


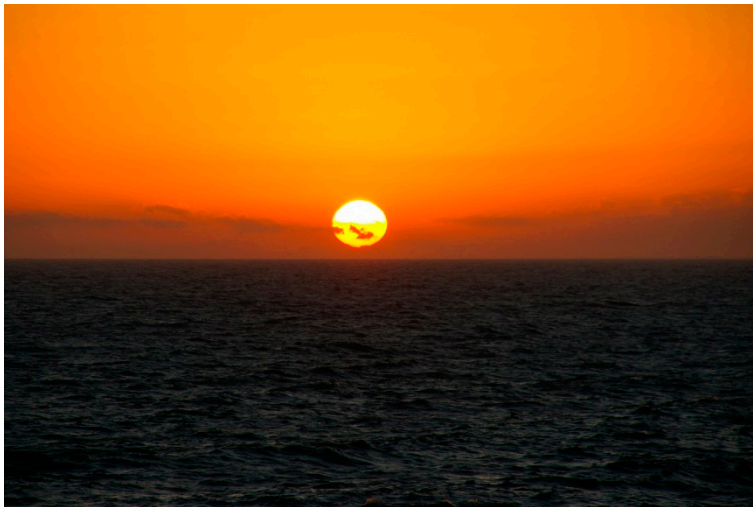
Light Exposure in Microscopy – How can Cell Survival be Increased?

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Fluorescence Microscopy of Living Cells



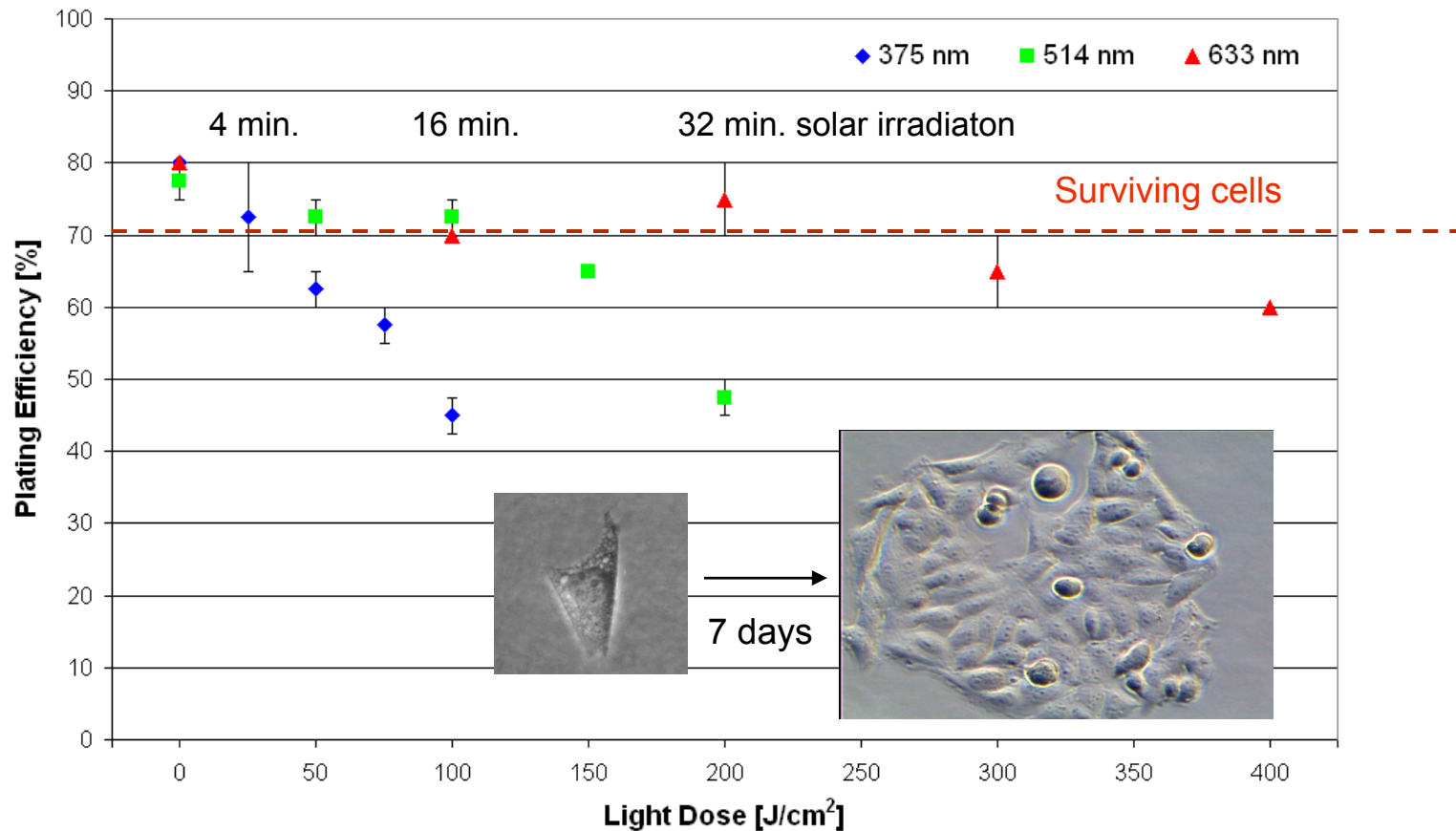
- How much light do we need for microscopy and how much light can we apply to living cells?
- Can we use or even exceed solar irradiance?
- How long will cells endure this irradiation?

Solar irradiance:

$$1 \text{ kW/m}^2 = 100 \text{ mW/cm}^2 = 1 \text{ mW/mm}^2 = 1 \text{ nW}/\mu\text{m}^2$$

$$1 \text{ J/cm}^2 = 10 \text{ s of solar irradiation}$$

Viability of U373-MG Glioblastoma Cells upon Irradiation (colony formation assay; native cells)

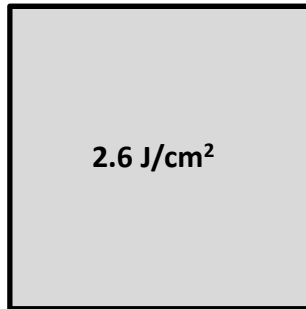


Cell Viability upon Irradiation

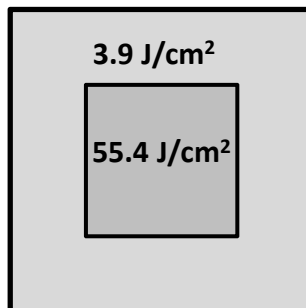
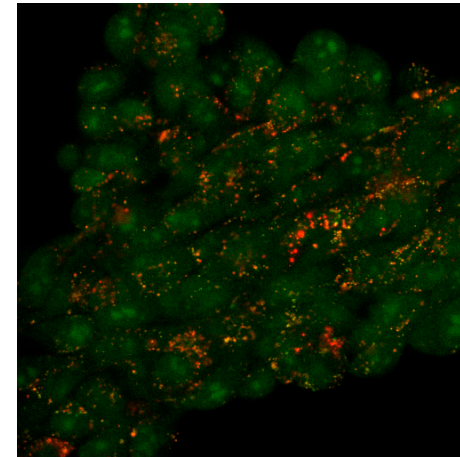
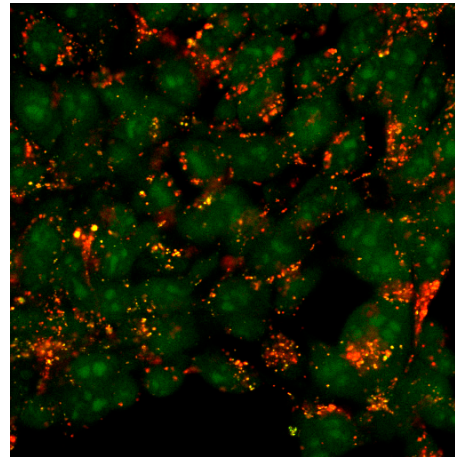
- native cells and fluorescent markers -

Cell line	Marker	Conc. [μM]	λ_{ex} [nm]	Max. light dose [J/cm ²]	Solar exposure time [s]
U373-MG	–		375	25	250
U373-MG	–		514	100	1000
U373-MG	–		633	200	2000
U373-MG	Laurdan	8	391	10	100
CHO-K1	DiA	5	488	10	100
CHO-K1	DiO	5	488	10	100
CHO-K1	GFP-Mem		488	10	100
CHO-K1	R 123	5	488	5–20	50–200
CHO-K1	MTO	0.05	514	50	500
CHO-K1	GFP-Mito		488	5	50

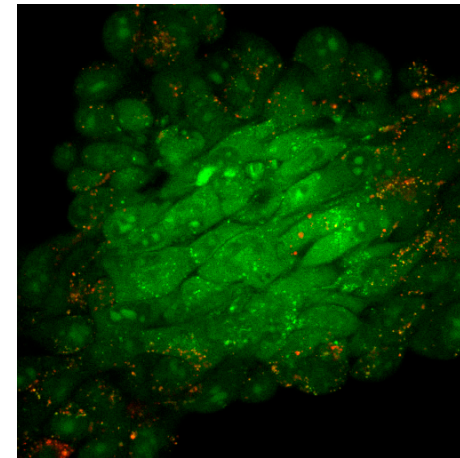
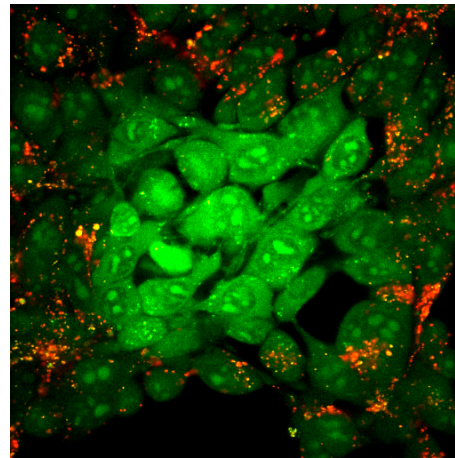
Example: 3T3 Fibroblasts + Acridine orange (5 μM , 30 min.)



Prior tp illumination



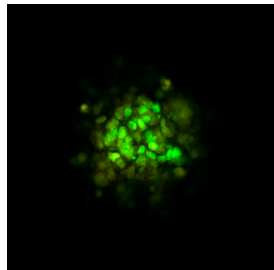
After illumination



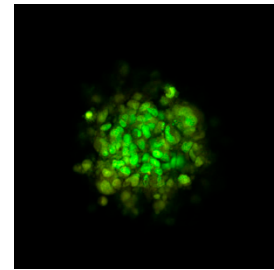
Fluorescence Microscopy with Axial Resolution

Methods:

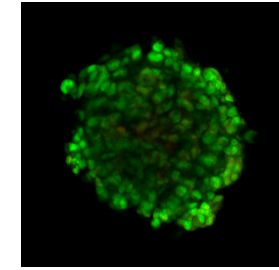
- Laser Scanning
Microscopy (LSM)



$z = 10 \mu\text{m}$

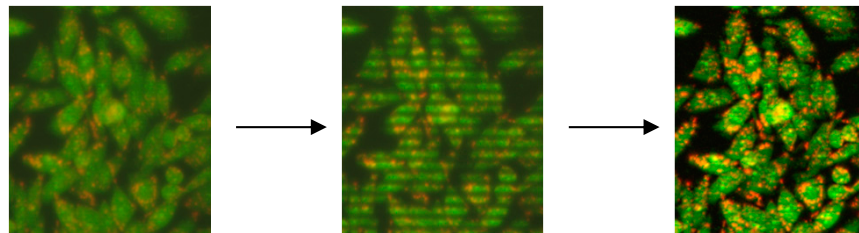


$z = 15 \mu\text{m}$

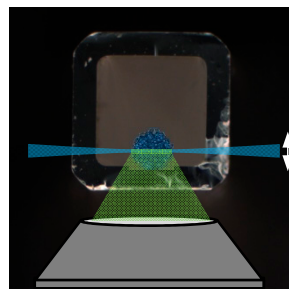


$z = 60 \mu\text{m}$

- Structured
Illumination
Microscopy (SIM)

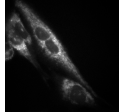
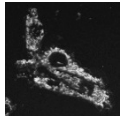
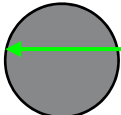
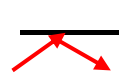


- Light Sheet
Fluorescence
Microscopy (LSFM)



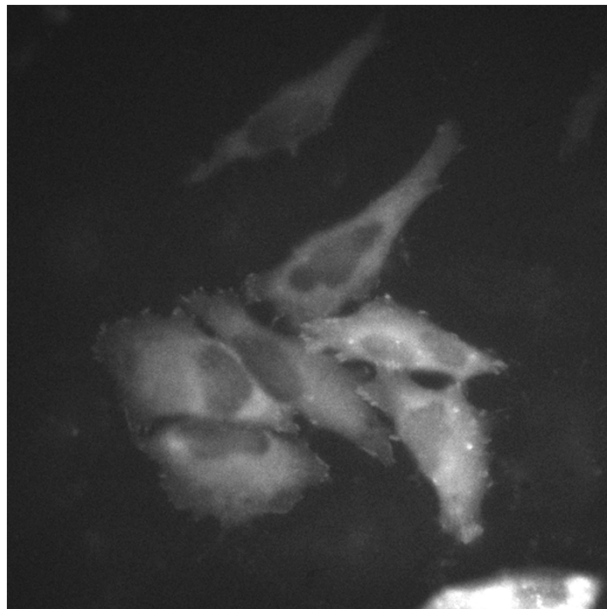
Only planes under
investigation are illuminated
→ minimum light exposure

Maximum Light Exposure to Living Cells in 3D Microscopy

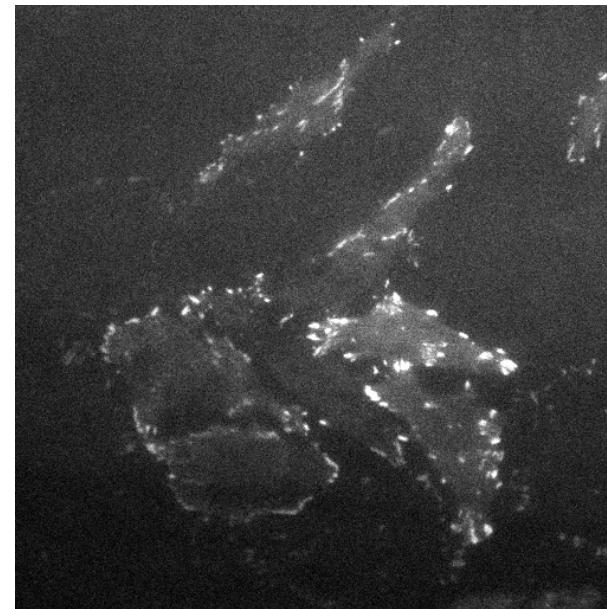
Method	Experiment	Max.light dose [nJ/ μm^2]	Irradiance [nW/ μm^2]	Max. no. of images
 Widefield microscopy	Autofluor.	250	1	250
 LSM	Fluor. marker	100	1	20
 Light Sheet (N layers)	Fluor. marker	100	1	$N \times 100$
 TIRFM	Fluor. marker	300	1	100–300
Single Molecule Methods	Fluor. Marker (low conc.)	2,000	500	≤ 1
STED (650 nm)	Fluor. Marker	100–500	30,000	not relevant

Total Internal Reflection Fluorescence Microscopy (TIRFM)

Membrane Associated Paxilline (Pax-EYFP) / Focal Adhesions



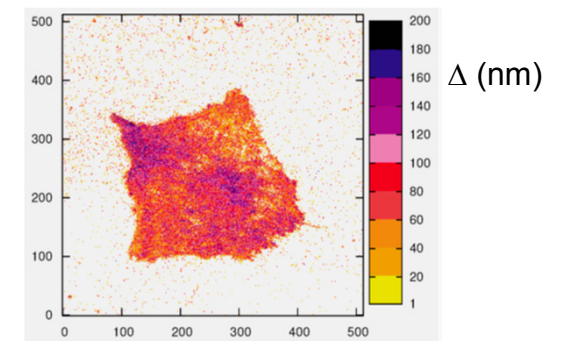
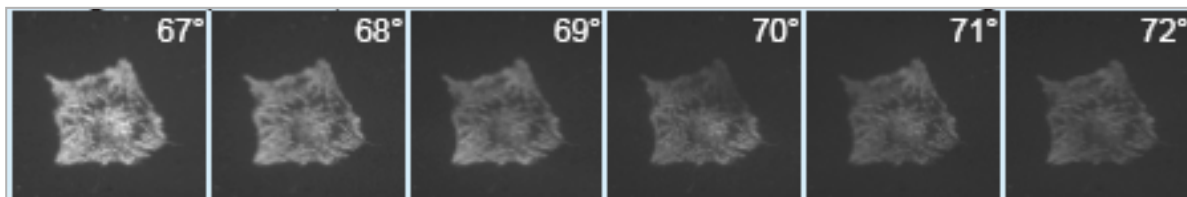
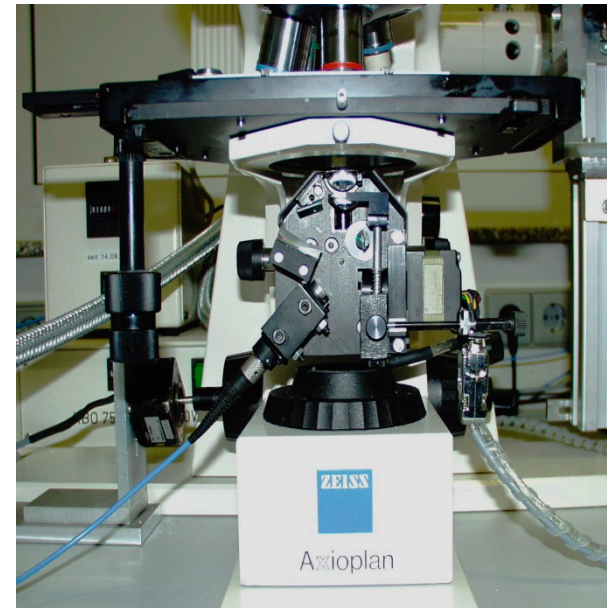
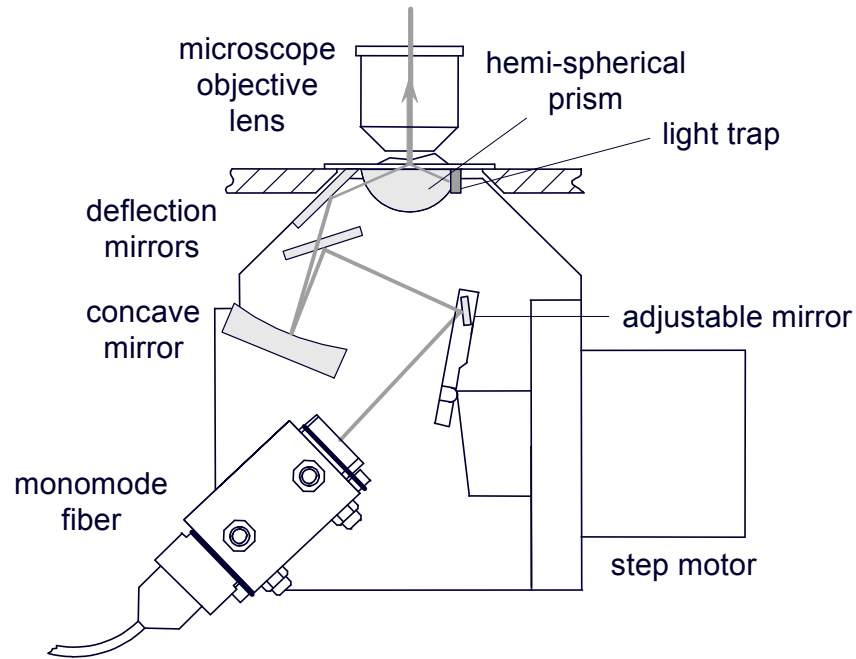
Conventional fluorescence microscopy



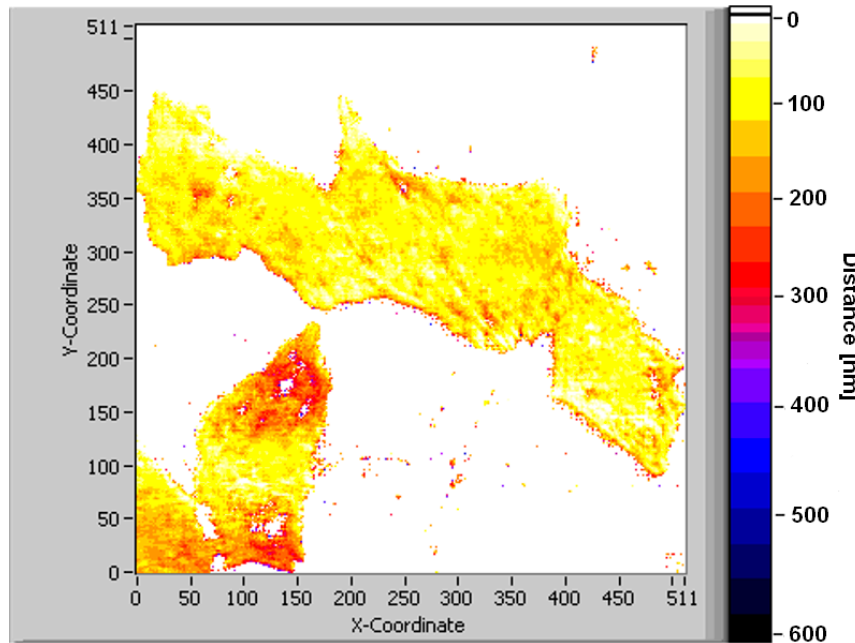
TIRFM

$\lambda_{\text{ex}} = 470 \text{ nm}; \lambda_{\text{d}} \geq 530 \text{ nm}$

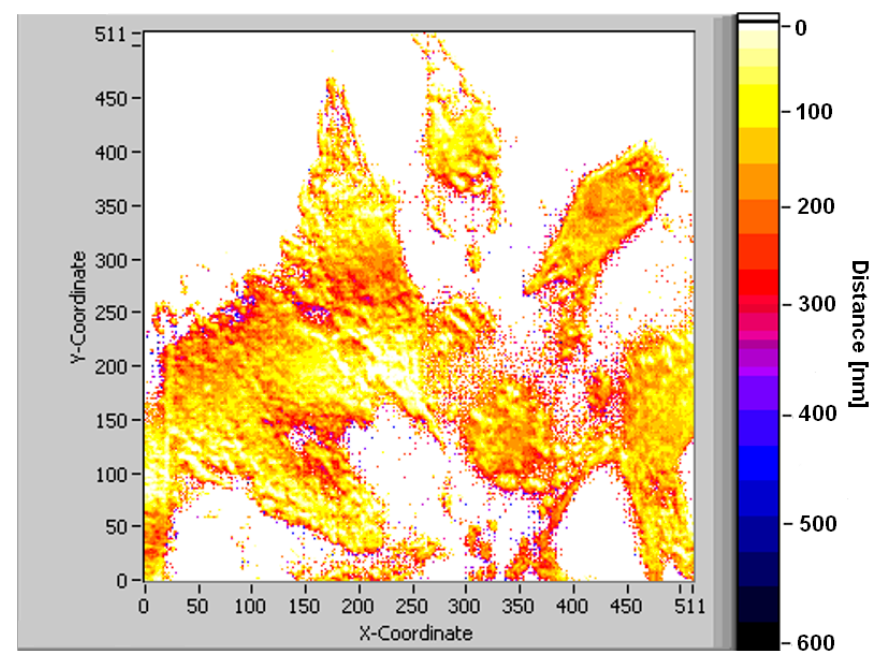
Variable-angle Total Internal Reflection Fluorescence Microscopy (TIRFM)



Nanometre Cell-Substrate Topology of Glioblastoma Cells - using the fluorescent membrane marker laurdan -



U251-MG tumour cells

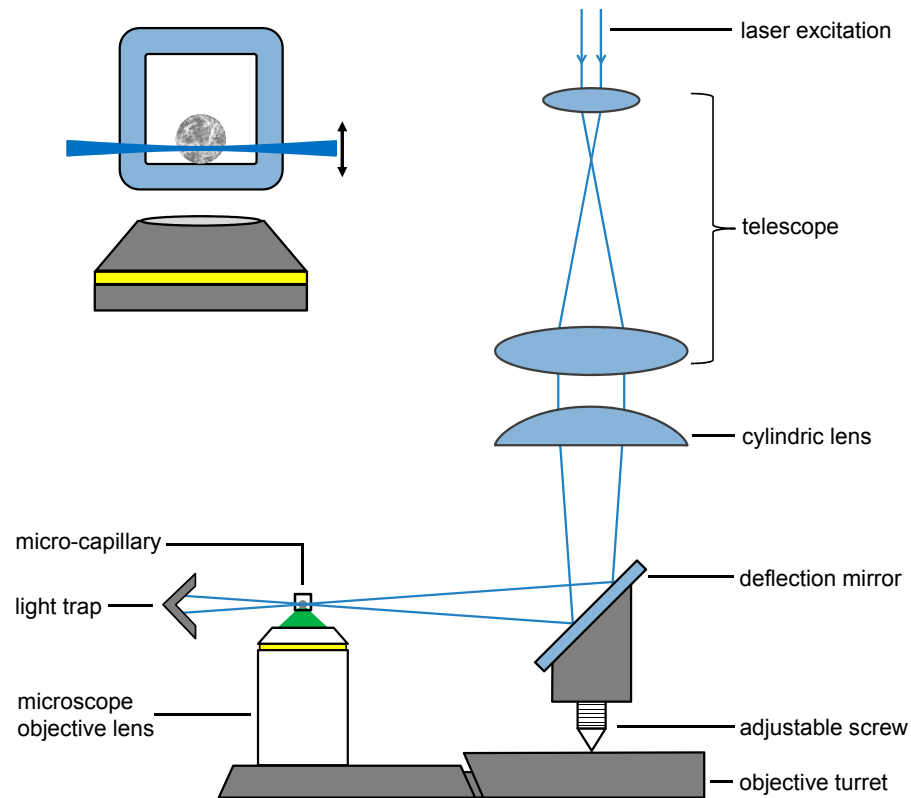


U251-MG with tumour suppressor gene TP53

Cell-substrate topology offers a criterion to distinguish tumour cells and less malignant cells

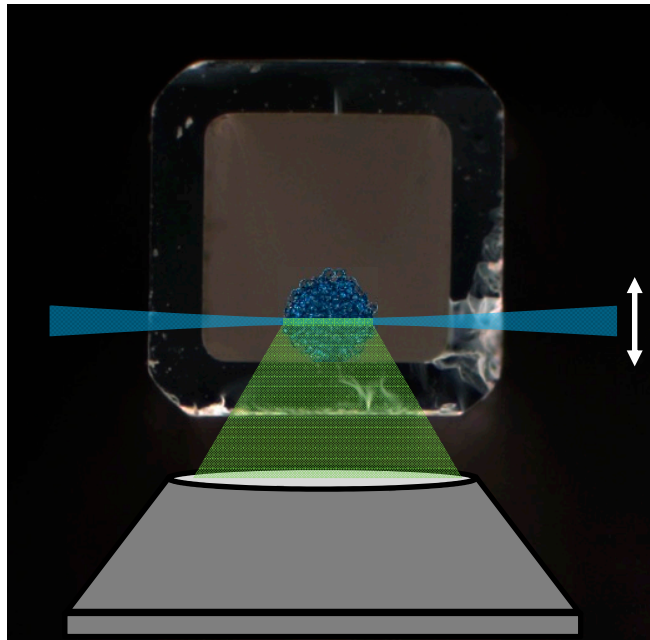
Cells provided by J. Mollenhauer, Dept. of Molecular Oncology, University of South Denmark, Odense

Light Sheet Fluorescence Microscopy (LSFM)

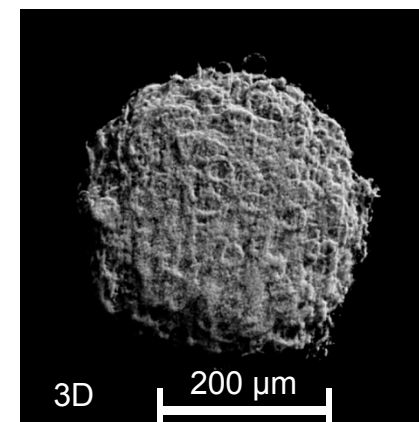
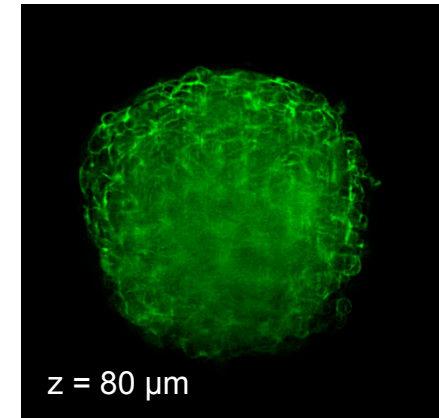
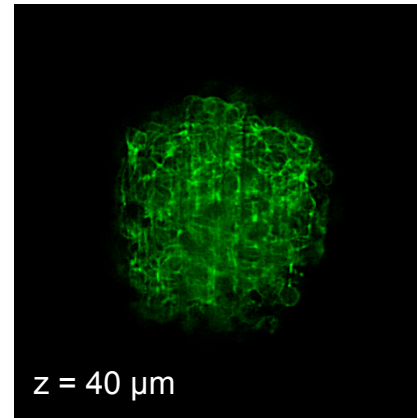


T. Bruns, S. Schickinger, R. Wittig and H. Schneckenburger, "Preparation strategy and illumination of 3D cell cultures in light sheet-based fluorescence microscopy," *J. Biomed. Opt.* 17, 101518 (2012).

Light Sheet Fluorescence Microscopy (LSFM) (CHO-GFP-Mem)

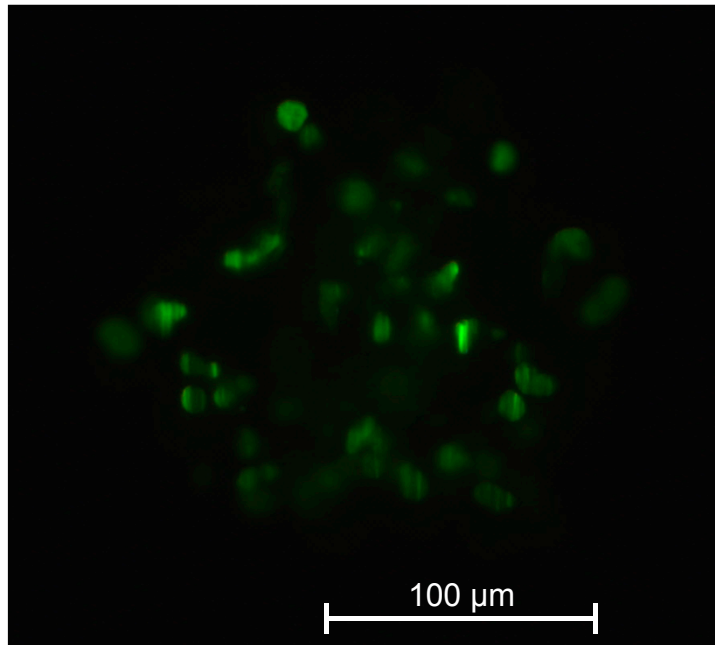


Selected parameters:
Beam waist: $\Delta z = 5\text{--}10\ \mu\text{m}$
Beam width: $\Delta y \approx 8\ \text{mm}$
Focal depth: $\Delta x \approx 150\text{--}200\ \mu\text{m}$

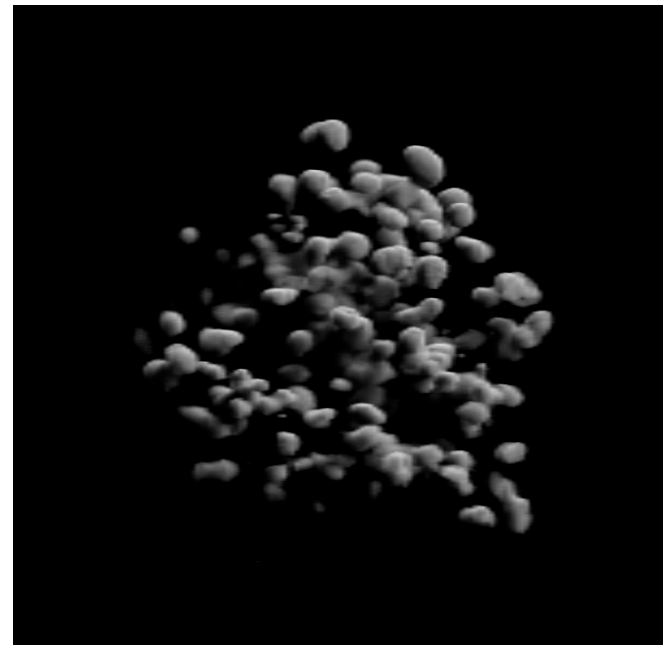


LSFM Application: 3D Imaging of Necrotic Cells

(Rotenone: 1 μM , 3 h; CellTox: 2 h; $\lambda_{\text{ex}} = 470 \text{ nm}$, $\lambda_{\text{d}} \geq 515 \text{ nm}$)



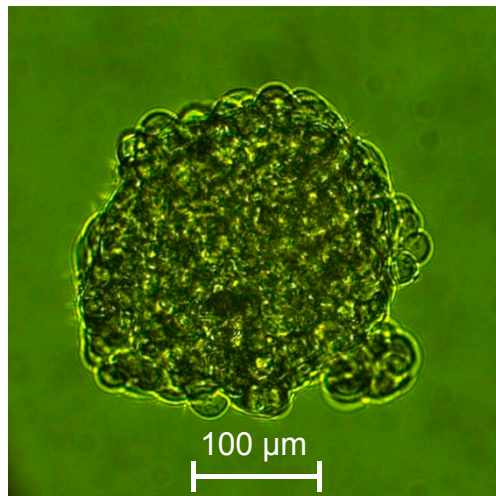
Single Plane: $z = 50 \mu\text{m}$; $d \leq 10 \mu\text{m}$



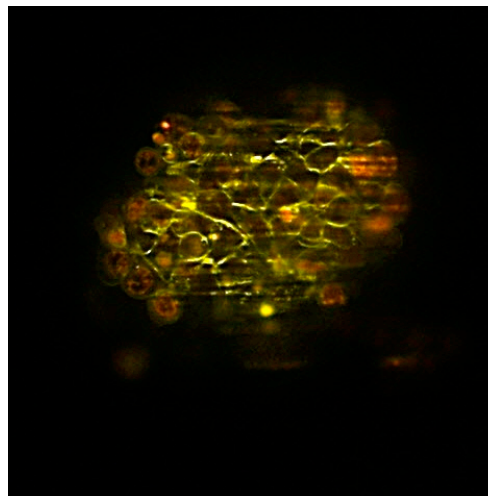
3D Reconstruction

LSFM Application: Uptake / Interaction of a Cytostatic Drug

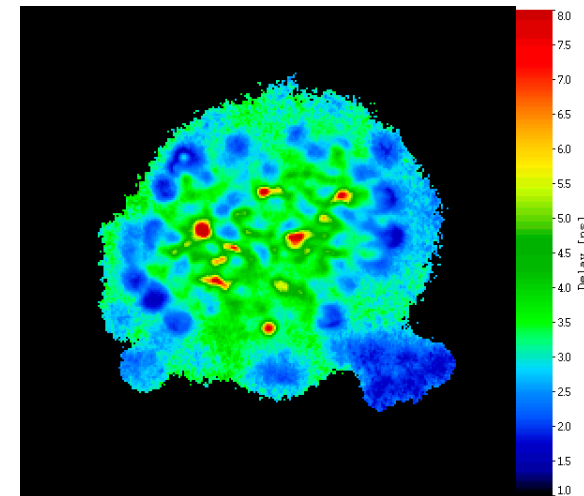
MCF-7 Breast Cancer Cells, Doxorubicin: 8 μM , 6 h, $\lambda_{\text{ex}} = 470 \text{ nm}$, $\lambda_{\text{d}} \geq 515 \text{ nm}$



Transillumination



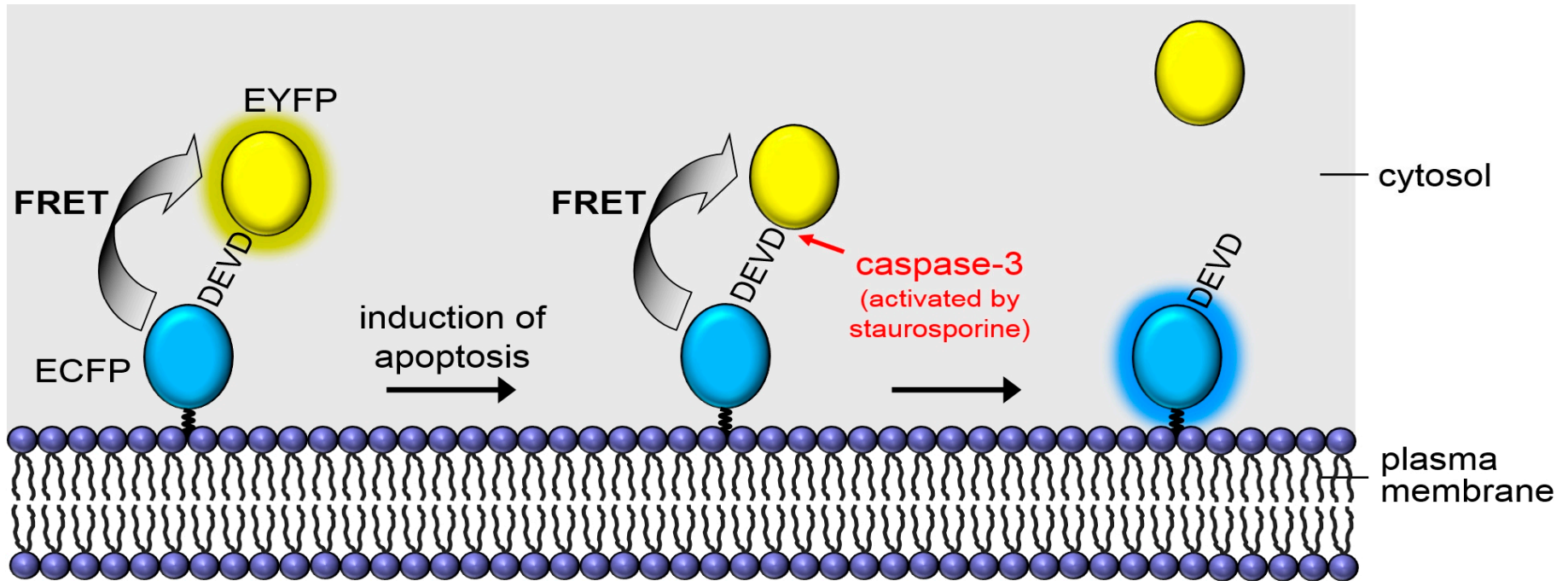
Fluorescence (single plane)



Fluorescence Lifetime

Fluorescence lifetime imaging (FLIM) is used to probe intermolecular interactions of doxorubicin and to identify a degradation product

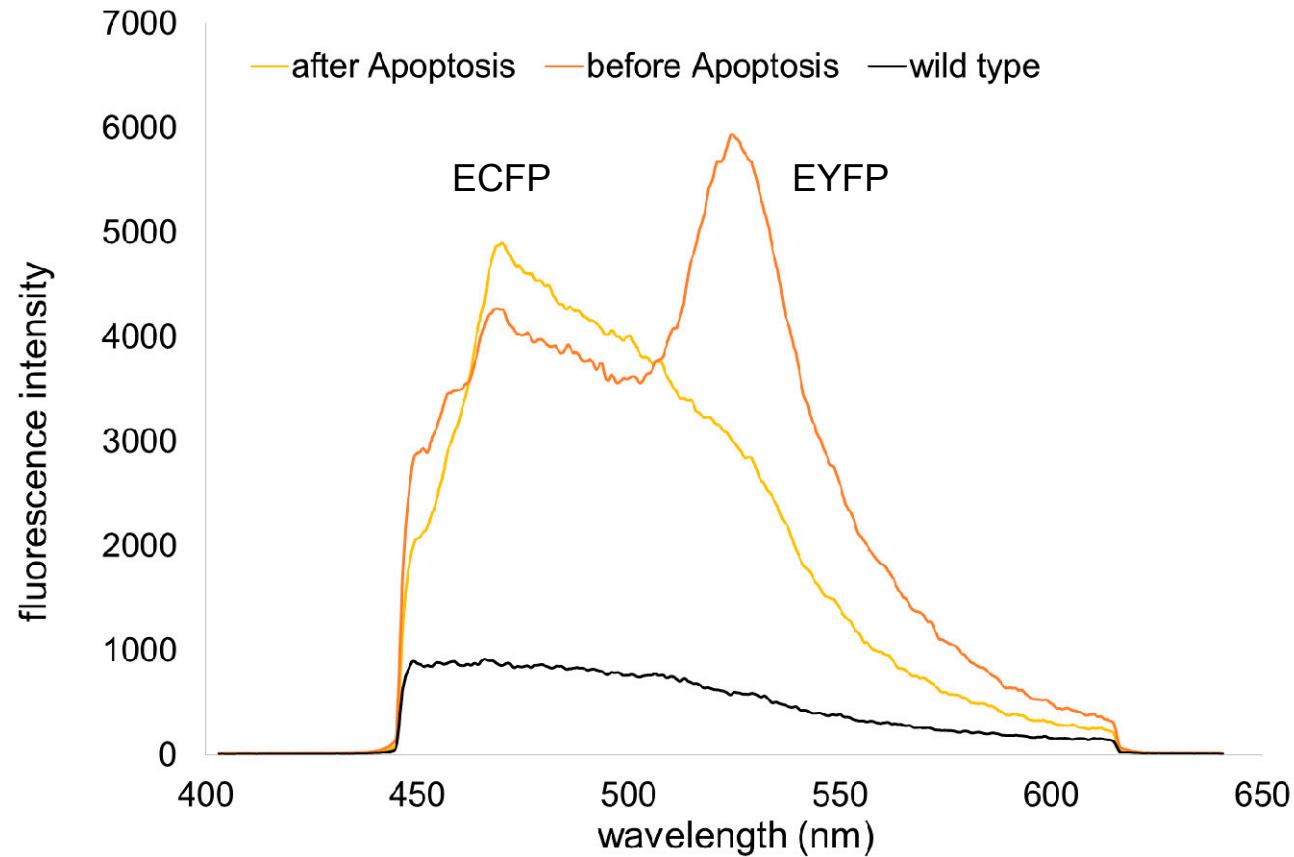
Förster Energy Transfer (FRET) Based Sensor for Apoptosis



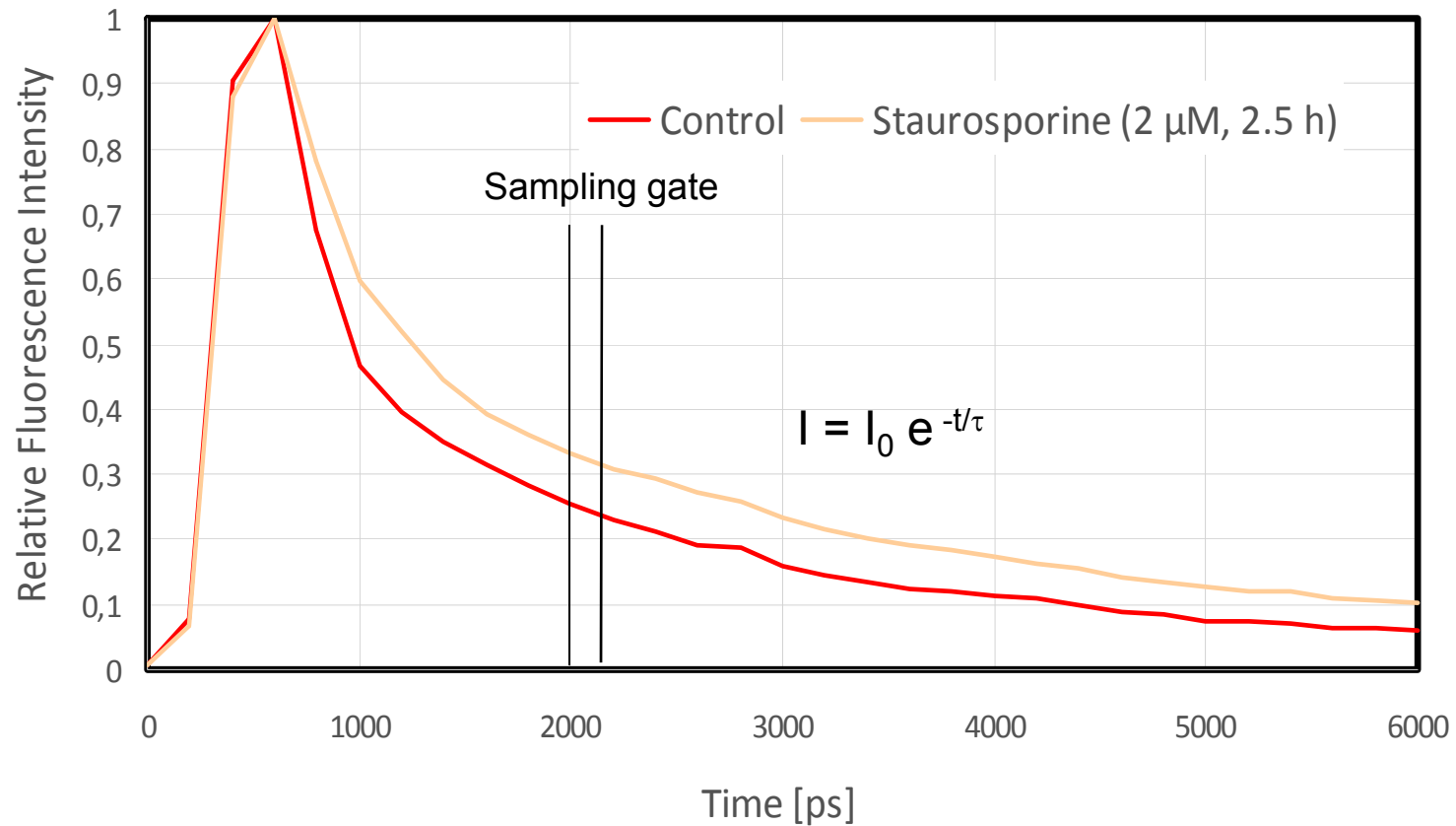
Non-radiative energy transfer from enhanced cyan fluorescent protein to enhanced yellow fluorescent protein via a cleavable peptide linker DEVD

Fluorescence Spectra prior to and subsequent to Apoptosis

$\lambda_{\text{ex}} = 391 \text{ nm (ECFP)}$

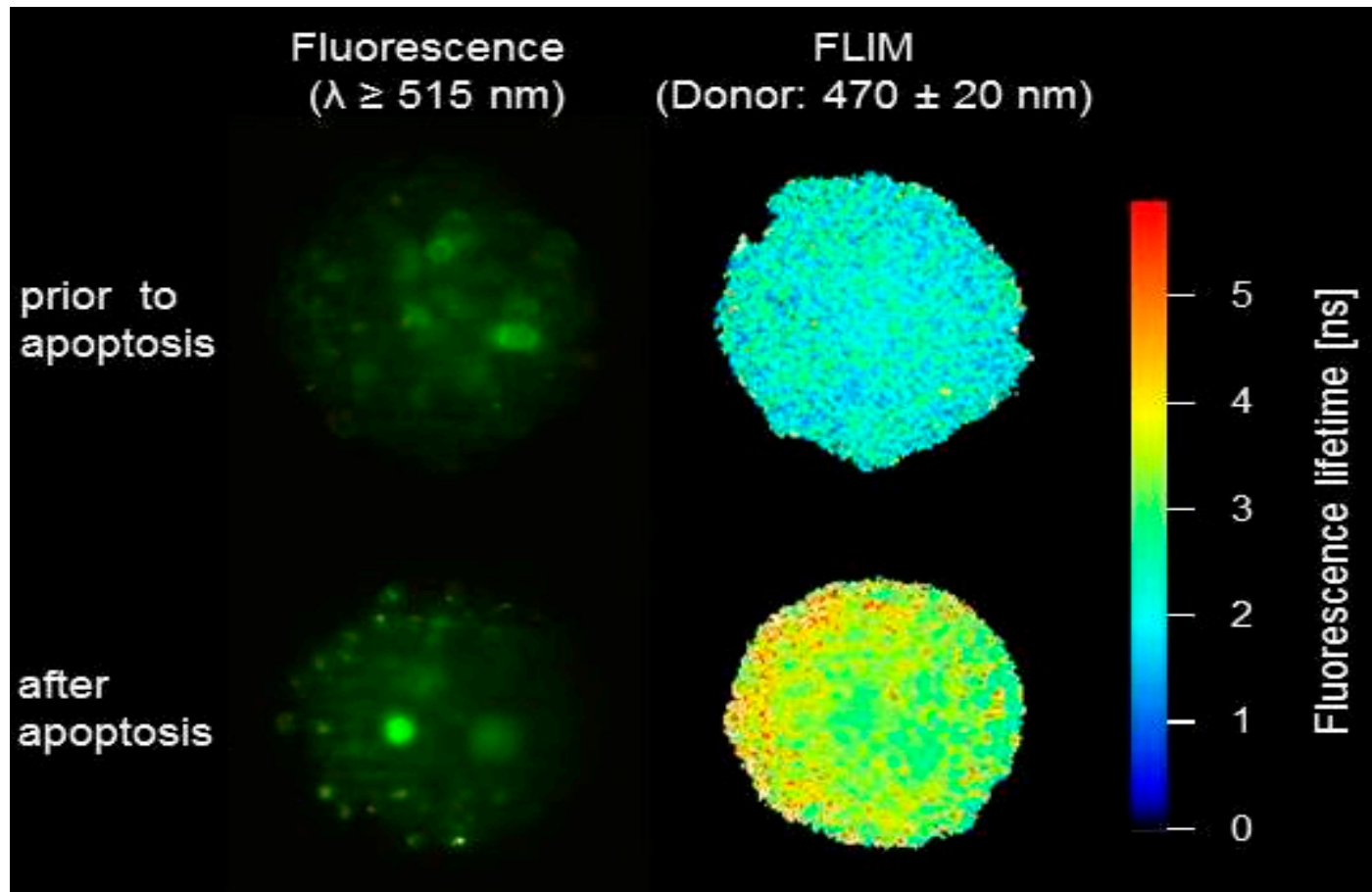


Fluorescence Decay Profiles prior to and subsequent to Apoptosis - HeLA-Mem-ECFP-DEVD-EYFP -



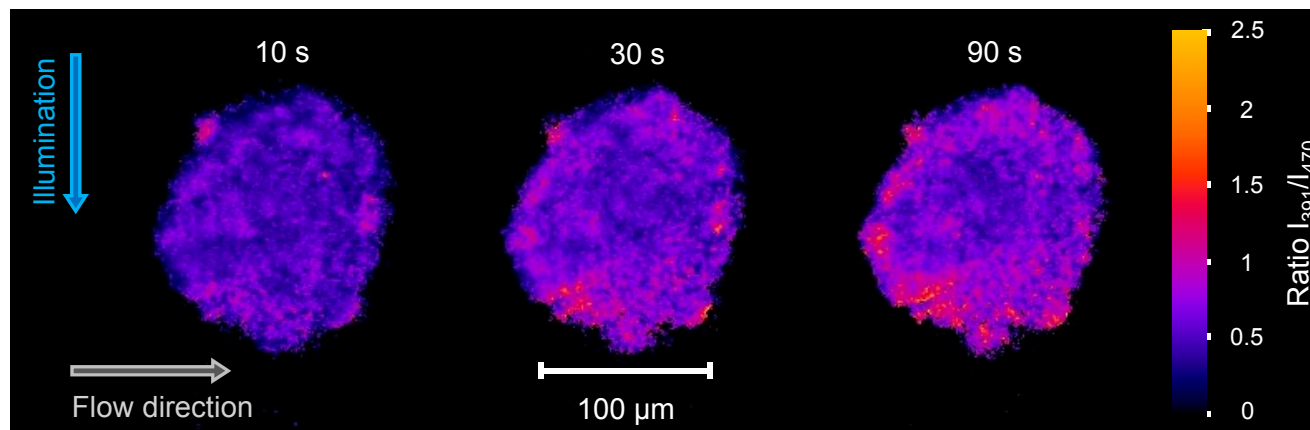
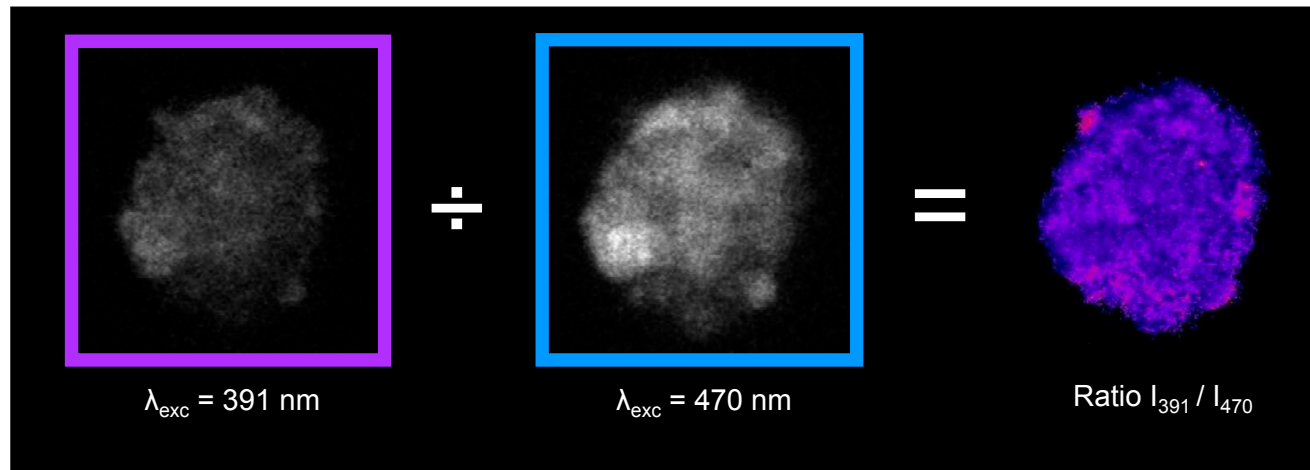
FRET-Based Membrane Associated Sensor for Apoptosis

- LSFM / FLIM of ECFP in HeLa Cells; $\lambda_{\text{ex}} = 391 \text{ nm}$ -




LSFM Application: Redox Imaging upon Addition of H_2O_2

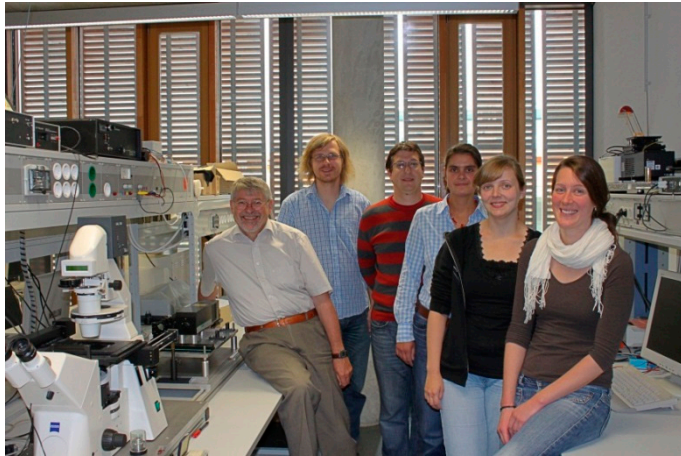
U251-MG glioblastoma cells with redox sensitive Grx1-roGFP2



Summary

- Light exposure in microscopy is limited due to phototoxicity
 - Maximum light doses for cell survival - dependent on wavelengths and fluorescence markers – are typically in the range of 5–200 J/cm² corresponding to 50 s – 2000 s of solar irradiance
 - Light sheet fluorescence microscopy (LSFM) needs minimum light doses for 3d samples, e. g. multicellular tumour spheroids (examples including apoptosis and necrosis, uptake of cytostatic drugs, redox imaging)
 - TIRFM needs minimum light doses for cell surfaces or membranes (examples including focal adhesions, cell-substrate topology)
- 

Acknowledgment / Literature



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The authors thank B. Angres and NMI Reutlingen for providing HeLa cells expressing the FRET sensor as well as R. Wittig (ILM Ulm) for his cooperation.

- H. Schneckenburger, P. Weber, M. Wagner, S. Schickinger, V. Richter, T. Bruns, W.S.L. Strauss, R. Wittig: Light exposure and cell viability in fluorescence microscopy, *J. Microsc.* 245 (2012) 311–318.
- M. Wagner, P. Weber, H. Baumann, H. Schneckenburger: Nanotopology of cell adhesion upon variable-angle total internal reflection fluorescence microscopy (VA-TIRFM), *J. Vis. Exp.* 68 (2012) e4133.
- T. Bruns, S. Schickinger, H. Schneckenburger: Single plane illumination module and micro-capillary approach for a wide-field microscope, *J. Vis. Exp.* 15(90) (2014) e51993.
- S. Schickinger, T. Bruns, R. Wittig, P. Weber, M. Wagner, H. Schneckenburger: Nanosecond ratio imaging of redox states in tumour cell spheroids using light sheet based fluorescence microscopy, *J. Biomed. Opt.* 18(12) (2013) 126007.
- P. Weber, S. Schickinger, M. Wagner, B. Angres, T. Bruns, H. Schneckenburger: “Monitoring of apoptosis in 3d cell cultures by FRET and light sheet fluorescence microscopy”, *Int. J. Mol. Sci.* 16(3) (2015) 5375–5385.