

Biological tools for microbial control: *Leuconostoc mesenteroides* and *Lactiplantibacillus plantarum* strains isolated from Apulian honeys as promising antifungal agents

Ester Presutto^{1*}, Maria Lucia Valeria de Chiara², Vittorio Capozzi², Giuseppe Spano¹, Mariagiovanna Fragasso¹

¹Department of Agriculture Food Natural Science Engineering, University of Foggia, Via Napoli 25, 71122, Foggia, Italy

²Institute of Sciences of Food Production, National Research Council (CNR), c/o CS-DAT, Via Michele Protano, Foggia, Italy

*Corresponding author: ester.presutto@unifg.it



Università di Foggia



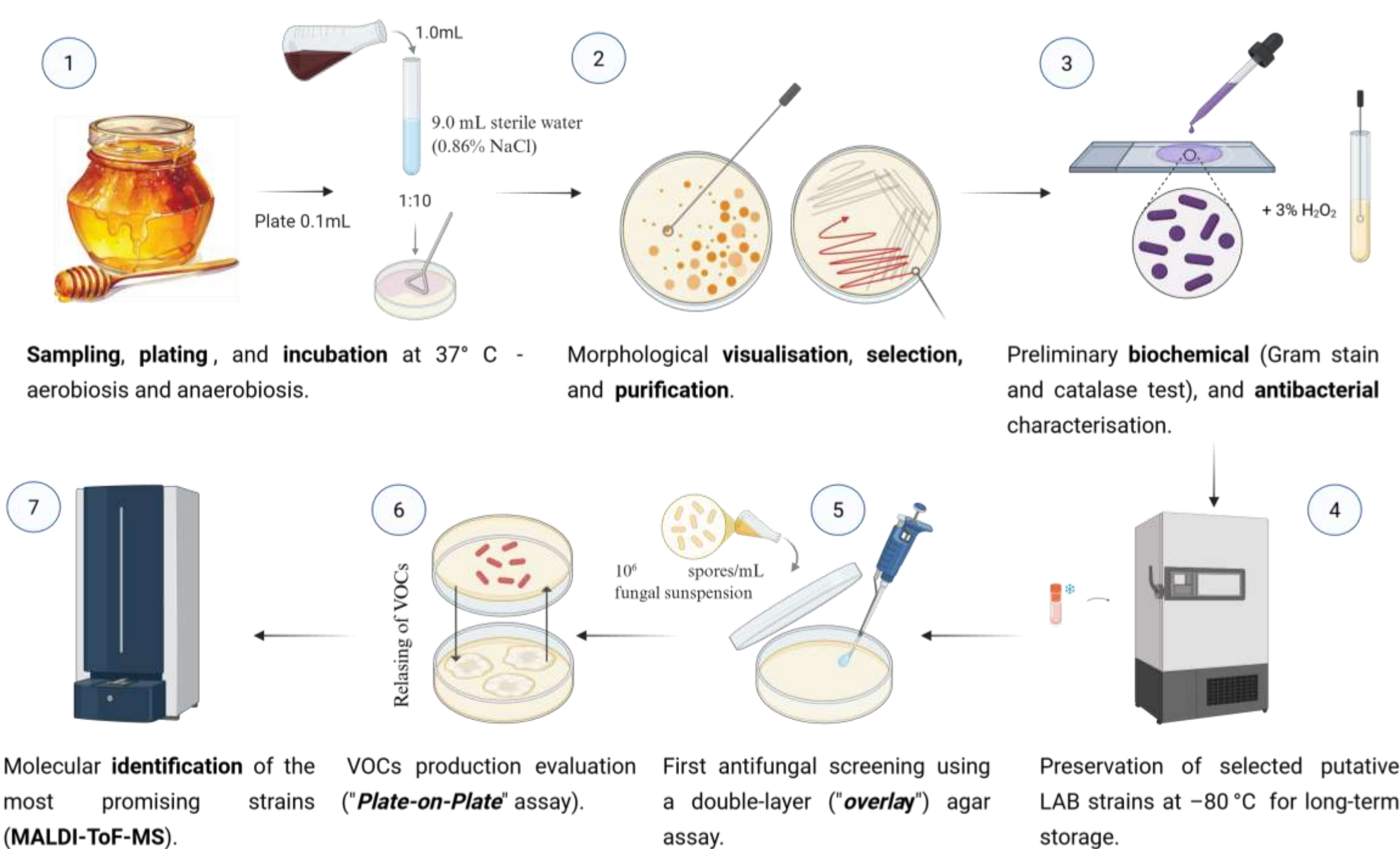
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INTRODUCTION & AIM

Lactic acid bacteria (LAB) are emerging as valuable agents for microbial biocontrol, due to their ability to produce broad-spectrum antagonistic compounds, including organic acids, bacteriocins, bioactive peptides, short-chain fatty acids, and volatile organic compounds (VOCs).

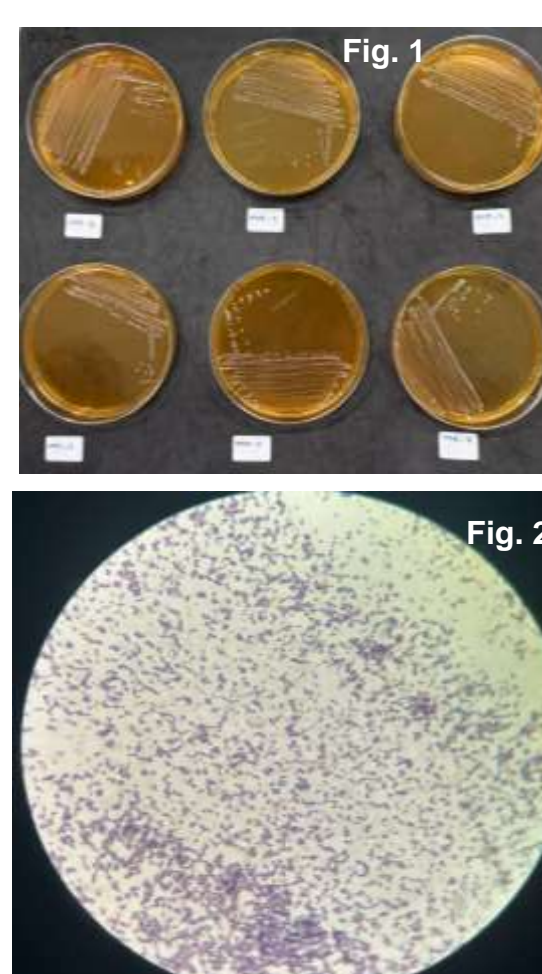
The use of LAB as natural preservatives aligns with current trends toward sustainable and eco-friendly food production systems, offering an alternative to synthetic chemical preservatives. The aim of this study was to assess the antifungal potential of 10 LAB strains belonging to the species *Leuconostoc* (*Leuc.*) *mesenteroides* and *Lactiplantibacillus plantarum*, isolated from five varieties of Apulian honey: i) honeydew honey, ii) French honeysuckle (*Sulla*) honey, iii) wildflower honey, iv) coriander honey, and v) *Eucalyptus* honey. Antifungal activity was investigated through a sequential assay combining the agar overlay method and the 'Plate-on-Plate' technique, tested against *Aspergillus* (*A.*) *niger*, *Penicillium* (*P.*) *crustosum*, *P. roqueforti*, *Aureobasidium* (*Au.*) *pullulans* and *Geotrichum* spp.

METHODS



Tab. 1

ID strains	Isolation source	Species
MME-1	Honeydew honey	<i>Leuconostoc mesenteroides</i>
MME-2	Honeydew honey	<i>Leuconostoc mesenteroides</i>
MME-3	Honeydew honey	<i>Leuconostoc mesenteroides</i>
MME-4	Honeydew honey	<i>Leuconostoc mesenteroides</i>
MME-9	Honeydew honey	<i>Lactiplantibacillus plantarum</i>
MM1	Wildflower honey	<i>Lactiplantibacillus plantarum</i>
MM9	Wildflower honey	<i>Lactiplantibacillus plantarum</i>
MS1-CB	<i>Sulla</i> honey	<i>Lactiplantibacillus plantarum</i>
ME-4	<i>Eucalyptus</i> honey	<i>Lactiplantibacillus plantarum</i>
MC-1	Coriander honey	<i>Lactiplantibacillus plantarum</i>



LAB strains were preliminarily selected based on morphological and biochemical characterisation (Figs. 1–2). The most promising isolates were then identified at the species level by Matrix-Assisted Laser Desorption/Ionization–Time of Flight Mass Spectrometry (MALDI-ToF MS).

CONCLUSIONS

The results support the application of LAB from non-conventional matrices as bioprotective agents in food and packaging systems, promoting the development of 'green' solutions in line with the principles of sustainability and the circular bioeconomy.

ACKNOWLEDGMENTS

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RESULTS & DISCUSSION

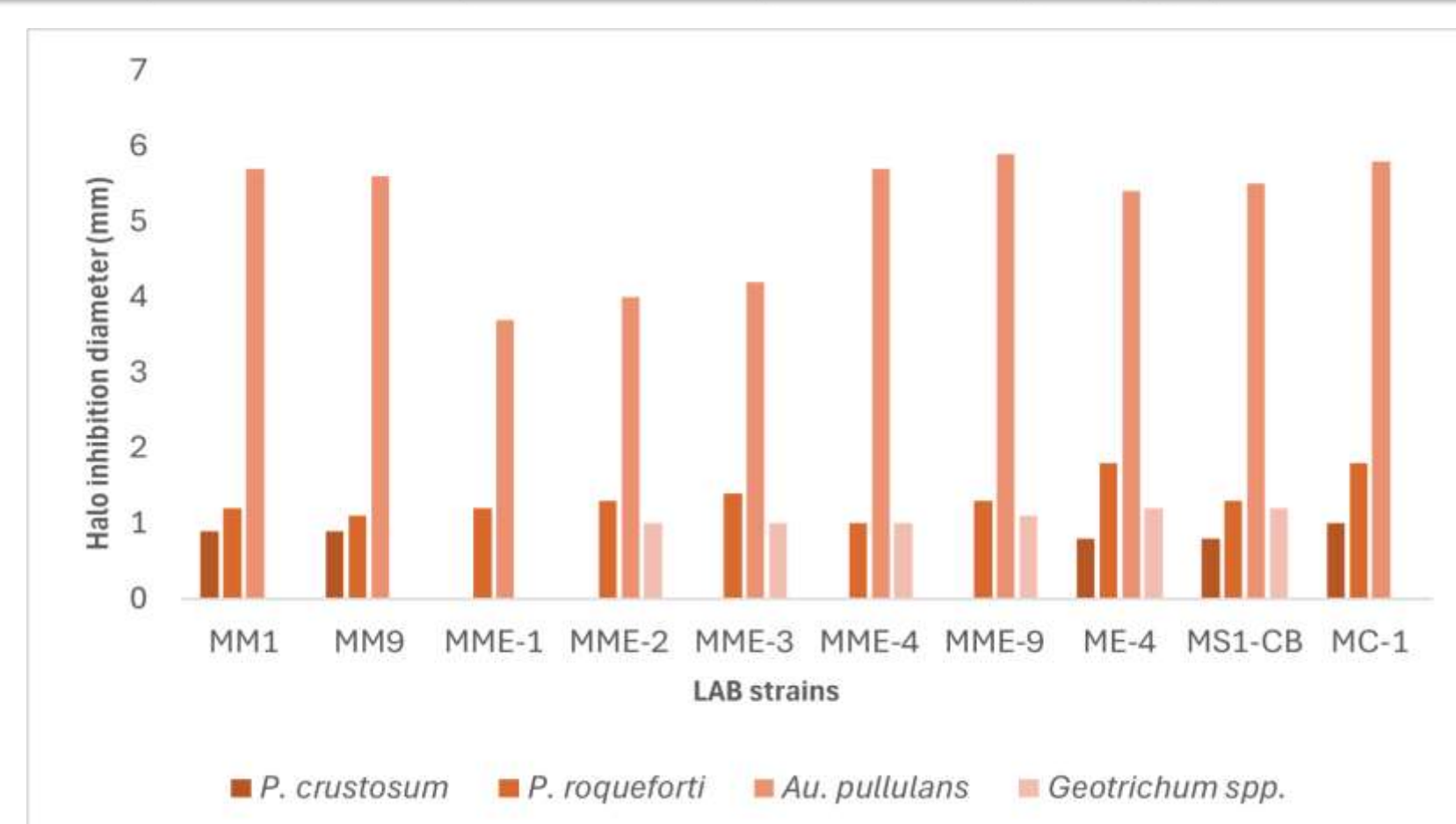


Fig. 3. Halo inhibition diameter (mm) of LAB strains using the overlay assay.

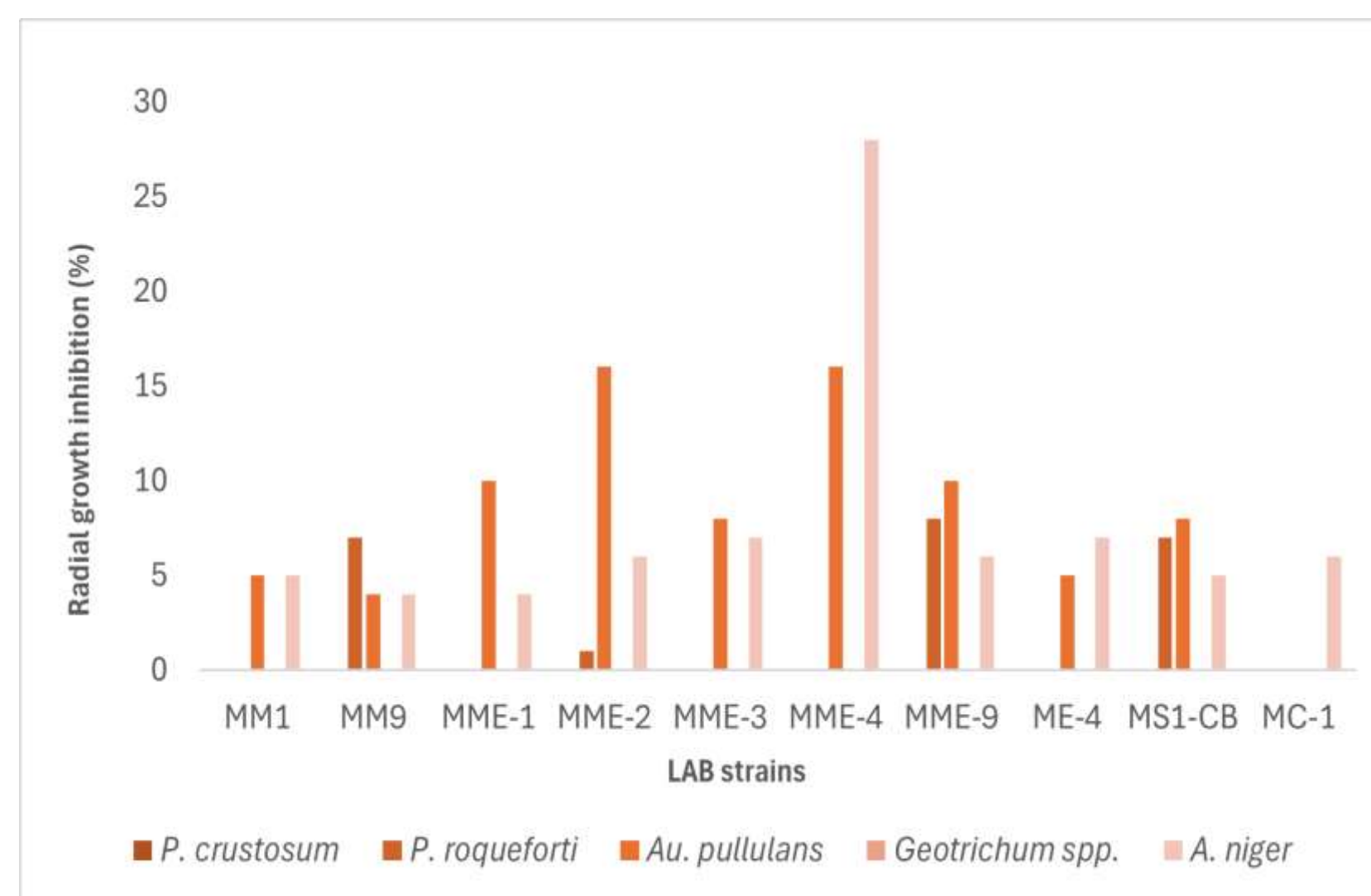
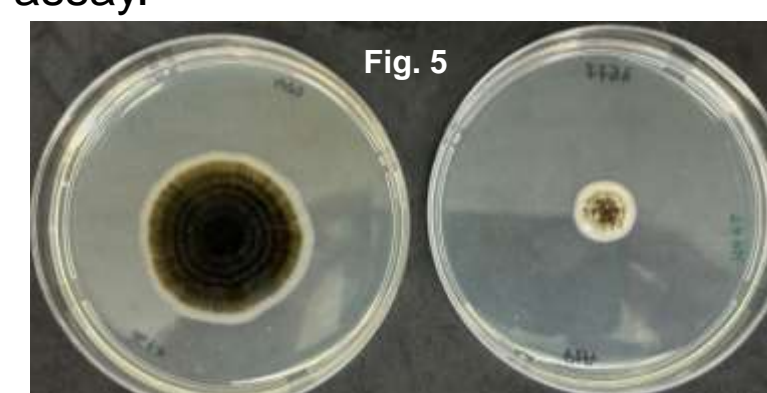


Fig. 4. Radial growth inhibition (%) by LAB VOCs using the 'Plate-on-Plate' assay.



In both cases, all strains exhibited moderate to high inhibitory activity against *P. roqueforti* and *Au. Pullulans* (Fig. 3-4). *Leuc. mesenteroides* MME-2 exhibited clear inhibition halos against *Au. pullulans* (Fig. 5). Complete inhibition of *A. niger* mycelial growth was observed exclusively for *Leuc. mesenteroides* MME-4 in the 'Plate-on-Plate' assay, suggesting the involvement of bioactive VOCs within the shared headspace (Fig. 6).

FUTURE WORK / REFERENCES

- Integrate omic approaches to link genotype and antimicrobial phenotype;
- Validate selected LAB strains in real food/packaging systems;
- Assess honey-derived LAB in sustainable food preservation and biocontrol system.