

Synthesis of Chitosan–Alginate–Gelatin Aerogel Using Freeze Drying and Dehydrator Techniques as a Matrix for Virgin Fish Oil-Based Oleogel

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INTRODUCTION & AIM

Omega-3 fatty acids (**EPA** and **DHA**) play a vital role in health, but the body converts less than 5% of ALA, making direct intake highly recommended. **Virgin fish oil**, a rich source of **DHA**, faces challenges such as low solubility, oxidation, and undesirable odor. **Oleogels** offer an innovative solution to improve oil stability without altering its structure, featuring thermo-reversible, viscoelastic, and self-standing properties (Li *et al.* 2019).

The **indirect oleogelation** method using aerogels is considered safer for heat-sensitive compounds (Plazzotta *et al.* 2019). **Aerogels**, porous materials with high surface area, can effectively bind oils and are safe for food applications. The use of marine-based biopolymers such as **chitosan**, **alginate**, and **gelatin** as aerogelators remains underexplored, including **comparisons of drying methods**, presenting a promising research opportunity.

This study aims to develop and evaluate chitosan, alginate, and gelatin-based **aerogels as templates** for fish oil oleogel synthesis, as well as to assess the effect of conventional drying methods (**freeze drying** and **atmospheric drying**) on the **structure** and **absorption capacity** of the aerogels.

METHOD



REFERENCES

- Li L, Wan W, Cheng W, Liu G, Han L. 2019. Oxidatively stable curcumin-loaded oleogels structured by β -sitosterol and lecithin: physical characteristics and release behaviour in vitro. *International Journal of Food Science and Technology*. 54(7):2502–2510. doi:10.1111/ijfs.14208.
- Plazzotta S, Calligaris S, Manzocco L. 2019. Structure of oleogels from karrageenan templates as affected by supercritical-CO₂-drying, freeze-drying and lettuce-filler addition. *Food Hydrocolloids*. 96:1–10. doi:10.1016/j.foodhyd.2019.05.008.
- Wang Z, Zhu W, Huang R, Zhang Y, Jia C, Zhao H, Chen W, Xue Y. 2020. Fabrication and characterization of cellulose nanofiber aerogels prepared via two different drying techniques. *Polymers*. 2583:1–13. doi:10.3390/polym12112583.

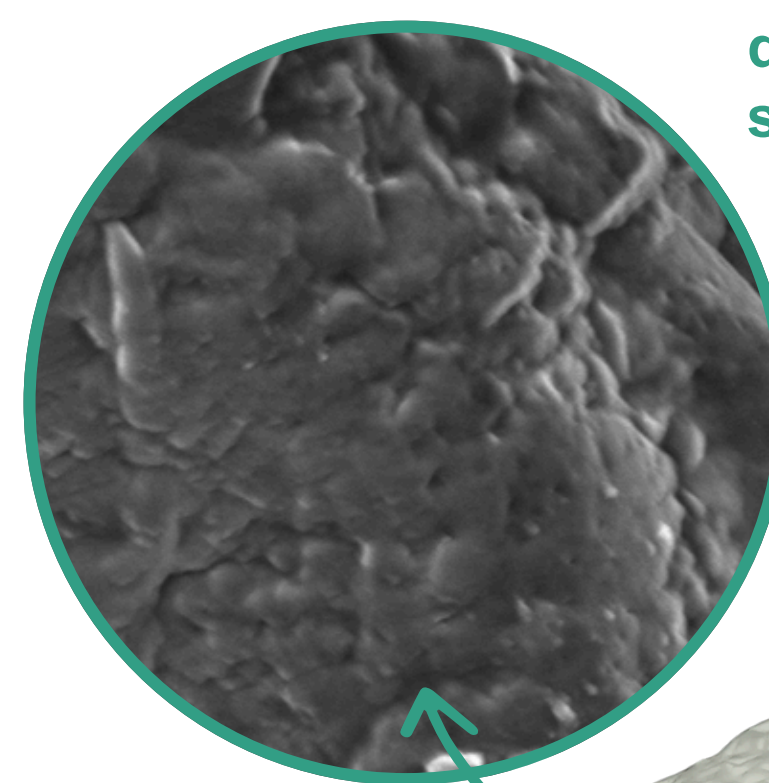
RESULTS & DISCUSSION

Cylindrical in shape, **smooth surface without bubbles or cracks**, **homogeneous semi-transparent white color**, **elastic** and **not brittle**.

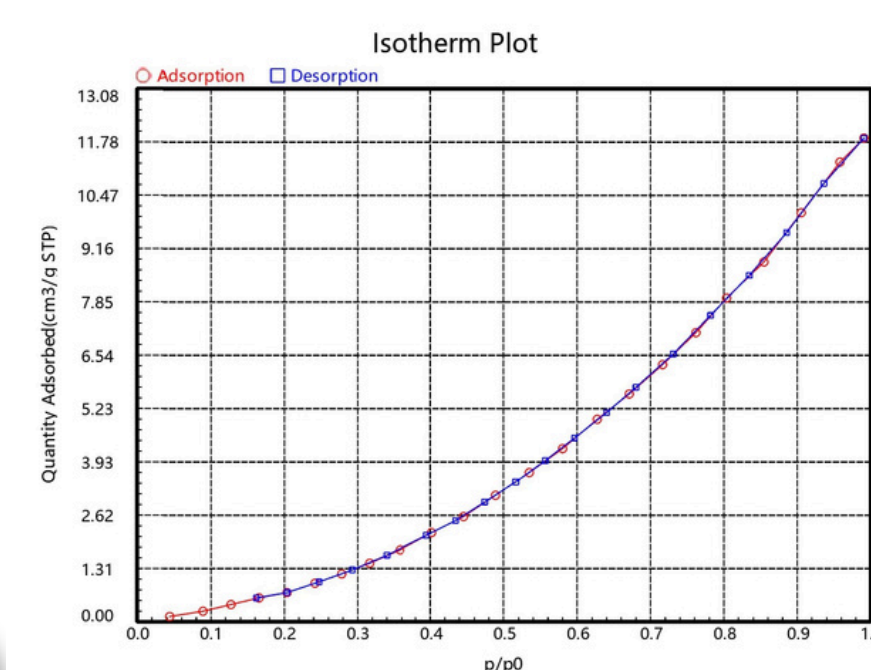


Swelling Ratio
9,63±0,94%

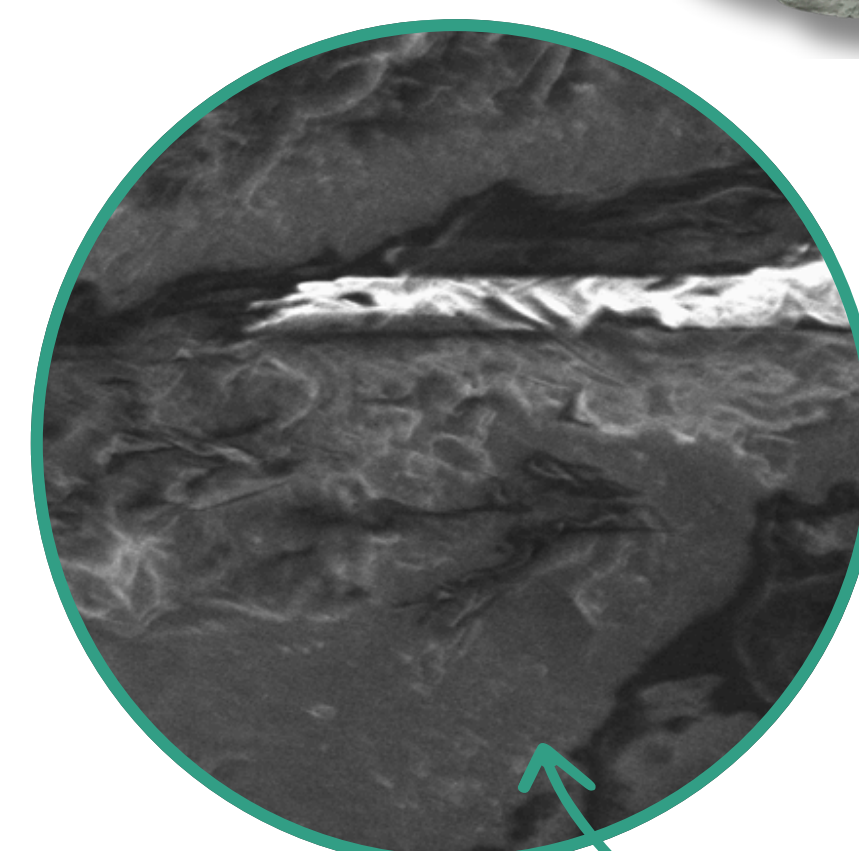
A layered morphology with a **wavy surface**, irregular **large cavities**, and a preserved **three-dimensional porous structure** resulting from sublimation.



Surface area
15,629 m²·g⁻¹
Average pore diameter
4,701 nm



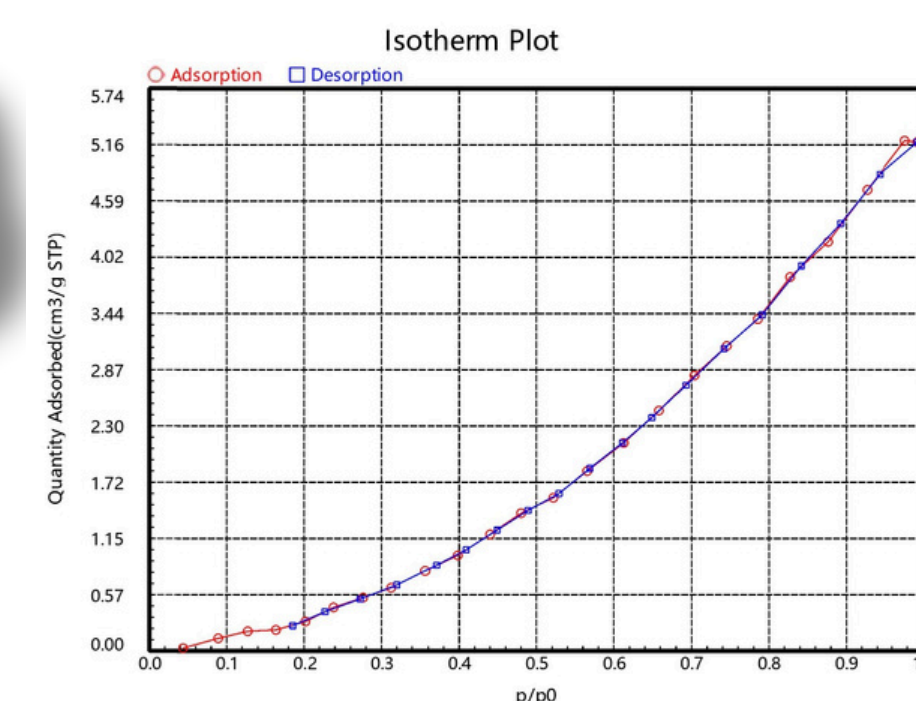
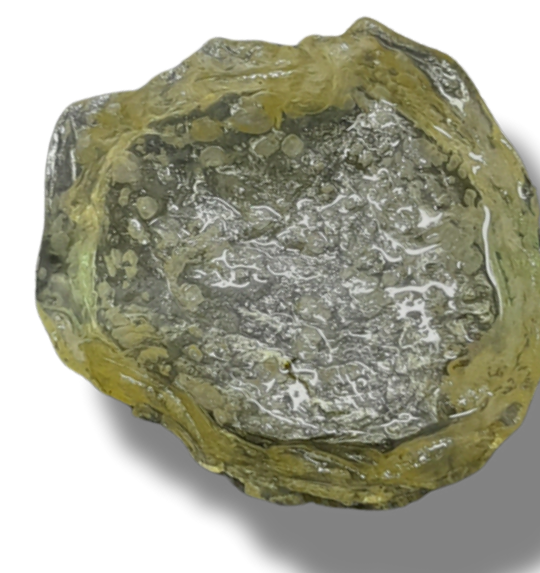
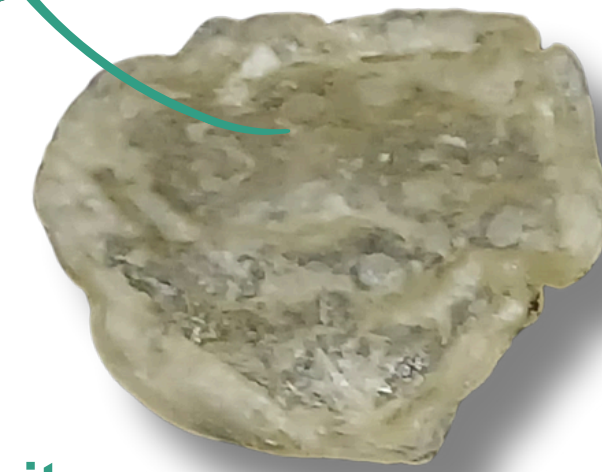
Aerogel Freeze Drying



Aerogel Dehydrator

Absorption capacity
Value **not detected** significantly

Oil Binding Capacity
Value **not detected** significantly



Average pore diameter
4,450 nm

Surface area
7,241 m²·g⁻¹

Absorption capacity
4,48±0,21 g/g
Oil Binding Capacity
85,59 ± 3,73%

A **dense, folded, and wrinkled surface** with **no open pores**, resulting from **structural collapse** caused by **capillary forces** during water evaporation at **high temperatures**.

CONCLUSION & FUTURE WORK

Chitosan, alginate, and gelatin-based aerogels were **successfully developed as matrices** for oleogel synthesis, with **freeze drying showing superior** results. These aerogels feature an **open-porous three-dimensional** structure with an average **pore size of 4.701 nm**, **high absorption capacity of 4.48 ± 0.21 g/g**, and significantly higher **oil-binding capacity of 85.59 ± 3.73%**.

Optimization of the biopolymer composition is needed to achieve improved **rheological** and **morphological** properties. Future research could focus on **evaluating the oxidative stability** of oleogels during storage, as well as **characterizing their mechanical** and **thermal properties**.