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2 **Evaluating the Performance of Different Commercial and** 3 **Pre-commercial Maize Varieties Under Low Nitrogen** 4 **Conditions Using Affordable Phenotyping Tools**

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13 **Abstract:** Maize is the most commonly cultivated cereal in Africa in terms of land area and production. Low yields
14 in this region are very often associated with issues related to low Nitrogen (N), such as low soil fertility or low
15 fertilizer availability. Developing new maize varieties with high and reliable yields in actual field conditions using
16 traditional crop breeding techniques can be slow and costly. Remote sensing has become an important tool in the
17 modernization of field-based High Throughput Plant Phenotyping (HTPP), providing faster gains towards
18 improved yield potential, adaptation to abiotic (water stress, extreme temperatures, salinity) and biotic
19 (susceptibility to pests and diseases) limiting conditions, and even quality traits. We evaluated the performance
20 of a set of remote sensing indices derived from Red-Green-Blue (RGB) images and the performance of the field-
21 based Normalized Difference Vegetation Index (NDVI) and SPAD as phenotypic traits and crop monitoring tools
22 for assessing maize performance under managed low nitrogen conditions. Phenotyping measurements were
23 conducted on maize plants at two different levels: on the ground and from an airborne UAV (Unmanned Aerial
24 Vehicle) platform. For the RGB indices assessed at the ground level, the strongest correlations compared to yield
25 were observed with Hue, GGA (Greener Green Area) and GA (Green Area) at the ground level while GGA and
26 CSI (Crop Senescence Index) were better correlated with grain yield at the aerial level. Regarding the field sensors,
27 SPAD exhibited the closest correlation with grain yield, with a higher correlation when measured closer to
28 anthesis. Additionally, we evaluated how these different HTPP data contributed to the improvement of
29 multivariate estimations of crop yield in combination with traditional agronomic field data, such as ASI (Anthesis
30 Silking Data), AD (Anthesis Data), and Plant Height (PH). All multivariate regression models with an R^2 higher
31 than 0.50 included one or more of these three agronomic parameters as predictive parameters, but with RGB
32 indices at both levels increased to R^2 over 0.60. As such, this research suggests that traditional agronomic data
33 provide information related to grain yield in abiotic stress conditions, but that they may be potentially
34 supplemented by RGB indices from either ground or UAV phenotyping platforms. Finally, in comparison to the
35 same panel of maize varieties grown under optimal conditions, only 11% of the varieties that were in highest
36 yield producing quartile under optimal N conditions remained in the highest quartile when grown under
37 managed low N conditions, suggesting that specific breeding for low N tolerance can still produce gains, but that
38 low N productivity is also not necessarily exclusive of high productivity in optimal conditions.

39 **Keywords:** Maize, Low nitrogen, High Throughput Plant Phenotyping, Remote Sensing

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41 **1. Introduction**

42 Maize is the most commonly cultivated cereal in Africa in terms of land area and production [1]. Low yields
43 in this region are largely associated with drought stress, low soil fertility, weeds, pests, diseases, low input
44 availability, low input use and inappropriate seeds [2]. After water, nitrogen (N) is the single most important
45 input for maize production, and the lack of N is considered the principal constraint to cereal yields in areas with
46 more than 400 mm average annual rainfall in sub-Saharan Africa [3]. Plants scientists face the challenge of solving
47 these limitations while considering the additional implications of climate change on food security [4]. In that
48 sense, affordable technologies capable of monitoring crop performance, involve yield prediction, or assessing
49 phenotyping variability for agronomical or breeding purposes are aimed at surpassing the bottlenecks in the way
50 of full exploitation of this technology [5,6]. One of the first non-destructive and analytical tools was the
51 chlorophyll meter – based on radiation absorbance by leaves in the red and near-infrared regions (usually at 650
52 and 940 nm). As relative leaf chlorophyll content readings have an indirect and close relationship with leaf N
53 concentration and leaf chlorophyll concentration (SPAD) [7,8].

54 Remote sensing has become an important tool in the modernization of field High Throughput Plant
55 Phenotyping (HTPP), including improvements in yield potential, adaptation to abiotic stressors (drought,
56 extreme temperatures, salinity), biotic (susceptibility to pests and diseases) limiting conditions, and even quality
57 traits [6,9,10]. The Normalized Difference Vegetation Index (NDVI) [11] is one of the most well-known vegetation
58 indices derived from multispectral remote sensing, as it includes visible and near infrared radiation [12,13]. As a
59 low-cost alternative, various RGB-based Vegetation Indices (RGB-VIs) can be calculated from commercial Red
60 Green Blue (RGB) cameras that have proven able to predict grain yield, quantify nutrient deficiencies, and
61 measure disease impacts [14,15]. The RGB images can be processed using the Breedpix code that enables the
62 extraction of RGB-VIs in relation to different properties of color, which often demonstrate performance similar to
63 or slightly better than that of the better-known NDVI [16]. The RGB-VIs proposed here, namely Hue, Saturation,
64 Intensity, Green Area (GA) and Greener Green Area (GGA) (the last two are based on pixel selections of Hue of
65 60-180 and 80-180, respectively) and L, a*, and b* from the CIE-Lab color space, are readily obtainable from
66 zenithal pictures of canopies and by using the appropriate calculations [16].

67 The aim of this study is to evaluate the performance of different commercial and pre-commercial maize
68 varieties under low nitrogen conditions using affordable HTPP tools. We evaluated the selection of maize
69 varieties using a set of remote sensing indices derived from RGB images acquired from a UAV (Unmanned Aerial
70 Vehicle) and at the ground level compared with the performance of the field-based NDVI and SPAD sensors, and
71 then we tested their capacity for yield estimation both alone and in combination with standard agronomical
72 variables, such as ASI (Anthesis Silking Data), AD (Anthesis Data), and Plant Height (PH).

73 **2. Materials and methods**

74 *2.1. Plant material and growing conditions*

75 Field trials were conducted at the CIMMYT (International Center for Maize and Wheat Improvement)
76 regional station located in Harare, Zimbabwe (-17.800 S, 31.050 E, 1498 m.a.s.l.). The soil of the station is
77 characterized by a pH slightly below 6. This study consisted of two different conditions: the first was Optimum
78 Nitrogen (OP) with a standard fertilization application [10] and was the Low Managed Nitrogen (LOW) that was
79 25-35% less N fertilizer compared to the OP growing conditions. A set of 49 maize hybrids were developed at
80 CIMMYT and 15 commercial maize varieties in Zimbabwe. Seeds were sown during the wet season, on December
81 16th, 2015 and the harvested-on May 12th, 2016.

82 *2.2. Remote Sensing and proximal (ground) data collection*

83 Remote sensing evaluations were performed on seedlings (less than 5 leaves) during the last week of January.
84 RGB-VIs were evaluated for each plot at terrestrial and aerial levels. RGB aerial images were acquired using an
85 Unmanned Air Vehicle (UAV, Mikrokopter OktoXL, Moormerland, Germany) flying under remote control at
86 about 50 m. The digital camera used for aerial imaging was a Lumix GX7 (Panasonic, Osaka, Japan). Images were
87 taken at 16-megapixel resolution using a 4/3" sensor, 20mm focal length, 1/160 second shutter speed, and auto-
88 programmed aperture. These images were taken at the rate of every 2 s from 50 m for the duration of the flight. At

the ground, level, one conventional digital photograph was taken per plot with an Olympus OM-D (Olympus, Tokyo, Japan), holding the camera about 80 cm above the plant canopy in a zenith and focusing near the center of each plot. The images were acquired with a resolution of 16 megapixels with a 4/3" Live MOS sensor with a focal length of 14 mm, activated at a speed of 1/125 seconds with the aperture programmed in automatic mode.

The NDVI of individual plots at ground level was determined with a ground-based portable spectroradiometer with an active sensor (GreenSeeker handheld crop sensor, Trimble, USA). The RGB images at aerial and ground level were taken on January 28th, 2016 with the NDVI. SPAD was measured in two different dates, once on February 18th, 2016 (SPAD¹) and then again on March 1st, 2016 (SPAD²) using a portable Minolta SPAD-502 chlorophyll meter (Spectrum Technologies Inc., Plainfield, IL, USA).

2.3. Image processing and statistical analyses

For the RGB images, the Microsoft Image Composite Editor (ICE; Microsoft Research Computational Photography Group, Redmond, USA) was used to produce an accurate image mosaic as seen in Figure 1. A total of 63 overlapping images were used for mosaic. Through the open source image analysis platform FIJI (Fiji is Just ImageJ; <http://fiji.sc/Fiji>), regions of interest were established at each row for the plots to be cropped. RGB pictures were subsequently analyzed using a version of the Breedpix 0.2 software adapted to JAVA8 and integrated as the CIMMYT MaizeScanner plugin within FIJI (<https://github.com/George-haddad/CIMMYT>). With the Breedpix software code, the images were processed to convert RGB values into indices based on the models of Hue-Intensity-Saturation (HIS), CIE-Lab and CIE-Luv cylindrical-coordinate representations of colors. Additionally, Crop Senescence Index (CSI) was calculated in agreement with [15,17]. The Triangular Greenness Index (TGI) was calculated as the area of a triangle formed by the reflectance values of the Blue, Green and Red bands [18]. Finally, the Normalized Green Red Difference Index (NGRDI) is calculated as the difference between the green and red digital numbers differentiates between plants and soil, and the sum normalizes for variations in light intensity between different images [19]. All statistical analyses were done using R and R Studio (<http://cran.r-project.org>, <http://www.rstudio.com>, R Studio, Boston, USA).

3. Results and Discussion

3.1. The effect of optimal condition and low managed nitrogen on grain yield

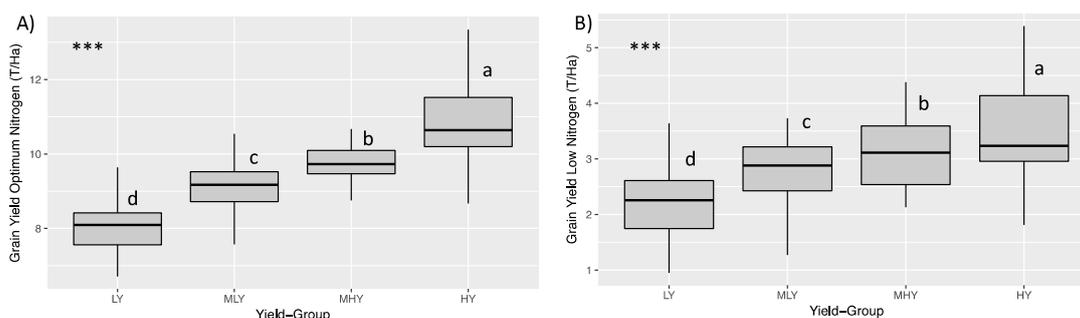


Figure 1. LY (Low Yield), MLY (Medium Low yield), MHY (Medium High Yield) and HY (High Yield) maize variety in two different conditions: (A) Optimum Nitrogen (OP) and (B) Low Nitrogen (LOW). Each value is the mean \pm SD for each genotype (n= 48 per quartile with 16 different variety). Bars with the different letters are significantly at $P < 0.001$.

The results showed (Figure 1) that maize varieties grown under optimal conditions, 11% of the varieties that were in highest yield producing quartile under optimal N conditions remained in the highest quartile when grown under managed low N conditions, suggesting that specific breeding for low N tolerance can still produce gains, but that low N productivity is not necessarily exclusive of high productivity in optimal conditions. In some cases, it has been reported that the genotypes selected under low N fertilization input are not truly adapted to N-

rich soils [20]. In [21] suggest that when the plant material performs relatively well under low N input, it should be selected under N deficiency conditions for which yield reduction does not exceed 35–40%.

3.2. Performance of remote sensing indices and field sensors in estimating grain yield

Table 1. Grain yield correlations with all proximal remote sensing variables from the RGB images taken from the UAV aerial platform, RGB images from the ground, and SPAD and NDVI field sensors. These indices are defined in the Introduction and Materials and Methods. Levels of significance: *, $P < 0.05$; ***, $P < 0.001$.

| RGB indices/ aerial | | | RGB indices/ ground | | | Additional Field Sensors | | |
|------------------------|--------|-----|------------------------|--------|-----|------------------------------|--------|--|
| | R | P | | R | P | | R | |
| GGA | 0.1978 | *** | GGA | 0.2339 | *** | SPAD ¹ (18/02/16) | 0.2936 | |
| GA | 0.1659 | *** | GA | 0.2175 | *** | SPAD ² (01/03/16) | 0.2564 | |
| Hue | 0.1449 | *** | Hue | 0.2351 | *** | NDVI | 0.1404 | |
| Intensity | 0.0932 | *** | Intensity | 0.0090 | | | | |
| Saturation | 0.1819 | *** | Saturation | 0.0515 | * | | | |
| Lightness | 0.0848 | *** | Lightness | 0.0208 | * | | | |
| a* | 0.1275 | *** | a* | 0.1467 | *** | | | |
| b* | 0.1573 | *** | b* | 0.0080 | | | | |
| u* | 0.1470 | *** | u* | 0.2021 | *** | | | |
| v* | 0.0884 | *** | v* | 0.0002 | | | | |
| CSI | 0.1830 | *** | CSI | 0.1031 | *** | | | |
| TGI | 0.0527 | * | TGI | 0.0019 | | | | |
| NGRDI | 0.1645 | *** | NGRDI | 0.0007 | | | | |

The RGB indices Hue, GGA and GA calculated from images taken at ground level demonstrated the best correlations with GY, outperforming other RGB indices (Table 1). GA quantifies the portion of green pixels to the total pixels of the image and is a reliable estimation of vegetation cover [22] and the values of GA in both observation levels were consistently below 60%. The ground and aerial measurements were taken at the same time on the same day, variation in environmental variables such as light intensity and brightness can be assumed to be negligible. On the other hand, all RGB indices from the ground and aerial levels didn't show significant differences between quartiles. This may be best explained considering that the data for our study were collected at an early phenological stage when the plants were not yet at full canopy cover and they didn't yet show the full range of symptoms of N deficiency. N deficiency can reduce plant growth rates, but also other later factors that affect GY, including leaf chlorophyll content, soluble protein content, photosynthetic rate and related enzyme activities of the maize plant during grain filling [23].

NDVI has been used with satisfactory results in many prediction models of yield in wheat at the field level [24], using field, airborne and satellite imagery. Regarding NDVI, it's the values clearly highlight and the variability is low, with more than 90% of values being in the range 0.55-0.8. These results support the previously reported saturation of reflectance spectra in the red and near-infrared regions, such that increasing leaf area does not involve a parallel increase in NDVI values [25]

SPAD is used to measure relative chlorophyll content in plant leaves and it has been effectively used to diagnose N status and predict GY potential in maize [26]. In maize, chlorophyll meters provide a convenient and reliable way to estimate leaf N content during vegetative growth [27] and over a large time scale after anthesis [28]. We can see this as a decline in relative chlorophyll content between the two SPAD measurements. This may be because when the crops are in the first phase (SPAD1), i.e., vegetative phase, young developing roots and leaves behave as sink organs for the assimilation of inorganic N and the synthesis of amino acids originating from the N taken up before flowering and then reduced via the nitrate assimilatory pathway. After flowering, (SPAD2), the N accumulated in the vegetative parts of the plant is remobilized and translocated to the grain [29].

In multivariate analyses, the estimation for yield using different combinations of RGB images from the field and UAV platforms, field sensors and traditional agronomical field measurements provided improved results over the single index results presented in Table 1. Combining RGB images and proximal field sensors resulted in

174 R² values of 0.403 and 0.384 for the ground and aerial RGB data, respectively. Further improvements were
175 observed when using also employing the traditional agronomical field measurements ASI, AD and PH, resulting
176 in R² values of 0.6157 and 0.6154 using RGB ground and aerial VIs, respectively. This suggests that the use of the
177 more time consuming field sensors may be replaced with either ground or aerial RGB data when used in
178 combination with the traditional agronomical field measurements for optimal results.

179 4. Conclusions

180 Maize hybrid technology may show promise for improving much-needed GY in low N environments and
181 the current range of variability in performance suggests the possibility of potential for further improvements. We
182 need to take advantage of known effects of low N on physiological processes to focus our efforts to bring HTPP
183 to low N breeding. For HTPP, RGB sensors can be considered as functional technology from the ground or a
184 UAV, but also, similar to SPAD, NDVI or any other agronomic or general plant physiological measurement, these
185 measurements must be carefully planned for an adequate growth stage in order to optimize their benefits to plant
186 breeding. Possible gains with new technologies with regards to equipment and time costs, especially in larger
187 breeding platforms.

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194 Regional Office in Harare, Zimbabwe. SCK carried out the UAV flights and image pre-processing for the obtainment of aerial
195 measurements. AG-R and JLA conducted the field measurements and the collection of samples. MLB processed the images,
196 analyzed the samples and wrote the paper under the supervision of JLA and SCK with the specific contributions from all of
197 the other authors.

198 **Conflicts of Interest:** The authors declare no conflict of interest.

199 **Abbreviations:** N: Nitrogen, NDVI: Normalized Difference Vegetation Index, HTPP: High Throughput Plant Phenotyping,
200 RGB: Red-Green-Blue, GA: Green Area, GGA: Green Greener Area, CIMMYT: International Maize and Wheat Improvement
201 Center, OP: Optimum Nitrogen, LOW: Low Managed Nitrogen, UAV: Unmanned Aerial Vehicle, ASI: Anthesis Silking Data,
202 AD: Anthesis Data, and PH: Plant Height, CSI: Crop Senescence Index, TGI: Triangular Greenness Index, NGRDI: Normalized
203 Green Red Difference Index, HIS: Hue-Intensity-Saturation, LY: Low Yield, MLY: Medium Low Yield, MHY: Medium High
204 Yield, HY: High Yield.

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