

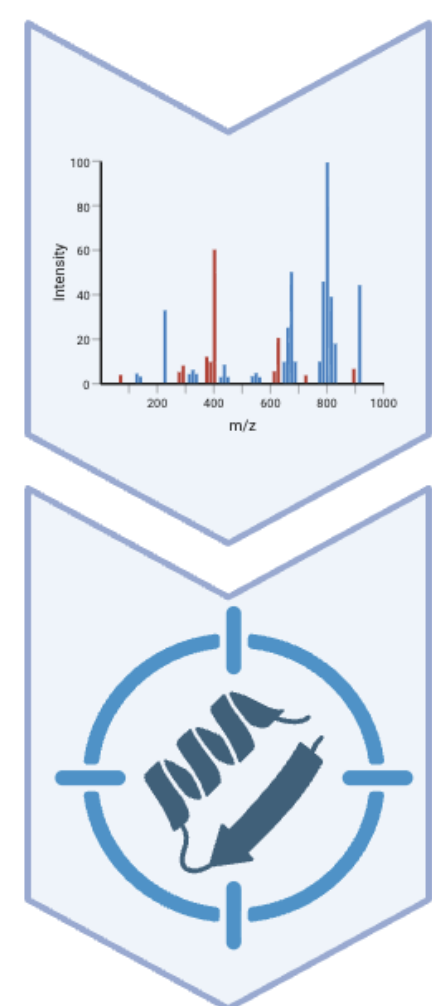
Discovery of Novel Ion Channel Modulators from *Physalia physalis*Zuzanna Tomkielska ^{1,2}, Jorge Frias ¹, Ana Casas ², Nelson Simões ¹, and Duarte Toubarro* ¹¹ Center of Biotechnology of Azores (CBA), University of Azores, 9500-321 Ponta Delgada, Portugal² Mesosystem Investigação & Investimentos by Spinpark, Barco, 4805-017 Guimarães, Portugal

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INTRODUCTION & AIM

Animal venoms are increasingly recognized as rich sources of bioactive molecules with therapeutic potential, particularly peptides targeting voltage-gated ion channels, which play critical roles in physiological processes and are implicated in a wide range of diseases. Among these, ShK-like peptides, originally discovered in sea anemone venom, have emerged as potent and selective modulators of Kv channels, highlighting the potential of venom-derived molecules in drug development. In this study, we report the discovery of 30 ShK-like domains in the venom of *Physalia physalis*, identified through proteomic analysis and predicted to selectively modulate specific Kv channel subtypes. **The aim of this work was to identify, characterize, and functionally validate one of these novel peptides, with a particular focus on their inhibitory activity against the Kv1.3 channel, a key therapeutic target involved in autoimmune disorders, pain, and neurological diseases.**

METHOD



Proteomic analysis

Venom was analyzed by LC-MS/MS (CIEX TripleTOF 6600), and spectra were searched against NCBI (Cnidaria-restricted) and a custom transcriptome-derived database (NCBI Accession SUB13325220).

Structure prediction and docking

3D structures of the ShK-like peptides were predicted using AlphaFold2 and subsequently docked against a panel of human Kv1 channels with ZDOCK.

Selection of the peptide based on the docking results

ShK26_D5

Vector construction

One of the most promising ShK variant was expressed in *E. coli* using the pET-40 vector, targeted to the periplasmic space in fusion with DsbC to promote correct disulfide bond formation, and carrying a His-tag.

Heterologous expression

Protein production was carried out in *E. coli* C41 using AI medium ZYM-5052 (2h at 37°C followed by 6h at 30°C, yield approx. 1 g/L pellet per cell culture) and purified by FPLC using a HisTrap and Desalting column.

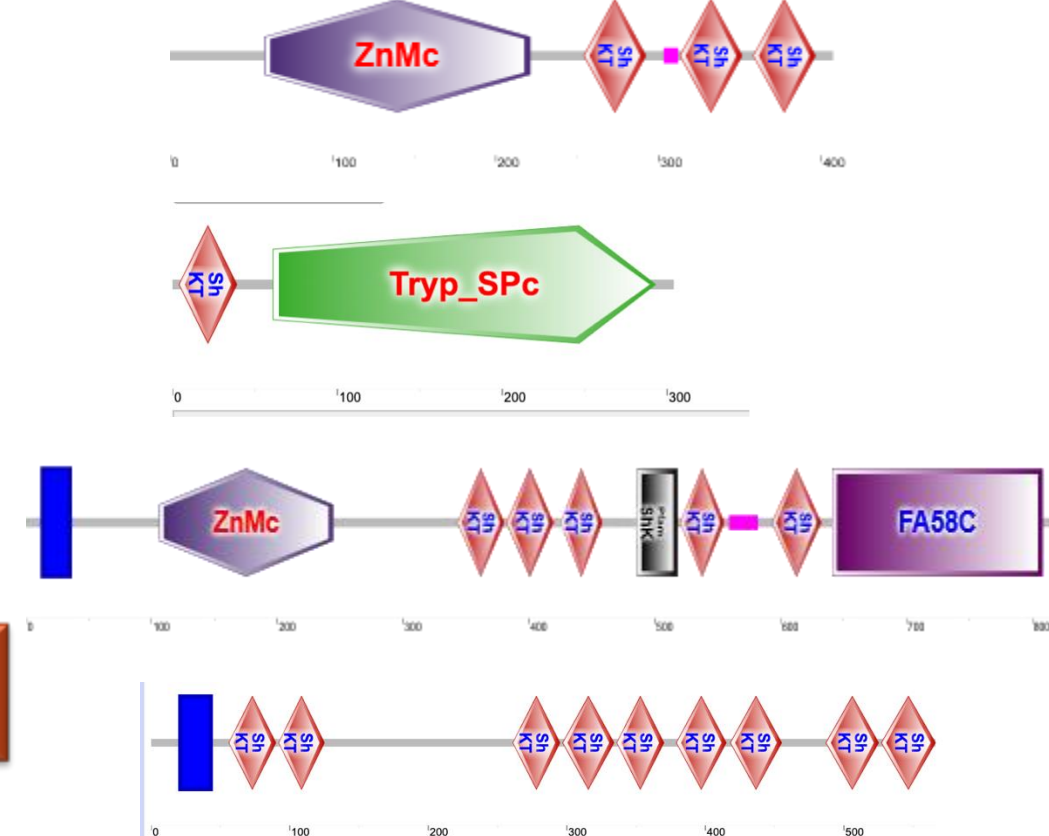
Target validation

Functional characterization was performed using automated whole-cell patch-clamp electrophysiology (SynchroPatch 384i) on fibroblasts heterologously transfected with Kv1.3, and Quinidine as the reference compound. A peak pulse protocol, from a holding potential of -80 mV to +60 mV over 300 ms, was applied every 20 seconds to monitor the outward current.

Radioligand competition assay

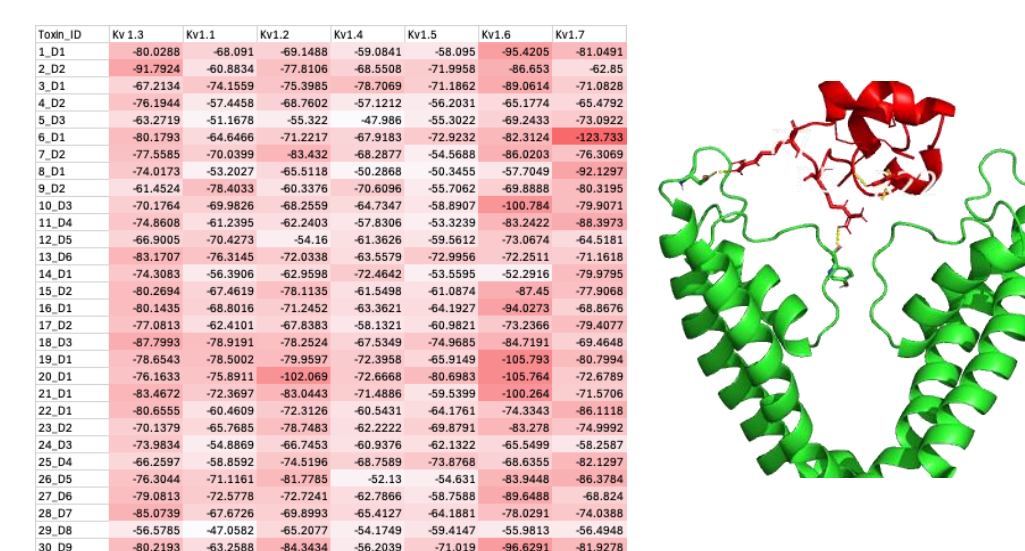
Evaluation of the affinity of the compound (10 μ M) for the Kv channel in the human cerebral cortex was determined in a competition assay with radiolabeled α -[125I]dendrotoxin (20nM and 50nM for non-specific binding).

RESULTS & DISCUSSION

1. Proteomic analysis of *P. physalis* venom revealed the presence of multiple ShK-like domains.

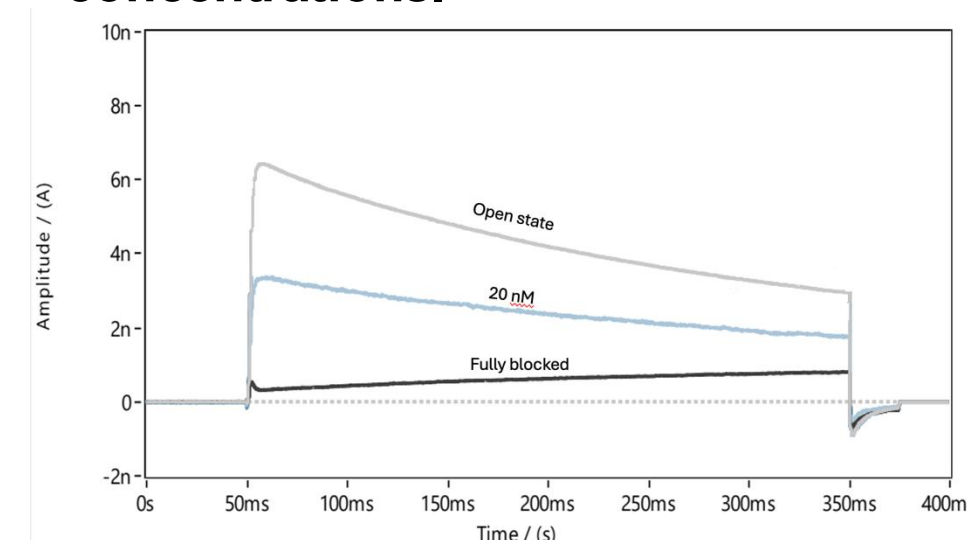
Multiple ShK-like peptides tandemly arranged in a single coding sequence.

3. Docking against the panel of human Kv1 channels reveals peptides with Kv1.3 preference.



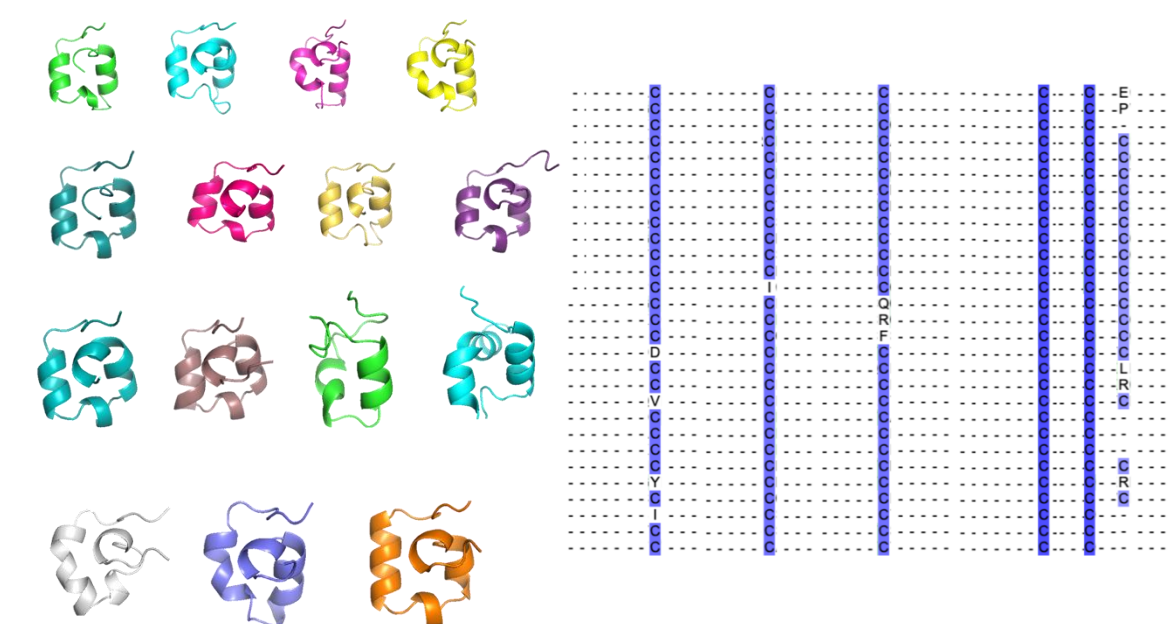
Peptide ShK26_D5 was selected for recombinant production based on its **low binding energy** and **direct binding to the pore** of the Kv1.3. **Green** structure represents the pore of the Kv1.3 channel while **red** represents ShK26_D5 peptide.

5. Whole-cell patch-clamp confirmed activity of ShK26_D5 at nanomolar concentrations.



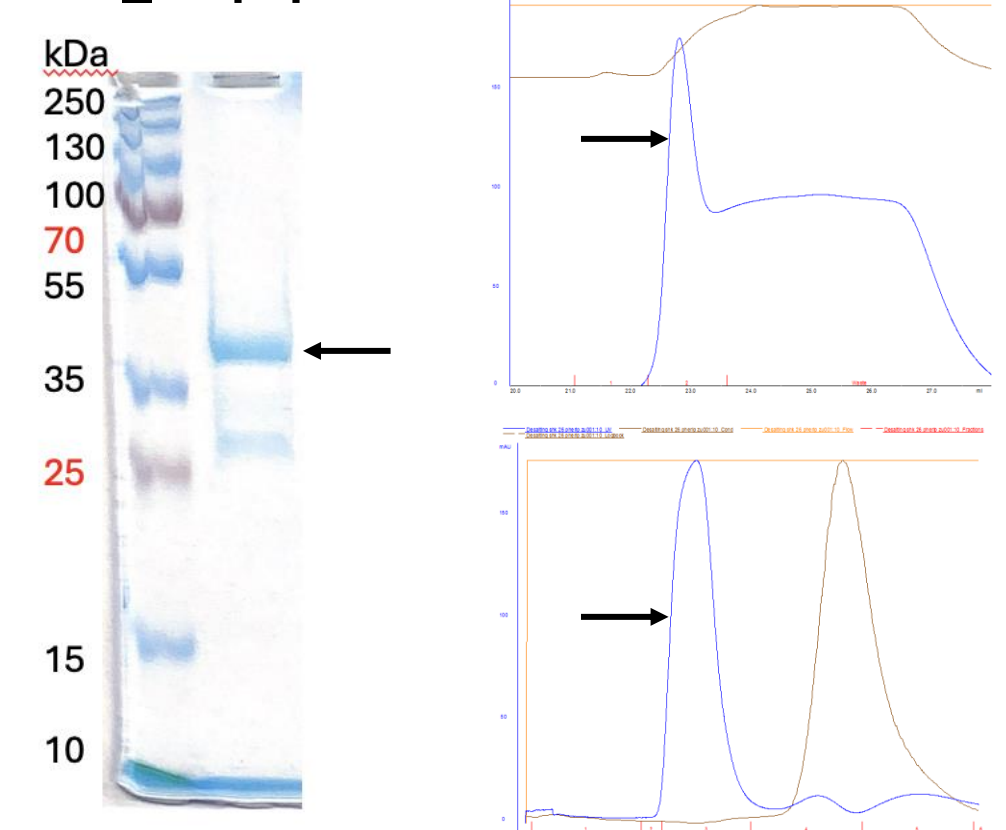
Patch-clamp recordings demonstrate that **20 nM** of the peptide effectively **inhibits Kv1.3 currents**, confirming its strong blocking activity. Open state (grey), inhibition with peptide (blue), and fully blocked channel (black).

2. AI-based platform predicts the 3D structure of the peptides with highly conserved Cys residues.

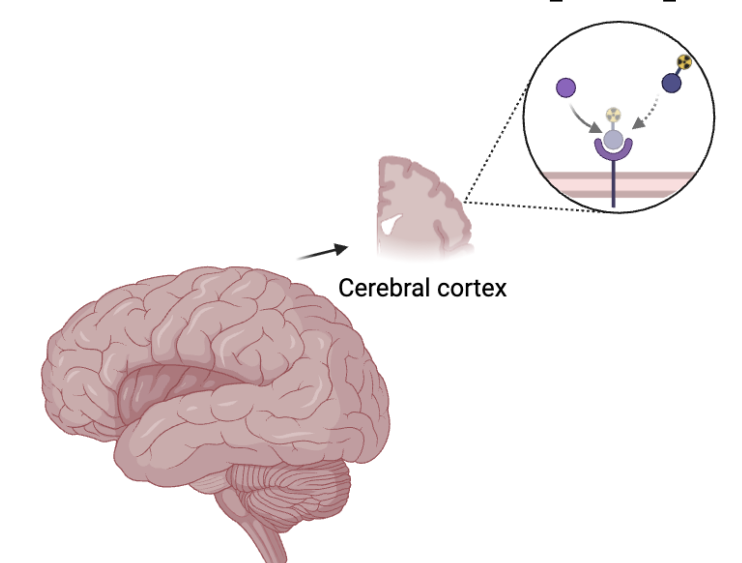


- Highly conserved 2-3 disulfide bridges that stabilize the structure
- 33-47 residues

4. Expression and purification of the ShK26_D5 peptide.



ShK26_D5 was expressed in a **soluble form**. Upper chromatogram: His-tag purification; lower chromatogram: desalting.

6. Competition assay on a cerebral cortex tissue with radiolabeled α -[125I]dendrotoxin

The recombinant ShK26_D5 peptide showed **low competition (4%)** against Kv1.1, Kv1.2, and Kv1.6, even at **200x higher concentrations** than the reference compound, which minimizes the risk of neurological side effects.

CONCLUSION

- Physalia physalis* venom contains 30 ShK-like peptides with potential as selective Kv channel modulators.
- The recombinant ShK26_D5 peptide showed **blocking activity on the Kv1.3 channel**, with minimal competition against Kv1.1, Kv1.2, and Kv1.6, reducing the likelihood of off-target interactions associated with severe side effects, placing it as a **promising candidate for therapeutic applications in Kv1.3-mediated disorders**.

FUTURE WORK / REFERENCES

- Future studies should address functional validation in neuronal and immune cells and in a vertebrate model to evaluate therapeutic potential and safety.

ACKNOWLEDGEMENT

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