

### LIGAND-BASED PHARMACOPHORE MODEL AND QSAR STUDIES ON HERBICIDES TARGETING PHOTOSYSTEM II FROM CHLAMYDOMONAS REINHARDTII

Ana Borota, Simona Funar-Timofei, Luminita Crisan\* Department of Computational Chemistry, Institute of Chemistry of Romanian Academy, Timisoara, Mihai Viteazul Avenue, 24, 300223 Timisoara, Romania Iumi\_crisan@acad-icht.tm.edu.ro

Dedicated to the 150<sup>th</sup> anniversary of the Romanian Academy

✤ The resistance of weeds is a problem which can be overcome by finding new herbicides. For this purpose, beyond the experimental methods, in silico approaches can be helpful, as a starting point.

Abstract

✤ In this regard, pharmacophore mapping and 3D-QSAR studies were carried out on several series of herbicide, already known to act on the Photosystem II (PS II) D1 protein. Using PHASE software, three pharmacophore features, H-bond acceptor (A), hydrophobic (H) and aromatic ring (R) were taken into account to be the best hypothesis.

✤ For this hypothesis an atom-based 3D-QSAR model was generated with statistically significant parameters (the correlation coefficient of regression (R<sup>2</sup>) of 0.839, the standard error of estimates (SD) of 0.370, the Fisher test (F) of 53.7 for the training set, the external explained variance Q<sup>2</sup> = 0.640, the Pearson-R = 0.916 and Root Mean Square Error (RMSE) = 0.572, for the test set).

✤ This hypothesis, validated by the 3D atom-based QSAR approach, assures the selection of novel scaffolds of herbicide derivatives and can be used for the design of new chemical entities active on the PS II D1 protein.

 Our aim is to develop a valid pharmacophore model based on different scaffolds of PS II D1 herbicide derivatives (pyrimidine, pyridine, cinnoline, triazine and quinine) which further can be used for screening molecular databases in order to find potential new herbicides and for the prediction of their activity.

Aim

### Data set selection and processing

Material and Methods

- The datasets consisting of 58 inhibitors of the D1 protein in photosystem II (PSII D1) were collected from literature [5] and Pubchem database [6,7] AID1101260 and AID1101262.
- In case of ten compounds which show multiple experimental activities, we considered their average values.
- All structures were converted from smiles code into 3D structures, and ionization states and tautomers in the pH range of 6.2±0.3 were generated, using the LigPrep module [8] of Schrödinger suite [9].
- The conformational space for each ligand was developed with the help of ConfGen module [10,11] using the default options. 217 compounds resulted after conformer generation and energy minimization based on the OPLS-2005 force field.
- The pharmacophore hypotheses were generated using eight most active (with pIC50>7) compounds, while the threshold for inactivity was set to 5 using the Phase module [12-14] of Schrödinger suite [9].

## Material and Methods

Table 1. The structure of the most active compounds (1 to 8), the unaligned ligands (9 and 10) and the lessactive compounds (11 and 12) and their herbicidal activity in logarithmic units



fppt.com

### Pharmacophore modeling and validation

Material and Methods

➤ The "Develop Pharmacophore Model" module of Phase software [12-14] implemented in the Schrödinger suite was used in order to generate all possible pharmacophore hypothesis using four PLS factors. The number of PLS factors was increased, but the model statistics or predictive ability did not improve.

➤ The pharmacophore validation was carried out by atom-based 3D-QSAR regression including both internal and external validation. The training set includes 80% randomly selected molecules, whereas the remaining 20% were denominated to validate the model (test set). The external predictive ability for the test set prediction using Pearson-R was considered and the models which have values greater than 0.6 were selected.

➤Taking into account this statistical parameter but also high value of Q2 test (correlation coefficient of prediction for the test set) and R2 training (correlation coefficient for the training set) we selected the best QSAR model.

- ✓ Ten pharmacophore (Table 2) hypotheses based on different scaffolds of PSII D1 herbicide derivatives were generated using three minimum sites: H-bond acceptor (A), hydrophobic (H) and aromatic ring (R). The selected hypothesis AHR.7 (Figure 1) was used for the generation of the 3D QSAR model using four PLS factors. This model was built using the PHASE descriptors as independent variables and the herbicidal activity values (expressed as pIC50 values), as dependent variables. Two unaligned ligands (compound no 9 and no 10) of AHR.7 hypothesis, were excluded as outlier, see Table 1.
- ✓ A graphical representation of the significant favourable and unfavourable features for the herbicidal activity of the compounds that resulted when the QSAR model is applied is shows in Figures 3 to 6.



**Figure 1**. The pharmacophore hypothesis AHR.7 (acceptor (A1, pink), hydrophobic (H6, green), ring (R8, orange)) aligned to the compound **2** with best Fitness score = 3

**Figure 2**. The plot of observed versus predicted herbicidal activities for the model with the pharmacophore AHR.7 hypothesis



**#** Number of factors in the partial least squares regression model; **SD** - standard deviation of the regression;  $\mathbf{R}^2$  - the coefficient of determination; **F** - the ratio of the model variance to the observed activity variance; **P** - the significance level of variance ratio; **Stability** – the stability of the model predictions; **RMSE** – the root-mean-square error in the test set predictions;  $\mathbf{Q}^2$  - value for the predicted activities, analogous to  $\mathbf{R}^2$ , but based on the test set predictions; **r** (Pearson-R) - value for the correlation between the predicted and observed activity for the test set;

\*for the training set; <sup>\$</sup> for the entire data set; # for the test set.

#### Table 2. The statistical parameters obtained for the QSAR models

M	ode1	#	SD*	R2*	F*	P*	Stability <sup>\$</sup>	RMSE#	Q <sup>2#</sup>	1 <sup>#</sup>
		1	0.596			6.14E-07		0.627	0.455	0.710
AT	IR.3	2	0.452			4.37E-10		0.511	0.639	0.814
AL	цс.5	3	0.361	0.840	54.200	1.97E-12	0.364	0.510	0.640	0.802
		4	0.241	0.931	100.500	6.30E-17	0.237	0.470	0.694	0.834
	AHR.4	1	0.598			6.90E-07		0.578	0.538	0.778
ΔŦ		2	0.456			5.72E-10		0.559	0.568	0.757
	ш.т	3	0.372			5.13E-12	0.394	0.645	0.424	0.658
	-	4	0.261	0.919	84.700	6.63E-16	0.225	0.629	0.453	0.677
		1	0.644			9.91E-06		0.506	0.646	0.876
AHR.2	IR 2	2	0.386			1.78E-12		0.489	0.669	0.847
		3	0.325			4.46E-14	and the second se	0.594	0.512	0.728
		4	0.258			1.98E-16		0.570	0.550	0.756
		1	0.633			4.70E-08		0.652	0.532	0.767
AH	R.12	2	0.511			2.61E-11		0.590	0.617	0.817
		3	0.465			2.35E-12		0.644	0.544	0.779
		4	0.376			1.88E-15		0.636	0.555	0.808
		1	0.545			3.05E-08		0.495	0.694	0.842
AH	R.14	2	0.431			9.16E-11		0.629	0.507	0.722
		3	0.341		12.5	3.56E-13		0.567	0.599	0.786
		4	0.297			2.95E-14		0.552	0.620	0.795
		1	0.659			2.93E-07		0.657	0.525	0.742
AF	IR.7	2	0.507			1.82E-11		0.484	0.743	0.889
		3	0.428			7.18E-14		0.574	0.637	0.860
		4	0.370			9.03E-16		0.572	0.640	0.916
		1	0.681			1.27E-06		0.556	0.627	0.897
AHR.6	IR.6	2	0.505			1.52E-11		0.546	0.640	0.855
		5	0.433			1.16E-13		0.546	0.640	0.835
		4	0.337			2.07E-17		0.650	0.490	0.717
AHR.11		1	0.597			6.26E-07		0.642	0.430	0.661
	R.11	2	0.462			5.08E-10		0.581	0.532	0.734
		3			and the second s	2.19E-13		0.700	0.322	0.605
		4	0.276			7.26E-16		0.723	0.276	0.552
		1	0.634			5.71E-06		0.715	0.362	0.637 0.826
AF	IR.9	2	0.480			1.86E-09		0.547	0.627	
		1	0.389		1.000	8.29E-12 1.67E-16		0.717 0.662	0.360 0.453	0.657 0.718
		1				0.000122	and the second second			0.622
		2	0.752	Contraction of the second s				0.731	0.355	
AH	IR.5	2	0.585			8.16E-09		0.512	0.684	0.838
		4	0.487			1.65E-11		0.642	0.503	0.711
	-	4	0.389	0.822	47.400	7.47E-15	0.012	0.643	0.501	0.712

com.



**Figure 3**. The QSAR model visualized in the context of the best aligned compound (no **2**) with AHR.7



Figure 4. The QSAR model visualized in the context of the most active compound (no 3) of the test set



Figure 5. The QSAR model visualized in the context of the less active compound (no 11) of the training set



Figure 6. The QSAR model visualized in the context of the less active compound (no 12) of the test set

## Conclusion

- Pharmacophore-based 3D-QSAR study of PSII D1 inhibitors is carried out in order to explain the structural features of some herbicide derivatives (pyrimidine, pyridine, cinnoline, triazine and quinine) required for their inhibitory activity.
- The selected 3D-QSAR model indicates a significant correlation and a good predictive capacity. One hydrogen bond acceptors (A), one lipophilic/hydrophobic group (H) and one aromatic ring (R), as pharmacophore features, are important for the PSII D1 herbicidal activity. The best hypothesis AHR.7, in this study, is characterized by the best values of the R<sup>2</sup> regression coefficient (0.839) and the highest values for the Pearson-R coefficient (0.916).
- In future studies this pharmacophore model will be used for screening molecular databases in order to find potential new herbicides.

## Acknowledgements

This project was financially supported by Project 1.1 of the Institute of Chemistry of the Romanian Academy. The authors thank Dr. Ramona Curpăn (Institute of Chemistry Timisoara of Romanian Academy), for providing access to Schrödinger software acquired through the PN-II-RU-TE-2014-4-422 projects funded by CNCS-UEFISCDI Romania.

## References

1. J. Barber, Russian in Biokhimiya, **79** (2014) 248-262.

2. M.D. Lambreva, D. Russo, F. Polticelli, V. Scognamiglio, A. Antonacci, V. Zobnina, G. Campi, G. Rea, Curr. Protein Pept. Sci., **15 (**2014) 285-295.

3. http://herbicidesymptoms.ipm.ucanr.edu/MOA/Photosystem\_II\_Inhibitors (accessed August 2016).

4. M. Broser, C. Glöckner, A. Gabdulkhakov, A. Guskov, J. Buchta, J. Kern, F. Müh, H. Dau, W. Saenger, A. Zouni, J. Biol. Chem. **286** (2011) 15964–15972.

- 5. U. Egner, K.P. Gerbling, G.A. Hoyer, G. Kriiger, P. Wegnerb, Pestic. Sci., 47 (1996) 145–158.
- 6. Y.Wang, J. Xiao, T.O. Suzek, J. Zhang, J. Wang, Z. Zhou, L. Han, K. Karapetyan, S. Dracheva,
- B.A. Shoemaker, E. Bolton, A. Gindulyte, S.H. Bryant, Nucleic Acids Res., 40 (2012) D400-412.
- 7. PubChem BioAssay. https://pubchem.ncbi.nlm.nih.gov/ (accessed June 2016).
- 8. Schrödinger Release 2016-3: LigPrep, Schrödinger, LLC, New York, NY, 2016.
- 9. Small-Molecule Drug Discovery Suite 2016-3, Schrödinger, LLC, New York, NY, 2016.

10. K.S. Watts, P. Dalal, R.B. Murphy, W. Sherman, R.A. Friesner, J.C. Shelley, J.Chem. Inf. Model., **50** (2010) 534-546.

- 11. Schrödinger Release 2016-3: ConfGen, Schrödinger, LLC, New York, NY, 2016.
- 12. S.L. Dixon, A.M. Smondyrev, E.H. Knoll, S.N. Rao, D.E. Shaw, R.A. Friesner, J. Comput. Aided. Mol. Des. **20** (2006) 647-671.
- 13. S.L. Dixon, A.M. Smondyrev, S.N. Rao, Chem. Biol. Drug. Des. **67**(2006) 370-372.
- 14. Schrödinger Release 2016-3: Phase, Schrödinger, LLC, New York, NY, 2016.