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Oxidative stress and apoptosis are strongly involved in ricin-induced intestinal cell intoxication

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INTRODUCTION & AIM

Ricin, a type 2 ribosome-inactivating protein (RIP), is a lethal toxin purified from castor bean seeds, and it has been classified as a chemical and biological weapon according to Schedule 1 by the OPCW [1]. Ricin cytotoxicity derives from its lectin chain, which facilitates cell entry, and from its active chain, which can depurinate different substrates, thus leading to multiple cell death pathways [2]. Oral ingestion is highly toxic, with symptoms appearing within 24 hours and a human lethal dose estimated at 1-20 mg/kg [3]. While the systemic effects of ricin are well-known, its localized impact on the gastrointestinal tract remains a critical concern. This study aims to elucidate ricin intoxication in two human intestinal cell lines, named HT29 and Caco-2, and to determine ricin impact on the integrity of the epithelial barrier.

RESULTS & DISCUSSION

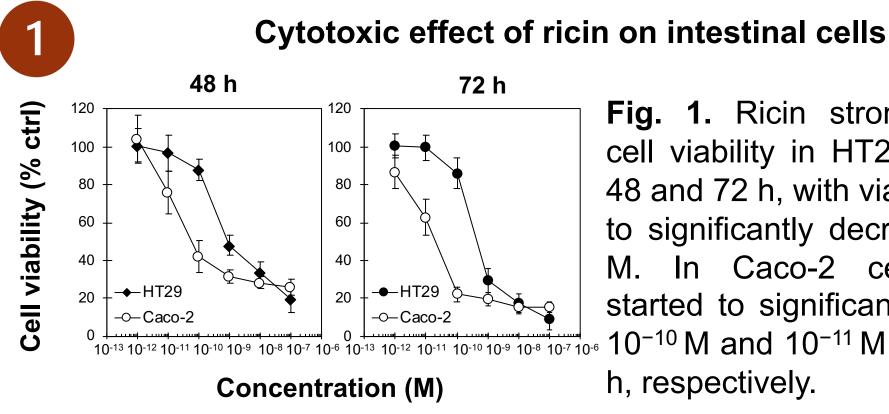


Fig. 1. Ricin strongly affected cell viability in HT29 cells, after 48 and 72 h, with viability starting to significantly decrease at 10⁻⁹ M. In Caco-2 cells, viability started to significantly reduce at 10⁻¹⁰ M and 10⁻¹¹ M at 48 and 72 h, respectively.

Effects of ricin on barrier integrity in Caco-2 monoculture

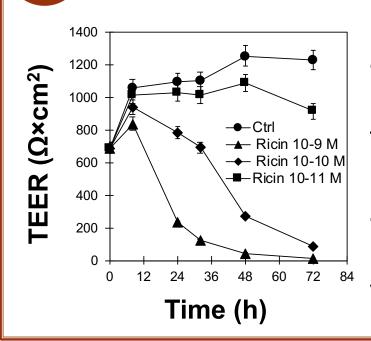


Fig. 2. At 10⁻⁹ M concentration, ricin strongly affected the barrier integrity after 24 h in Caco-2 monolayer, showing TEER values 5-fold lower than those of controls. At 10⁻¹⁰ M concentration, ricin was also able to reduce TEER values by about 1.3-fold with respect to control cells. At 10⁻¹¹ M concentration, ricin has no effect, and TEER values are comparable to controls.

Evaluation of cell death induced by ricin in HT29 and Caco-2 cells

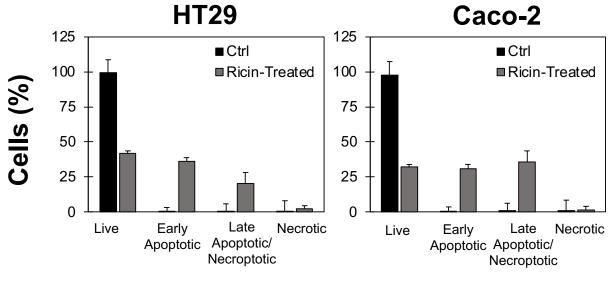


Fig. 3. At 10⁻⁹ M concentration, ricin was able to induce apoptosis/necroptosis after 48 h, without the involvement of necrosis.

Effects of ROS scavengers on ricin cytotoxicity in HT29 and Caco-2 cells

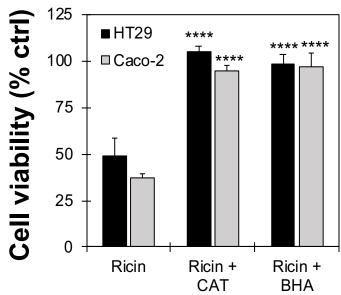


Fig. 4. Ricin cytotoxicity (at 10⁻⁹ M concentration) can be completely prevented after 48 h preincubating cells with antioxidant scavengers, demonstrating the involvement of oxidative stress in cell death mechanisms after ricin exposure.

METHODS

Ricin cytotoxicity was evaluated on two human intestinal cell lines, HT29 and Caco-2, in dose and time-response experiments, determining cell viability via an MTS-reduction assay. Cells (3 × 10³/well) were treated with scalar concentrations of ricin for 48 and 72 h. The effect of ricin on Caco-2 cell monolayer integrity was monitored by Trans-Epithelial-Electrical Resistance (TEER) measurements at 0, 8, 24, 32, 48 and 72 h after ricin intoxication. Cell death was determined through flow cytometry analysis of Annexin V/PI positivity and the involvement of oxidative stress in ricintreated cells was indirectly investigated after 48 h, pretreating cells with two reactive oxygen species scavengers, catalase (CAT) and butylated hydroxyanisole (BHA).

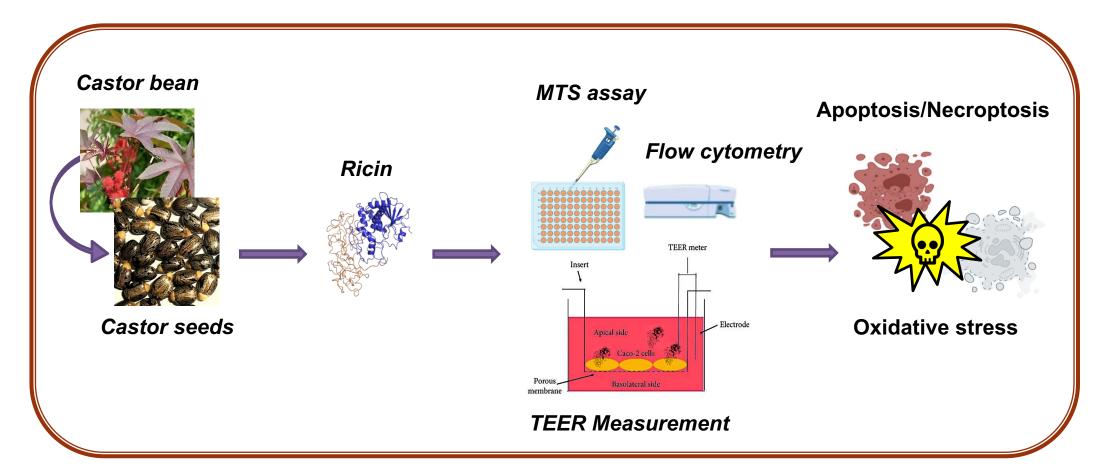


Fig. 1 Experimental Workflow

CONCLUSIONS

Our results demonstrate a dose- and time-dependent viability reduction in both HT29 and Caco-2 cells after ricin exposure.

TEER experiments demonstrate a dose- and time-dependent reduction in barrier integrity after ricin intoxication in Caco-2 cell monolayer.

Experiments on cell death mechanisms revealed a predominant role of apoptosis/necroptosis in ricin-intoxicated HT29 and Caco-2 cells after 48 h, without necrosis involvement.

Interestingly, both antioxidants used, CAT and BHA, can significantly protect HT29 and Caco-2 after 48 h of ricin exposure.

These results pave the way for new promising therapeutic strategies against oral ricin poisoning.

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