

21–23 May 2025 | Online

## Assessing the Antimicrobial Activity of Gemmotherapy Extracts

Melinda Héjja<sup>1,2</sup>, Éva György<sup>3</sup>, Ferenc Ádám Lóga<sup>4</sup>, Tünde Pacza<sup>2</sup>, Róbert Nagy<sup>2</sup>, Endre Máthé<sup>2</sup>

<sup>1</sup>Doctoral School of Nutrition and Food Science, Faculty of Agricultural and Food Sciences and Environmental Management, University of Debrecen, Böszörök Str. 128, HU-4032 Debrecen, Hungary

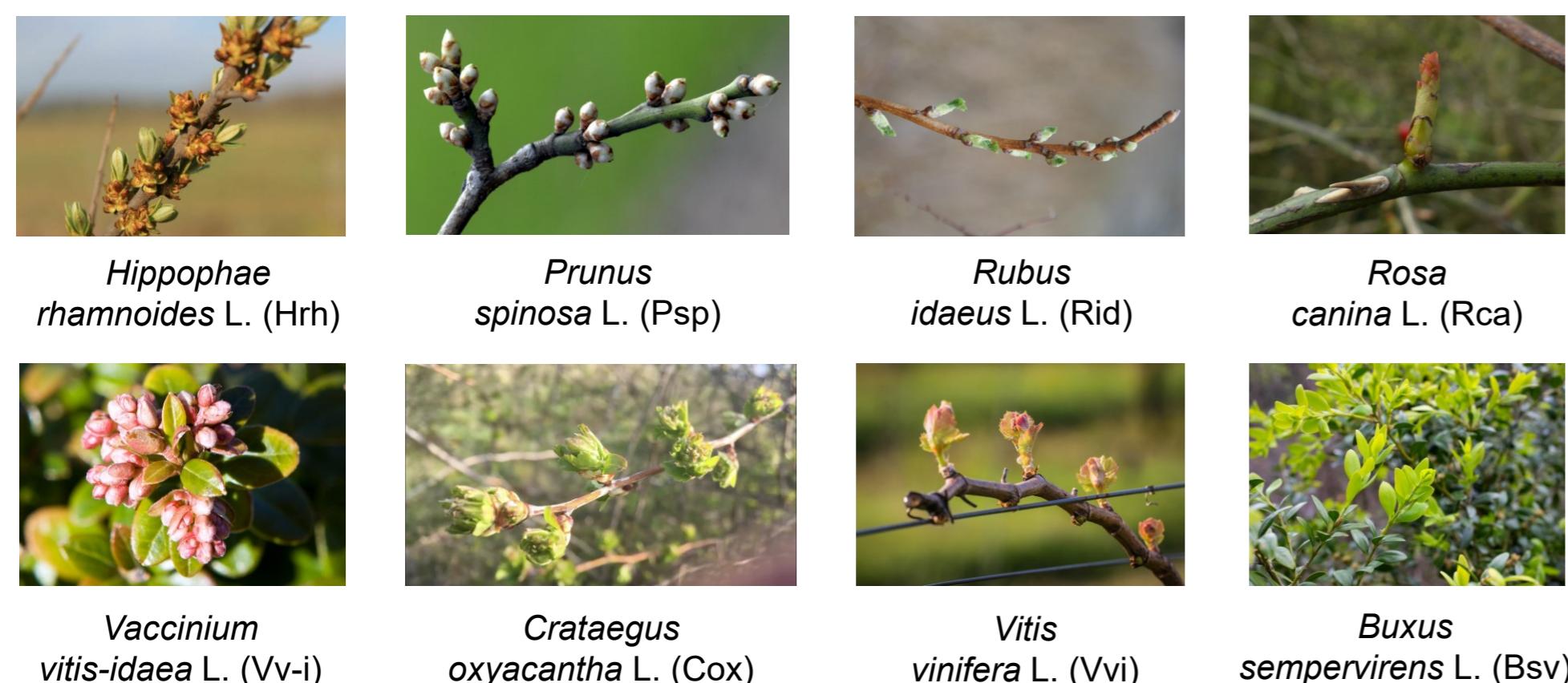
<sup>2</sup>Institute of Nutrition Science, Faculty of Agricultural and Food Sciences and Environmental Management, University of Debrecen, Böszörök Str. 128, HU-4032 Debrecen, Hungary

<sup>3</sup>Department of Food Science, Faculty of Economics, Socio-Human Sciences and Engineering, Sapientia Hungarian University of Transylvania, Libertății sq. 1, RO-530104 Miercurea Ciuc, Romania

<sup>4</sup>Biofrutti, Széchenyi street no. 68., HU-4100 Berettyóújfalu, Hungary

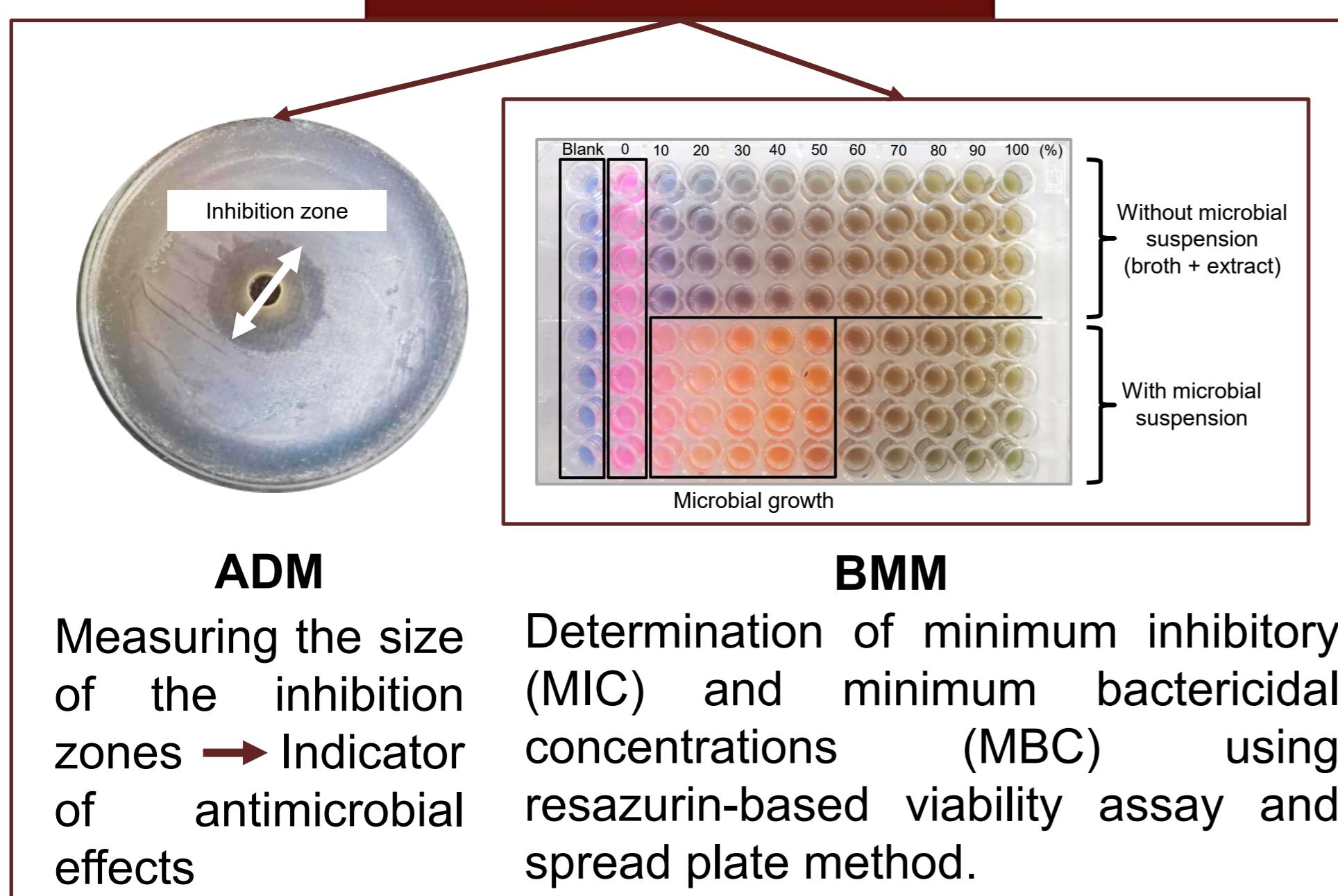
### INTRODUCTION & AIM

**Antibiotic resistance** is a growing global health concern, highlighting the need for novel antimicrobial agents. **Gemmotherapy extracts (GTE)** — derived from buds and young shoots rich in secondary metabolites, enzymes, hormones, trace elements, cytokines — may offer a more potent alternative to traditional fruit extracts. This study investigates the **antimicrobial activity** of **8 bud extracts** against **14 microorganisms** using **agar diffusion (AD)** and **broth microdilution (BM)** methods, and analyzes their **polyphenol, flavonoid, and antioxidant** content.



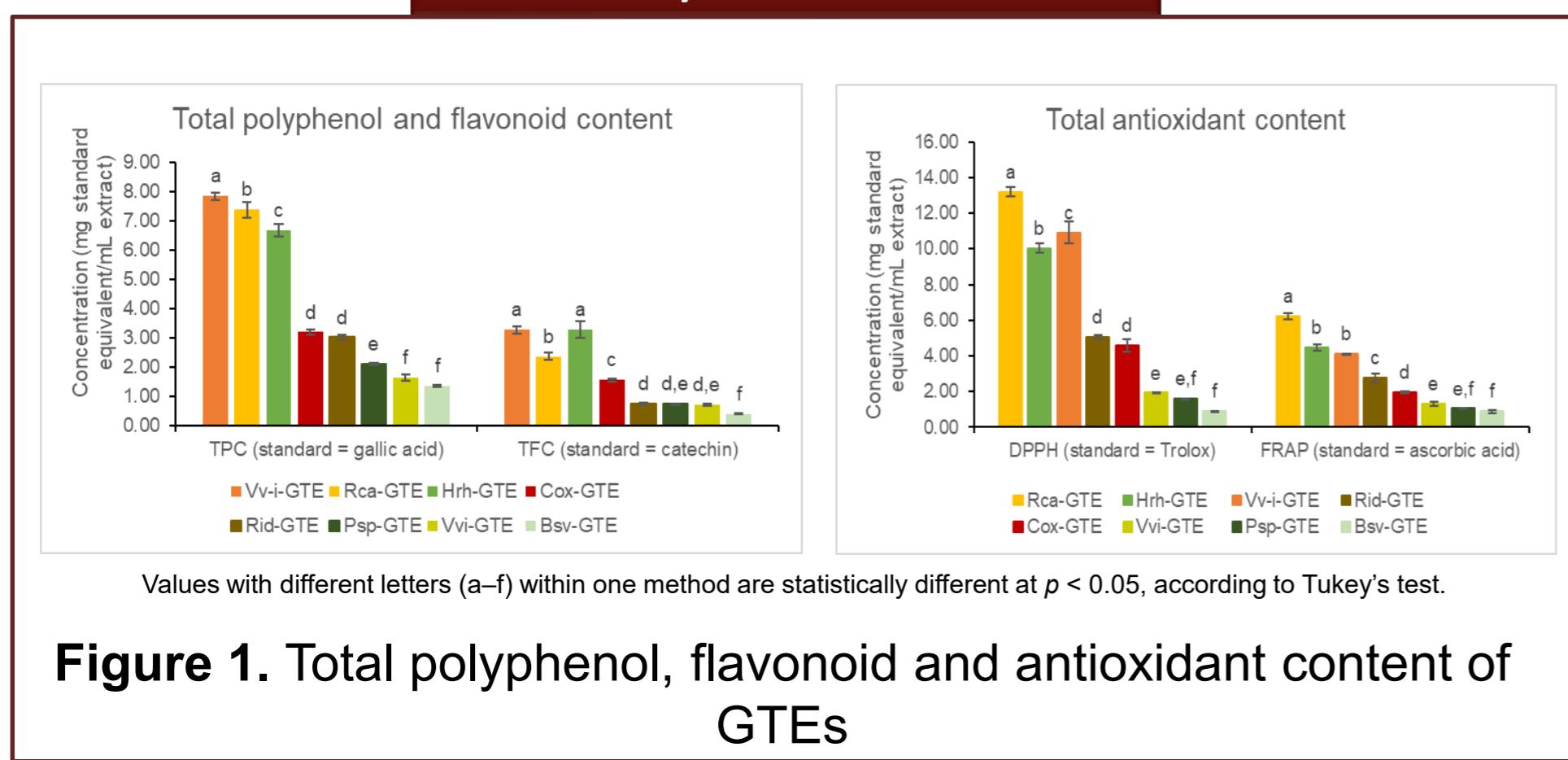
### METHOD

#### Antimicrobial tests



### RESULTS & DISCUSSION

#### Analytical results



**Table 1.** Minimum concentration of the extract (%) that produces a visible inhibition zone

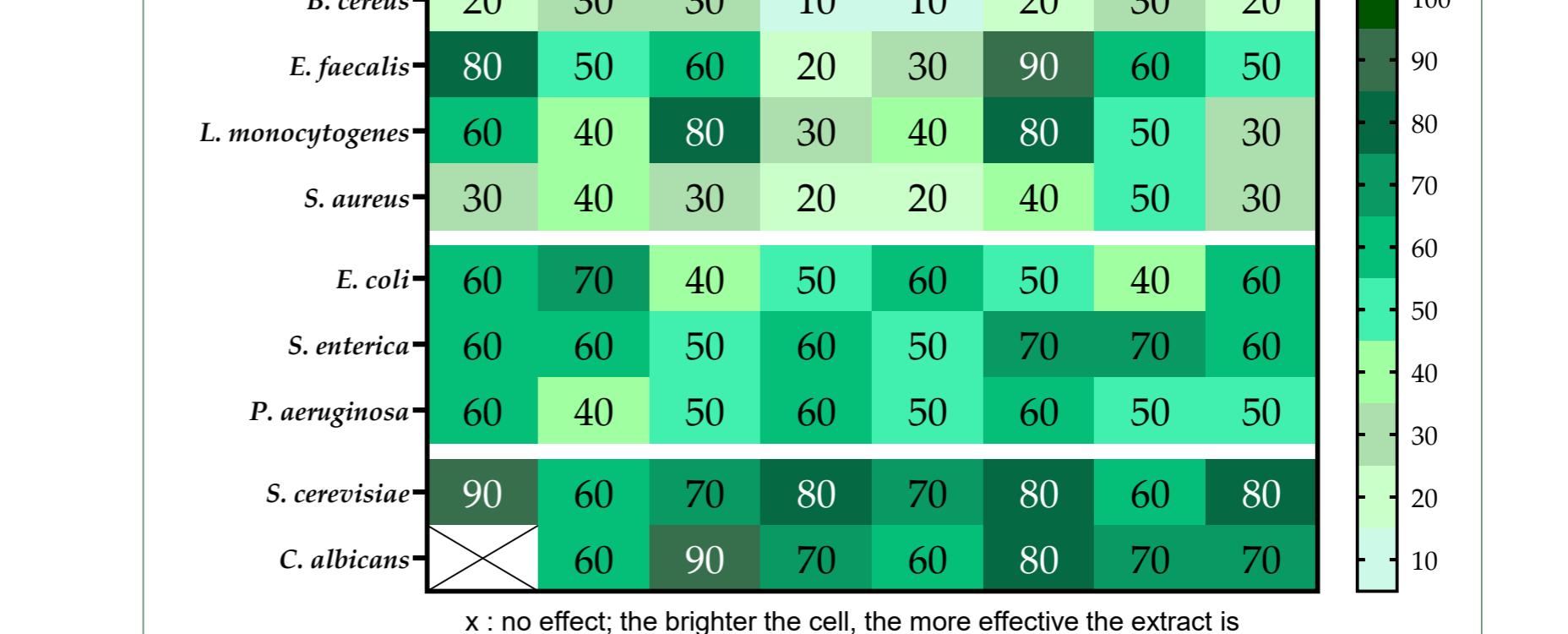
Microorganisms	Rid GTE	Psp GTE	Hrh GTE	Rca GTE	Vv-i GTE	Cox GTE	Bsv GTE	Vvi GTE
<i>Bacillus cereus</i> B.00076	nd	70	40	10	30	50	nd	100
<i>Enterococcus faecalis</i> B.01054	nd	nd	50	50	30	60	nd	nd
<i>Listeria monocytogenes</i> wild strain	nd	nd	100	10	40	nd	nd	nd
<i>Staphylococcus aureus</i> B.01055	70	70	40	20	10	20	nd	nd

nd – not detectable

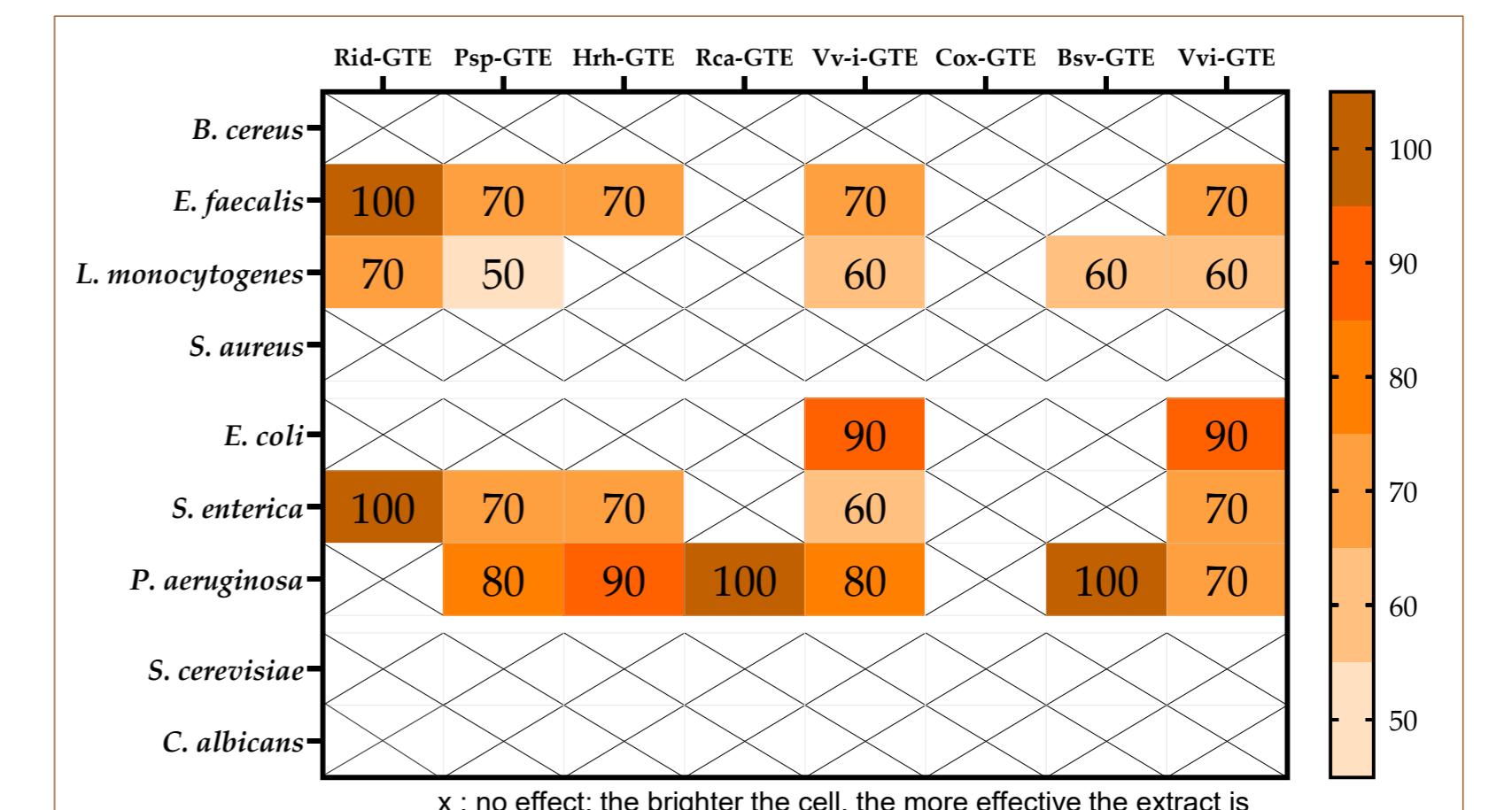
Resistant microorganisms:	Gram-negative bacteria	Yeasts	Molds
	<ul style="list-style-type: none"> <li><i>Escherichia coli</i> B.00200</li> <li><i>Salmonella enterica</i> subs. <i>enterica</i> B.00834</li> <li><i>Pseudomonas aeruginosa</i> B.01064</li> </ul>	<ul style="list-style-type: none"> <li><i>Saccharomyces cerevisiae</i> Y.00481</li> <li><i>Candida albicans</i> wild strain</li> </ul>	<ul style="list-style-type: none"> <li><i>Aspergillus niger</i> F.00071</li> <li><i>Aspergillus ochraceus</i> F.00850</li> <li><i>Aspergillus flavus</i> F.00048</li> <li><i>Penicillium expansum</i> F.00601</li> <li><i>Penicillium citrinum</i> F.00815</li> </ul>

### Agar diffusion method

**Table 1.** Minimum concentration of the extract (%) that produces a visible inhibition zone



**Figure 2.** Minimum inhibitory concentrations (%)



**Figure 3.** Minimum bactericidal concentrations (%)

### CONCLUSION

The examined extracts exhibit antimicrobial effects and may serve as useful adjuncts to antibiotic therapies.

Extracts of dog rose, lingonberry, and sea buckthorn exhibit the highest antioxidant and polyphenol contents and effectively inhibit the growth of most bacterial strains as determined by the AD and MIC methods.

### FUTURE WORK

Assessing the antimicrobial activity of mixed GTEs.

Assessing nutrigenomic effects in diabetic and leaky gut animal models.