, and safety.

#### **A**Safety Evaluation.

The in vitro safety test was performed to evaluate the safety of the strain to mice.

#### **RESULTS**

#### Table 1 Biochemical identification results of strain P2M1.

| Items                  | Results | Items                        | Results |
|------------------------|---------|------------------------------|---------|
| Sucrose                | +       | Xylose                       | -       |
| Ribose                 | -       | Lactose                      | +       |
| Trehalose              | -       | Galactose                    | +       |
| Glucose                | +       | Cellobiose                   | +       |
| Melezitose             | -       | Fructose                     | -       |
| Indole test            | -       | Gelatin of liquefaction test | -       |
| Nitrate reduction test | -       | Hydrogen peroxide test       | -       |

+: Positive result; -: Negative result.





| Table 2 Antibiotic sensitivity of L.reuteri P2M1. |                     |              |  |  |
|---|---------------------|--------------|--|--|
| Antibiotic  | Inhibition diameter | Sensitivity  |  |  |
| Amikacin  | $8.40 \pm 0.14$     | Resistance   |  |  |
| Clindamycin                                       | $12.43 \pm 0.19$    | Resistance   |  |  |
| Erythromycin                                      | $20.52 \pm 0.08$    | Sensitive    |  |  |
| Lincomycin  | $13.49 \pm 0.30$    | Resistance   |  |  |
| Enrofloxacin                                      | $7.59 \pm 0.25$     | Resistance   |  |  |
| Roxithromycin                                     | $16.89 \pm 0.16$    | Intermediate |  |  |
| Ampicillin  | $21.72 \pm 0.47$    | Sensitive    |  |  |
| Amoxicillin                                       | $32.89 \pm 0.62$    | Sensitive    |  |  |
| Cefuroxime  | 22.52 ± 1.11        | Sensitive    |  |  |
| Cefazolin   | $27.54 \pm 0.51$    | Sensitive    |  |  |

• The strain P2M1 exhibited glycolysis ability, the logarithmic growth phase was 2-10 h, and the pH was stable between 4.0 and 4.1 after 20 h. It has good low-pH and bile salt tolerance, and is sensitive to common antibiotics.

• The DPPH, ABTS and hydroxyl radical-scavenging rates of the fermentation supernatant of the strain were significantly higher than those of the intact cell suspension (P < 0.05), which were 88.79%, 93.52% and 92.15%, respectively.



• No death or organ lesions were observed in mice after intragastric administration of 0.2 mL of viable L. reuteri P2M1 solution at a concentration of 1×10<sup>9</sup> CFU/mL per day for two weeks.



L. reuteri P2M1 has a short logarithmic growth period, rapid growth of cell concentration, high yield of extracellular polysaccharides, good probiotic and antioxidant properties, and safety in vitro, and can be used as a candidate strain for probiotic preparations.

**CONCLUSION** 

1. Vougiouklaki, D., et al. (2023). Probiotic properties and antioxidant activity in vitro of lactic acid bacteria. Microorganisms, 11(5), 1264. 2. Yang, S., et al. (2023). Exopolysaccharides from lactic acid bacteria, as an alternative to antibiotics, on regulation of intestinal health and the immune system. Animal Nutrition, 13, 78–89.