

Synthesis of novel modified uracil for dual-purpose: Quenching and photoswitching

Mohamed E. Moustafa^{a,b} and Robert H. E. Hudson^a

^aDepartment of Chemistry, The University of Western Ontario, London, Ontario, Canada, N6A 5B7

^bDepartment of Chemistry, Faculty of Education, Suez Canal University, Arish, Egypt

Novel azo-based uracil derivatives have been synthesized with a photoswitching and quenching properties. The advantage of these derivatives over the traditional azobenzene photochromophore is maintaining the hydrogen bonding networks with the complementary nucleosides. *Trans-to-cis* Photoisomerization is observed up on illumination at 365nm, this process can be reversed by illumination thermally.

Keywords: Quencher, photoisomerization, dimethylphenylazo, uracil

Introduction

Hybridization of nucleic acids is considered the cornerstone for many biomedical applications as antigene and antisense. Recently, much effort has been devoted to regulate nucleic acid hybridization using external stimuli as light, pH,¹ electric field² and heat.³ Light is preferred over all other external stimuli due to many reasons such as efficiency, clean and does not produce any contaminants to the biological system under study.

Photoregulation of nucleic acids can be achieved via attaching a photoresponsive molecule to the single-stranded DNA. Azobenzene one of photochromic molecules that extensively has been widely used for photoregulation of nucleic acid through geometrical structural change from *trans* to *cis* isomer and reverse with response of optical excitation with light at suitable wavelength.⁴⁻⁹ Irradiation with UV light led to *trans-to-cis* isomerization while irradiation with visible light or thermally recovers the *trans* isomer. Incorporation of azobenzene to oligonucleotides was obtained via two approaches (i) as a side chain on the oligonucleotide phosphate backbone¹⁰ or (ii) as a linker between two oligonucleotide segments in the main chain of an oligonucleotide.¹¹ In principle,

introduction of several azobenzene residues to oligonucleotides led to an increase in melting temperature difference ΔT_m corresponding to the photo-induced isomerization. Despite the efficiency of azobenzenes in reversible photoswitching of nucleic acid-related activities, a desire to overcome the non-nucleosidic nature of the attached azobenzene moiety by designing azo-nucleobase analogue that able to maintain the H-binding ability and thusly enhance the stabilization of duplex formation.

4-(dimethylamino)azobenzene-4'-carboxylic acid (DABCYL), universal quencher, one of azobenzene derivatives that widely used as a quencher for nucleic acid detection using fluorescent probe technique. Here we report the synthesis of novel azo-based uracil derivatives with potential dual-purpose as photo trigger and/or as a quencher that mimics the structure of the universal quencher DABCYL. The modified uracil is designed to have 4-(dimethylamino)phenylazo moiety at C5 position due to at this position there is no interference with the hydrogen bonding sites of uracil and therefore retain the base pairing ability with adenine. During the progress of this work, two examples of azobenzene-modified 2'-deoxyuridine at C5 position have been reported.¹²

Results and discussions

We are interested in the synthesis of 4-(dimethylamino)phenylazo derivative as azobenzene analogue for several reasons: 1) 4-(dimethylamino)phenylazo moiety is able to induce a conformational changes by photoisomerization. 2) The presence of dimethylamino group which is susceptible to protonation facilitates the investigation of pH effect. 3) The structure of azo-based nucleoside is structurally mimics the universal quencher. 4-(dimethylamino)azobenzene-4'-carboxylic acid (DABCYL), therefore, may have potential quenching effect as shown in figure 1.

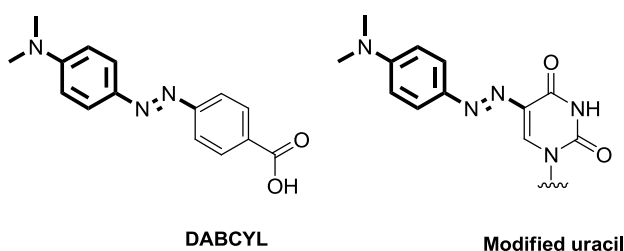
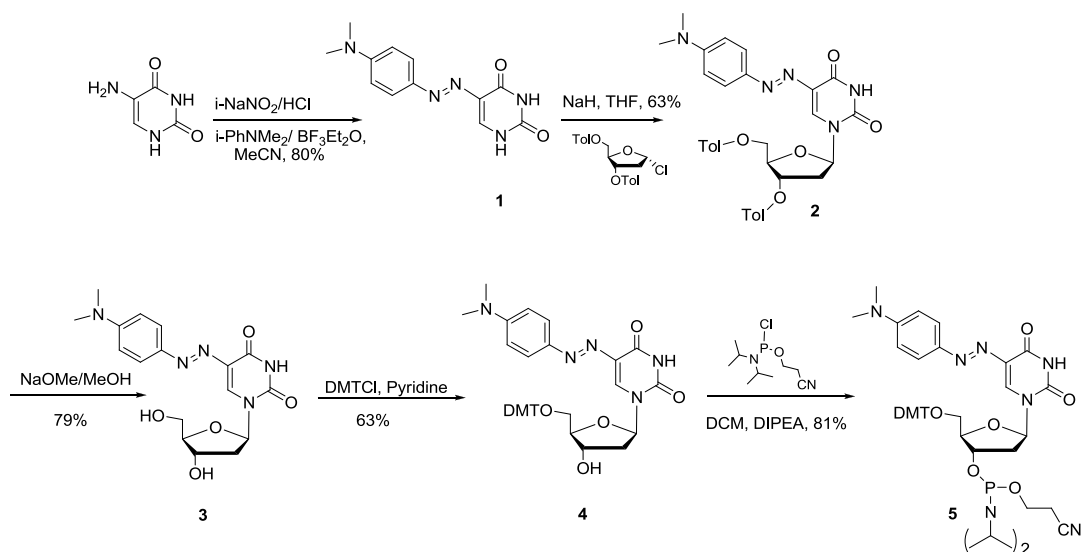


Figure 1: Comparison between dapcyl and the modified azo-based uracil

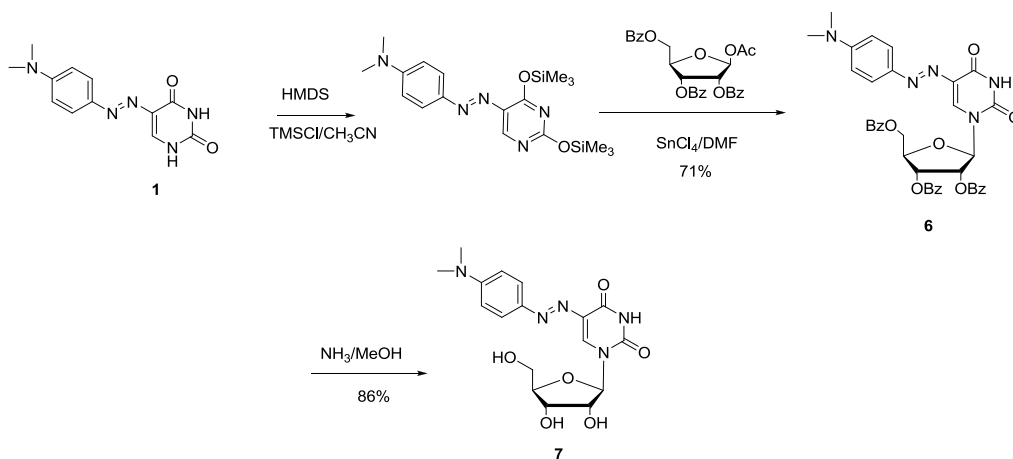
The synthesis of azo-based uracil **1**, was obtained by treatment of 5-aminouacil with NaNO_2 in the presence of HCl to afford the corresponding diazonium salt followed by coupling with dimethylaniline to give the desired product **1** in 80% yield. The synthetic approach to synthesize the phosphoramidite **5** for DNA synthesis purpose is depicted in scheme 1. Azo-modified uracil **1** then reacted with 1'- α -chloro-3,5-di-*O*-*p*-toluoyl-1,2-dideoxy- α -D-ribofuranose under basic condition to give the corresponding deoxyriboside **2** followed by removal of the toluoyl protecting groups using sodium methoxide in methanol giving the azo-based 2-deoxyriboside **3** in 79% yield. Standard methods were used to convert the unprotected nucleosides to 5'-dimethoxytrityl-protected derivative by using dimethoxytrityl chloride (DMTCl) in pyridine giving **4** in 63% yield followed by conversion to the corresponding cyanoethyl phosphoramidite derivative **5** in 81% yield.



Scheme 1. Schematic synthesis of deoxyuridine phosphoramidite possesses phenylazo moiety for photoswitching.

To get an access for RNA chemistry, ribonucleoside **7** was synthesized following Vorbrüggen procedure¹³ starting with refluxing azo-modified uracil **1** with hexamethyldisilazane (HMDS) and trimethylsilyl chloride (TMSCl) to form the corresponding silylated nucleoside followed by coupling with 1-*O*-acetyl-2,3,5-tribenzoyl ribofuranose in dimethylformamide (DMF) in the presence of SnCl_4 to afford **6** in 71%.

Removal of benzoyl groups was carried out using ammonia in methanol to afford azo-based riboside **7** in 86 % yield.



Scheme 2. Schematic synthesis of photochromic riboside.

Photoisomerization studies

Azobenzene is well known to isomerize from its mostly thermally stable planar *trans*-isomer to *cis*-isomer upon exposure to UV-light irradiation (300nm ~ 400nm), and the reverse process takes place corresponding to irradiation with visible light (> 400nm) or thermally (Fig. 2). This process is completely reversible under UV and visible irradiations. From this vein, the synthesized nucleosides carrying azophenyl moiety is expected to follow the same behavior that azobenzene follows.

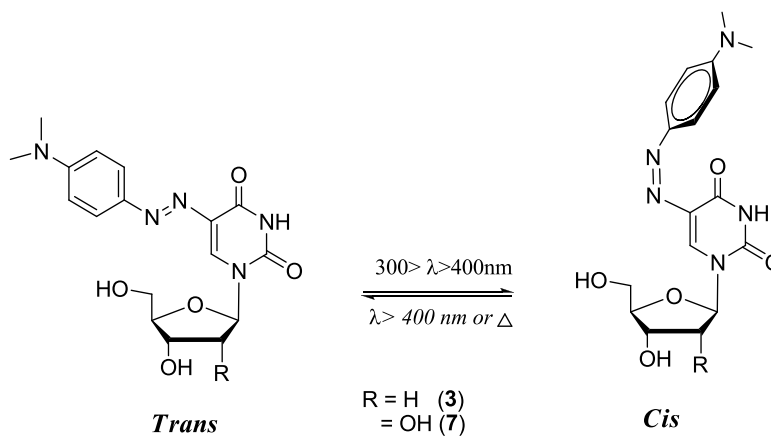


Figure 2. Schematic representation of reversible *trans*-*cis* photoisomerization of the photochromic nucleoside **3** and **7**.

To assess the photochromic behavior of the synthesized nucleosides **3** and **7** by monitoring the changes that may occur at their absorption with response of photoirradiation. As shown in figure 3, the absorption spectrum of *trans* isomer has a major peak centered at 454 nm (in dichloromethane for **3**) and at 443 nm (in acetonitrile for **7**) attributed to strong π - π^* transition which overlapped with the weak peak attributed to weak n - π^* transition. Upon irradiating with monochromic light at 365 nm, a decrease in the intensity of the absorption band was observed along with the irradiation time due to *trans*-to-*cis* isomerization, while the reverse *cis*-to-*trans* isomerization was achieved via thermal relaxation.

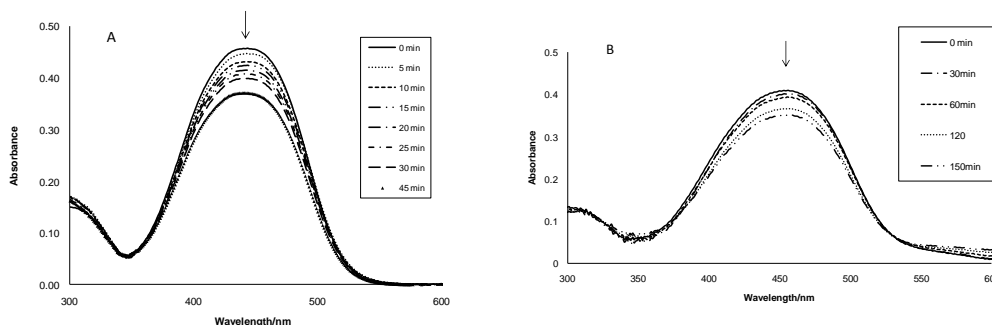


Figure 3. Absorption spectra of a) **3** in dichloromethane and b) for **7** in acetonitrile at 10 μ M concentrations for *trans*-*cis* photoisomerization upon illumination with UV-Light at 366nm.

To assess the quenching ability of the synthesized azo-based uracil **1**, a preliminary, qualitative study to quench the emission signals of fluorescein and pyrene luminophores has been made. As shown from the absorption spectra of **1** in ethanol, modified uracil **1** has absorption range 350-525 nm with maximum wavelength 436 nm, which is expectedly overlap with emission spectra of fluorophores used in the study. A solution of fluorophores in ethanol was titrated with aliquots of modified uracil **1**, as illustrated in

(Fig. 4), upon addition of small portions of **1** to the fluorophore, a subsequent quenching of fluorescent signal was observed.

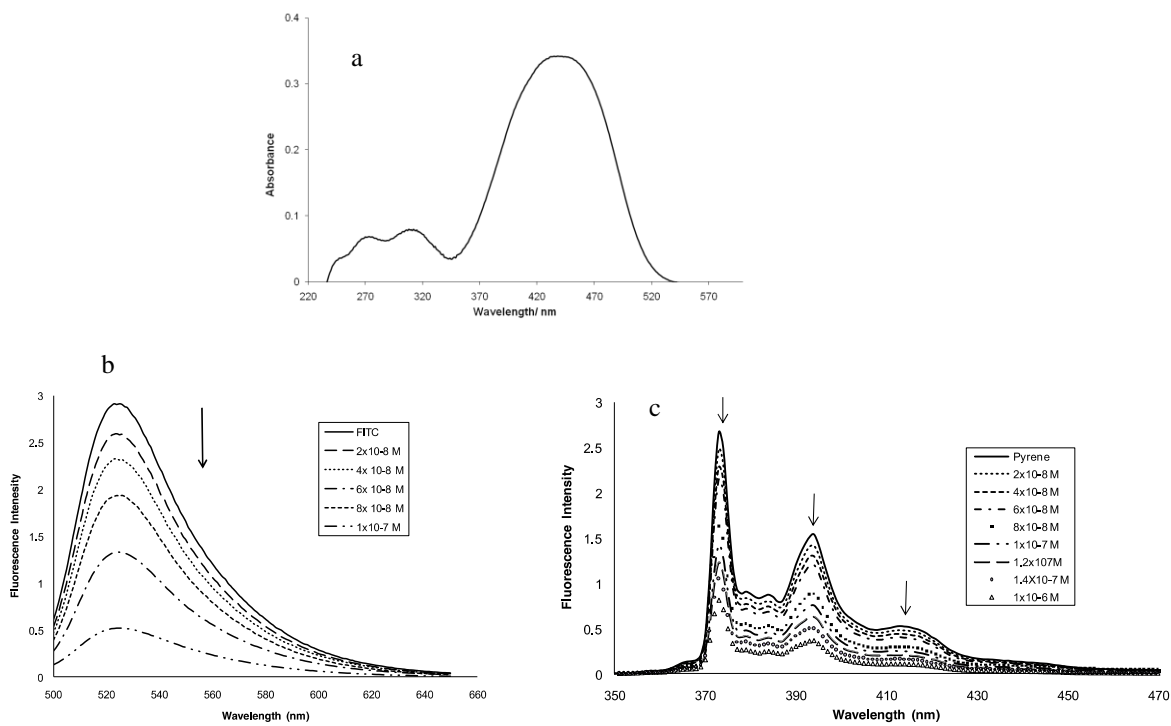


Figure 4:- a) Absorption spectrum of compound **1**, quenching of b) fluorescein (with excitation wavelength 488 nm) and c) pyrene (with excitation wavelength 330 nm) emission signals upon addition of various amount of azo-uracil **1** in ethanol.

pH-Sensitivity

Azobenzenes especially aminoazobenzene and dimethylaminoazobenzene attracted much attention due to their ability to change colour in solution with change of the pH. It is well established that protonation of aminoazobenzene derivatives led to the formation of two tautomeric azonium and ammonium forms.¹⁴ As illustrated in figure 5, Azonium ion is obtained by protonation on azo-nitrogen leading to delocalized of the lone-pair electrons of the aromatic ring and as consequence a red shift in absorption maximum was occurred. Ammonium ion is obtained by protonation at amino-nitrogen resulting in prevention the conjugation of electrons lone-pairs and thusly the UV-Vis spectrum looks similar to that of unsubstituted azobenzene due to diminish the effect of amino substituent.

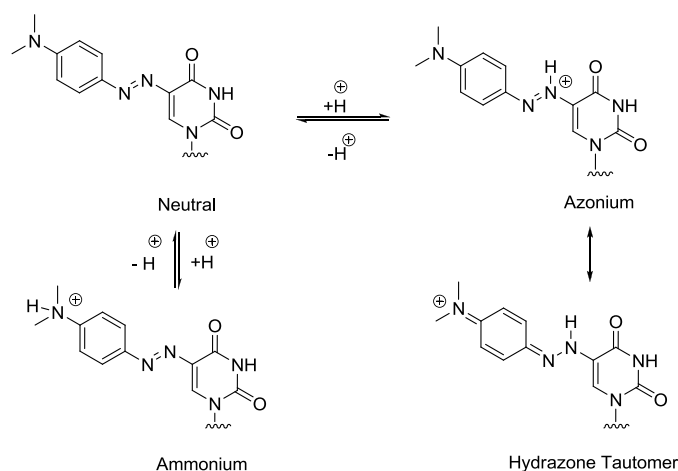


Figure 5. Molecular structure of dimethylaminoazophenyl(deoxy)uridine and its protonated forms. Sugar unit was removed for simplicity

The presence of an azo- and dimethylamino groups in the synthesized nucleosides promoting us to study the effect of acid. We investigated the effect of trifluoroacetic acid solution (0.005 M) in dichloromethane and ethanol on the nucleosides **3** and **7** respectively. Upon protonation, the band at λ_{\max} 411 nm, which is attributed to neutral *trans*-dimethylaminoazobenzene, has been red shifted to λ_{\max} 535 nm attributed to the azonium ion with appearance of new absorption band at λ_{\max} 318 nm attributed to ammonium ion of *trans*-dimethylaminoazobenzene.

Similarly, as can be seen in figure **6**, upon addition of TFA solution to deoxyriboside **3** in dichloromethane leads to a decrease in the π - π absorption band at λ_{\max} 455 which is attributed to neutral form with an increase in π - π absorption band centered at 618 nm and 350 nm attributed to azonium and ammonium ions respectively. There are two isosbestic points were observed at 376, 507 nm indicate that compound **3** exists either as non-protonated or protonated species in tautomeric equilibrium. While, upon titrating of **7** with small aliquots of TFA in ethanol, the π - π absorption band for neutral form at λ_{\max} 436 nm was decreased with appearance of new absorption bands centered at 623nm and 342 nm attributed to the protonated forms azonium and ammonium ion respectively. These results are in excellent agreement with the aforementioned study. Surprisingly, subjecting solutions of **3**, **7** in halogenated solvents as dichloromethane or chloroform to

TFA leads to color change from yellow to deep blue color while using solvent ethanol, methanol, ethyl acetate instead of halogenated solvents this color change was not observed.

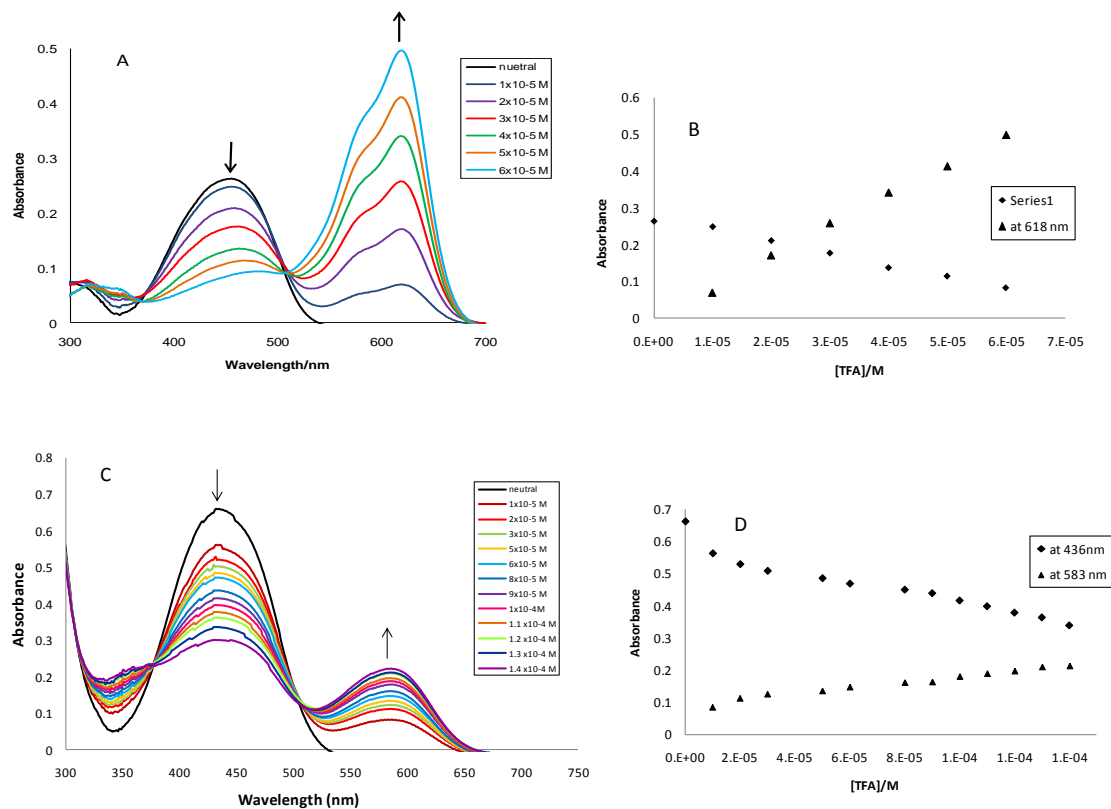


Figure 6. Changes in UV-Vis spectra of A) 3 at 6.2 μM concentration and C) 7 at 13.3 μM concentration upon addition of increasing concentrations of TFA, arrows indicate the progressive decrease of absorption band for neutral form and the growth of protonated form. B, D) Variation of absorbance vs TFA concentration for the respective absorption bands.

Conclusion

We have successfully designed and synthesized novel photochromic azo-based nucleosides with potential photoswitching and quenching properties for DNA and RND chemistry with potential duplex stabilization by retaining the ability of formation of hydrogen bond between the modified nucleobase and the complementary nucleobase,. These nucleosides showed ability to undergo *trans*-to *cis* isomerization upon irradiation with monochromatic

light at 365nm and reversibly the *trans* configuration was recovered thermally in the dark. In addition, it showed halochromic effect upon acid treatment. Therefore, the synthesized photochromic nucleosides may be contribute in photoregulation of nucleic acids.

Materials and Methods

General consideration. All solvents and chemicals were commercially available and were used as purchased. Reactions were monitored by thin layer chromatography (TLC) on pre-coated 0.2 mm Merck Kieselgel 60 TLC plates. Silica gel 60 F254 (Merck) plates and visualized with an ultraviolet light source at 254 nm. Silica gel (Merck Kieselgel 60, 15-40 mm) was used for column chromatography. ¹H NMR, ¹³C NMR and ³¹P NMR spectra were recorded at a Varian Mercury Plus spectrometer (400.09 MHz, 100.61MHz and 242.9 MHz respectively) at room temperature. Chemical shifts are reported in parts per million (δ), were measured from tetramethylsilane (0 ppm) and are referenced to the residual proton in the deuterated solvent: CDCl₃ (7.26 ppm), DMSO-*d*₆ (2.48 ppm), CD₃OD (4.87 and 3.31 ppm) for ¹H NMR and CDCl₃ (77.0 ppm), DMSO-*d*₆ (39.5 ppm) and CD₃OD (49.1) for ¹³C NMR. Multiplicities are described as s (singlet), d (doublet), t (triplet), m (multiplet), and br s (broad singlet). Coupling constants (*J*) are reported in Hertz (Hz). High resolution mass spectra (HRMS) were obtained using electrospray ionization time of flight methods (ESI-TOF).

Synthesis

5-((4-(dimethylamino)phenyl)diazenyl)uracil **1**

To a cold solution of 5-aminouracil (1.0 g, 7.87 mmol) in hydrochloric acid 1N (15 mL), a solution of 6.9% NaNO₂ (6 mL) was added dropwise over a period of 15 min. a buff solid was precipitated. The reaction mixture was stirred for further 30 min for complete precipitation. Solid was collected by filtration, wash with iced water and dry under vacuum to give 5-diazouracil (1.04 g, 84%) and used for the next step without further purification. to a suspension of 5-diazouracil (0.5 g, 3.2 mmol) in anhydrous acetonitrile (50 mL), borontrifluoride etherate (0.41 mL, 3.2 mmol) and dimethylaniline (0.38 mL, 3.2 mmol) were added to the mixture and was stirred for 2 h. The separated solid was

filtered and refluxed in EtOH (100 mL) for 1 h. The solid was collected and dried to afford (1.38 g, 80%) of **1**. ¹H NMR (DMSO-d₆) δ : 11.42 (s, 1H), 11.28 (s, 1H), 7.66 (s, 1H), 7.61 (d, *J* = 8.9 Hz, 2H), 6.79 (d, *J* = 8.7 Hz, 2H), 3.01 (s, 6H); ¹³C NMR (DMSO-d₆) δ : 161.8, 152.7, 151.3, 143.5, 130.7, 129.5, 124.9, 112.2, 47.6. HRMS (ESI): calcd. for C₁₂H₁₃N₅O₂ [M]⁺ 259.1069 found 259.1062.

5-((4-(dimethylamino)phenyl)diazanyl)2'-deoxyuridine-3',5'-di-(p-toluoyl) ester **2**

To a suspension of 5-((4-(dimethylamino)phenyl)diazanyl)uracil **1** (0.44 g, 1.72 mmol) in dry THF (20 mL), NaH (60% dispersion in mineral oil, 82 mg, 3.44 mmol) was added and stirred at r.t. for 2 h. A solution of 1- α -chloro-3',5'-di-O-toluoyl-2'-deoxyribose (0.69 g, 2.23 mmol) in dry THF (5 mL) was added to the mixture in one portion and the reaction mixture was stirred for 24 h under inert atmosphere. Solvent was evaporated under vacuum and the residue was dissolved in dichloromethane (30 mL), was washed with saturated aqueous sodium Na₂CO₃, brine and dried over anhydrous Na₂SO₄. The solution was filtered, concentrated, and the residue was subjected to purification by flash chromatography with CH₂Cl₂/CH₃OH (20:1, v/v) as eluent to give bis-toluoyl ester **2** as red solid. The product was obtained as a mixture of two isomers (0.65 g, 63%). ¹H NMR (DMSO-d₆) δ: 11.78 (br s, 1H), 8.02 (s, 1H), 7.92-7.89 (m, 2H), 7.80 (d, *J* = 7.8 Hz, 1H), 7.68 (d, *J* = 8.2 Hz, 1H), 7.50 (d, *J* = 8.9 Hz, 1H), 7.37-7.33 (m, 3H), 7.16 (d, *J* = 8.2 Hz, 1H), 6.91 (d, *J* = 8.2 Hz, 1H), 6.75 (d, *J* = 8.9 Hz, 1H), 6.69 (d, *J* = 9.3 Hz, 1H), 6.32 (m, 1H), 5.57 (d, *J* = 5.8 Hz, 1H), 5.01 (m, 1H), 4.57 (m, 1H), 4.41 (m, 1H), 3.02 (s, 6H), 2.97 (m, 1H), 2.61 (m, 1H), 2.37 (s, 3H), 2.26 (s, 3H); ¹³C NMR (CDCl₃) δ : 166.1, 165.7, 152.2, 144.3, 143.5, 129.8, 129.6, 129.1, 126.5, 126.6, 126.3, 125.7, 111.2, 87.9, 77.3, 74.6, 64.1, 40.1, 27.96. HRMS (ESI): calcd. 611.2380 for C₃₃H₃₃N₅O₇ [M]⁺ found 611.2391.

5-((4-(dimethylamino)phenyl)diazanyl)2'-deoxyuridine **3**

To a cold solution of the toluoyl ester **2** (0.5 g, 0.82 mmol) in dry CH₂Cl₂ (5 mL), NaOMe solution (0.2 M in MeOH) (9.8 mL, 1.96 mmol) was added dropwise over 15 min. The reaction mixture was stirred at r.t. for 1 h. An aqueous solution of 5% NH₄Cl was added to the mixture till the pH was 8 and the mixture was extracted with

ethylacetate (3 x 30 mL). The combined organics were washed with brine, dried over anhydrous Na₂SO₄ and concentrated under reduced pressure to afford red oil residue. The residue was subjected to flash chromatography using hexane/ CH₂Cl₂ (70:30, v/v) as the eluent to yield the deoxyriboside **3** as red powder (0.25 g, 79%). ¹H NMR (CD₃OD) δ: 8.50 (s, 1H), 7.79 (d, *J* = 7.8 Hz, 1H), 6.77(d, *J* = 7.8 Hz, 1H), 6.33 (t, *J* = 6.5 Hz, 1H), 4.43 (m, 1H), 3.96 (m, 1H), 3.83 (m, 1H), 3.75 (m, 1H), 3.20 (dd, *J* = 3.3 and 7.3 Hz, 1H), 3.03 (s, 6H), 2.40 (m, 2H); ¹³CNMR (CD₃OD): δ 163.5, 154.3, 151.5, 131.2, 126.4, 112.7, 89.3, 87.4, 72.1, 62.7, 41.8, 40.5. HRMS (ESI): calcd. 375.1543 for C₁₇H₂₁N₅O₅ [M]⁺ found 375.1535.

5-((4-(dimethylamino)phenyl)diazenyl)2'-deoxy-5'-O-(4,4'-dimethoxytrityl)uridine **4**

The free nucleoside **3** (0.2 g, 0.54 mmol) was dried by coevaporation with dry pyridine (3 x 5 mL) and then was dissolved in dry pyridine (10 mL). A solution of 4,4'-Dimethoxytritylchloride (0.269 g, 0.69 mmol) in dry pyridine (5 mL) and DIPEA (7.0 mmol, 1 mL) was added and the mixture was stirred at r.t. for 3 h. The reaction mixture was quenched by the addition of methanol (2 mL), was diluted with CH₂Cl₂ (75 mL) and then was washed with saturated aqueous solution of NaHCO₃ (3 x 50 mL). The organics were combined, dried over anhydrous Na₂SO₄ and filtered. The solvent was removed under reduced pressure to give red oil. The crude product was purified by flash column chromatography using CH₂Cl₂/ MeOH/ Et₃N (85: 10: 5, v/v) as elution solvent afforded the tetritylated derivative **4** as red solid (0.28 g, 63%). ¹H NMR (CDCl₃) δ: 8.25 (s, 1H), 7.65 (d, *J* = 8.9 Hz, 2H), 7.38 (d, *J* = 7.4Hz, 2H), 7.27 (d, *J* = 7.4 Hz, 4H), 7.22 (d, *J* = 7.0 Hz, 2H), 7.14 (m, 2H), 6.78 (d, *J* = 8.5 Hz, 3H), 6.51 (d, *J* = 8.5 Hz, 2H), 6.18 (m, 1H), 4.57 (m, 1H), 4.39 (m, 1H), 3.69 (s, 6H), 3.20 (m, 1H), 3.25 (m, 1H), 2.90 (s, 6H), 2.80 (m, 1H) 2.47 (m, 1H); ¹³C NMR (CDCl₃) δ: 161.4, 158.3, 152.0, 149.7, 144.5, 143.3, 135.6, 135.39, 130.4, 129.8 , 127.9, 127.7, 126.6, 125.0, 113.0, 111.1, 89.1, 86.2, 71.9, 64.1, 55.0, 45.6, 40.0. HRMS (ESI): calcd. 678.2928 for [M+H]⁺ C₃₈H₄₀N₅O₇ found 678.2906.

5-((4-(dimethylamino)phenyl)diazenyl)2'-deoxy-3'-(2-cyanoethyl-diisopropylphosphoramidite)-5'-O-(4,4'-dimethoxytrityl)uridine **5**

To a solution of DMT-deoxyuridine **4** (0.16 g, 0.24 mmol) in dry CH₂Cl₂ (5 mL), DIPEA (340 μL, 2.4 mmol), 2-cyanoethyl-(*N,N'*-diisopropylamino) chlorophosphite (105 μL, 0.47 mmol) was added. The reaction mixture was stirred at r.t. under ambient atmosphere for 3 h until deemed complete by TLC. Solvent was removed and the resultant residue was dissolved in CH₂Cl₂ (25 mL), washed with aqueous solution of 5% NaHCO₃ (25 mL), brine (20 mL) and was dried over anhydrous Na₂SO₄. After removal of solvent the crude product was subjected to flash chromatography using hexane/ ethylacetate/ Et₃N (80:15: 5 to 50:45:5, v/v) to yield the phosphoramidite derivative **5** (0.169 g, 81%). ¹H NMR (CDCl₃) δ: as two diastereomers: 8.19 and 8.17 (2s, 1H), 7.77 (d, *J* = 7.6 Hz, 2H), 7.38 (d, *J* = 7.6 Hz, 2H), 7.21 (m, 7H), 6.79 (d, *J* = 6.7Hz, 4H), 6.63 (d, *J* = 6.5 Hz, 2H), 6.29 (m, 1H), 4.80 (m, 1H), 4.54-4.46 (m, 2H), 3.75 (s, 6H), 3.57-3.48 (m, 2H), 3.03 (s, 6H), 3.38 (m,1H), 3.11 (m, 1H), 2.85 (m, 2H), 2.38-2.17 (m, 2H), 1.20 (s, 12H); ¹³C NMR (CDCl₃) δ : 160.8, 158.5, 152.3 149.5, 144.3, 143.5, 135.5, 129.9, 128.0, 126.9, 125.1, 113.2, 111.2, 88.7, 85.5, 74.1, 67.1, 55.0, 43.0, 40.2, 32.2, 29.6, 26.3, 24.4, 23.38; ³¹P NMR (CDCl₃): δ two diastereomeric peaks at 149.2 and 148.6. HRMS (ESI): calcd. 878.3880 for C₄₆H₅₅N₈O₈P, found 878.3889.

5-((4-(dimethylamino)phenyl)diazenyl)(2,3,5-Tri-O-benzoyl-b-D-ribofuranosyl)uridine **6**

A suspension of 5-((4-(dimethylamino)phenyl)diazenyl)uracil **1** (0.26 g, 1 mmol) in of 1,1,1,3,3,3-hexamethyldisilazane (HMDS) (20 mL) and of trimethylsilyl chloride (1 mL) was refluxed until a clear solution was obtained. The solution was allowed to cool, and excess HMDS was removed under reduced pressure. The silylated uracil derivatives were dissolved in CH₃CN (5 mL) and the solution was burged with nitrogen for 15 min prior to addition of SnCl₄ (10% excess) (127 μL, 1.2 mmol) to the reaction mixture with vigorous stirring. A solution of 1-*O*-Acetyl-2,3,5-tri-*O*-benzoyl-β-dribofuranose (0.56 g, 1.1 mmol) in CH₂Cl₂ (20 mL) was added dropwise to the reaction mixture. The solution was stirred at r.t. for 6 h at which the reaction came to completion as determined by TLC. The reaction mixture was quenched by aqueous saturated NaHCO₃ solution (100 ml) and was extracted with CH₂Cl₂ (3 x 50 ml) and was dried over anhydrous Na₂SO₄. The solvent was removed under reduced pressure leaving brown residue. The crude product was purified by flash column chromatography using elution with a gradient of hexane/

CH₂Cl₂ (from 100:0 to 60:40, v/v). Compound **6** was isolated as a red solid (0.5 g, 71%). ¹H NMR (CDCl₃) δ: 9.86 (br s, 1H), 8.05 (s, 1H), 7.96 (d, *J* = 7.4, 2H), 7.91 (d, *J* = 7.0, 2H), 7.70 (d, *J* = 7.4, 3H), 7.49 (m, 4H), 7.32 (m, 6H), 6.66 (d, *J* = 7.0, 2H), 6.29 (m, 1H), 4.82 (m, 1H), 4.72 (m, 3H), 3.06 (m, 1H), 3.03 (s, 6H); ¹³C NMR (CDCl₃) δ: 166.2, 165.3, 160.6, 152.7, 149.4, 143.4, 133.8, 133.4, 129.9, 129.8, 128.5, 125.5, 111.2, 88.6, 80.9, 74.2, 71.5, 64.2, 40.2. HRMS (ESI): calcd. for C₃₈ H₃₄ N₅O₉ [M+H]⁺ 704.2357 found 704.2358.

5-((4-(dimethylamino)phenyl)diazenyl)uridine **7**

Compound **6** (0.4 g, 0.57 mmol) was dissolved in saturated solution of ammonia in methanol (20 mL) and was stirred for 4 h at r.t. The solution was evaporated under pressure to give the crude product which was purified by flash column chromatography using CH₂Cl₂/MeOH (90: 10,v/v) yielding 0.19 g (86%). ¹H NMR (DMSO-d₆) δ: 7.73 (s, 1H), 7.64 (d, *J* = 8.9, 2H), 6.78 (d, *J* = 8.9, 2H), 6.14 (m, 1H), 5.07 (m, 1H), 4.90 (m, 1H), 4.62 (m, 1H) (Exchangeable 3OH), 4.52 (m, 1H), 4.12 (d, *J* = 5.4, 1H), 3.69 (m, 1H), 3.62 (m, 2H), 3.44 (m, 1H), 3.01 (s, 6H); ¹³C NMR (DMSO-d₆) δ: 160.8, 152.5, 143.2, 124.8, 111.9, 94.9, 84.9, 71.4, 70.6, 63.7, 62.79, 40.5. HRMS (ESI): calcd. for C₁₇H₂₁N₅NaO [M+Na]⁺ 414.1390 found 414.1382.

UV-Vis measurements:

All UV-Vis spectra were measured with a Varian Cary 300 Bio spectrophotometer using quartz cuvette cells of 1 mm of optical path.

Photoisomerization.

Trans-to-*cis* photoisomerization of nucleosides **3** and **7** were monitored by UV-Vis spectroscopy. A solution of the nucleosides at 1.06 x10⁻⁵ M concentration (**3** in dichloromethane) and (**7** in acetonitrile) were irradiated with UV-light at room temperature using a UVGL-58 hand held lamp (6 watt, 366 nm) at A distance of approximately 5 cm. The spectral change for absorption maximum peak at 455 nm (for **3**) and 436 nm (for **7**) were monitored with the time course of irradiation. While the *cis*-to-*trans* isomerization was achieved and confirmed by retrieving the maximum absorption

bands at 455 nm (for **3**) and 436 nm (for **7**) thermally by leaving the samples at the dark at room temperature.

Quenching analysis

Effect of acids

The spectral change of the synthesized photochromic nucleosides at concentration 6.2×10^{-6} M (in dichloromethane for **3**) and 1.5×10^{-5} M (in ethanol for **7**) were monitored upon addition of small aliquots of TFA solution (0.005 M).

Acknowledgements

The Authors are so grateful for the Natural Sciences and Engineering Research Council of Canada (NSERC) for financial support of this work. M. E. M. wishes to thank the Egyptian government for scholarship support.

References:

1. Bae Y., Fukushima S., Harada A., Kataoka K. Design of environment-sensitive supramolecular assemblies for intracellular drug delivery: polymeric micelles that are responsive to intracellular pH change. *Angew. Chem.* **2003**; 115:4788; *Angew. Chem. Int. Ed.* **2003**; 42: 4640-4643.
2. Osada Y., Okuzaki H., Hori H. Apolymeric gel with electrically driven motility. *Nature* **1992**; 355: 242-244.
3. Yamato M., Akiyama Y., Kobayashi J., Yang J., Kikuchi A., Okano T., Temperature-responsive cell culture surfaces for regenerative medicine with cell sheet engineering, *Prog. Polym. Sci.* **2007**; 32:1123-1133.
4. a) Yamana, K., Yoshikawa A., Nakano, H. Synthesis of a new photoisomerizable linker for connecting two oligonucleotide segments, *Tetrahedron Lett.*, **1996**, 37, 637-640. b) Yamana K., Yoshikawa A., Kan K., Nakano H. Synthesis of oligonucleotides containing a new azobenzene fragment with efficient photoisomerizability, *Bioorg. Med. Chem.* **1999**, 7, 2977-2983.
5. Asanuma H., Ito T., Yoshida, T., Liang, X., Komiyama M. Photoregulation of the formation of and dissociation of a DNA duplex by using the *cis-trans*

- isomerization of azobenzene, *Angew. Chem.* **1999**, 111, 2547-2549; *Angew. Chem. Int. Ed.*, **1999**, 38, 2393-2395.
6. Asanuma, H., Liang X., Yoshida T., Yamazawa A., Komiyama M. Photo-control of triplex formation by azobenzene-bearing oligo(thymidine), *Angew chem.*, **2000**, 112, 1372-1374. *Angew. Chem. Int. Ed.*, **2000**, 39, 1316-1318.
 7. Asanuma H., Tamaru D., Yamazawa A., Liu M., Komiyama, M. Photo-regulation of transcription reaction by T7 RNA polymerase by tethering an azobenzene in the promoter, *Chem. Bio. Chem.*, **2002**, 3, 786-789.
 8. Liu M.; Asanuma H., Komiyama M. Azobenzene-tethered T7 promoter for efficient photoregulation of transcription, *J. Am. Chem. Soc.*, **2006**, 128, 1009-1015.
 9. a) Shao Q, Xing B. Photoactive molecules for applications in molecular imaging and cell biology. *Chem. Soc. Rev.* **2010**; 39, 2835-2846, b) Ogasawara S, Maeda M. Straightforward and reversible photoregulation of hybridization by using a photochromic nucleoside, *Angew. Chem.* **2008**; 120, 8971-8974, *Angew. Chem. Int. Ed.* **2008**; 47, 8839-8843.
 10. Asanuma H, Yoshida T, Ito T, Koniyarna M. Photo-responsive oligonucleotides carrying azobenzene at the 2'-position of uridine. *Tetrahedron Lett.* **1999**; 40, 7995-7998. b) Asanuma H, Ito T, Yoshia T, Liang, X, Komiyarna M. Photoregulation of the formation and dissociation of a DNA duplex by using the cis-trans isomerization of azobenzene. *Angew. Chem. Int. Ed.* **1999**; 38, 2393-2395; c) Asanuma H, Ito T, Komiyama M. Photo-responsive oligonucleotides carrying azobenzene in the side-chains, *Tetrahedron Lett.* **1998**, 39, 9015-9018.
 11. a) Yamana K, Kan K, Nakano H. Synthesis of oligonucleotides containing a new azobenzene fragment with efficient photoisomerizability. *Bioorg. Med. Chem.* 1999; 7:2977-2983. b) Yamana K, Yoshikawa A., Noda R., Nakano H. Synthesis and binding properties of oligonucleotides containing an azobenzene linker. *Nucleosides, Nucleotides and Nucleic Acids.* **1998**; 17, 233-242.
 12. a) Mori S. , Morihiro K., Obika S., *Molecules*, **2014**, 19, 5109-5118; C5-Azobenzene-substituted 2'-Deoxyuridine-containing Oligodeoxynucleotides for Photo-Switching Hybridization. b) Kovaliov M., Wachtel C., Yavin E., Fischer

- B., Synthesis and evaluation of a photoresponsive quencher for fluorescent hybridization probes, *Org. Biomol. Chem.*, **2014**, 12, 7844-7858.
13. Balla U, Vorbrüggen H. Synthesis of nucleosides 17. A general synthesis of N-glycosides. 6. On the mechanism of the stannic chloride catalyzed silyl Hilbert-Johnson reaction. *J. Org. Chem.*, **1976**; 41:2084-2086.
14. a) Swasicki E. Physical properties of the aminoazobenzene dyes. iv. the position of proton addition, *J. Org. Chem.* **1957**; 22:365-367. b) Kumar S K, Patnaik A. Tunable Electronic Properties of a Proton-Responsive N,N-Dimethyl aminoazobenzene Fullerene (C₆₀) Dyad. *Chem. phys. chem.* **2010**; 11:3645-3655.