



SciForum MOL2NET

Phenolic Compounds Isolated From *Euphorbia phosphorea*

André Chaves de Albuquerque^{1,*}, Roseana Farias de Araújo Ramos², Lucas Silva Abreu², Yuri Manguiera do Nascimento², Josean Fachine Tavares^{1,2}, Vicente Carlos de Oliveira Costa²

¹ Department of Pharmaceutical Sciences, Health Sciences Center, Universidade Federal da Paraíba

² Postgraduate Program in Natural and Synthetic Bioactive Products, Health Sciences Center, Universidade Federal da Paraíba; E-Mail: josean@ltf.ufpb.br (J. F. T.)

* E-Mail: andrechaves05@gmail.com

Received: / Accepted: / Published:

Abstract: Euphorbiaceae is one of the largest families among angiosperms, being found in several distinct habitats and vegetation types, mainly in tropical and subtropical areas of America and Africa. Despite its notable economic importance, as well as its abundant occurrence in northeastern Brazil, few reports have been made about its metabolites and its pharmacological activities. The genus *Euphorbia* L. is globally distributed, but specially abundant in arid and semi-arid regions, where its occurrence is mostly as succulent xerophytes. As reported by previous studies, constituents from species of the genus *euphorbia*, such as tannins and flavonoids, have shown relevant activity in wound healing, conferring the genus therapeutical potential. This study conducted phytochemical investigations on the species *Euphorbia phosphorea*, obtained from the semi-arid region of Paraíba. Aerial parts of *E. phosphorea* were collected from Serra Branca, Paraíba, Brazil. The fresh material was dried in a circulating air oven, pulverized and macerated with ethanol, yielding an ethanol extract. The obtained extract was then partitioned with the organic solvents hexane, dichloromethane, ethyl acetate and n-butanol to obtain the respective soluble fractions. The ethyl acetate fraction was submitted to sephadex LH-20 column chromatography with methanol as eluent. The 88 sufractions obtained were then analysed in thin layer chromatography and gathered according to their retention factor similarity. ¹H NMR spectrum of fraction 47-51 revealed characteristics of polyphenolic compounds, being confirmed later by analysing its ¹³C-APT NMR and LC-MS data. Applying chromatographic and spectroscopic techniques, the study of *Euphorbia phosphorea* led to the obtaining of ellagic acid and corilagin, contributing to the phytochemical knowledge of the genus *Euphorbia*.

Keywords: Euphorbiaceae; *Euphorbia*; Tannins; Polyphenols.

1. Introduction

Euphorbiaceae is one of the largest families among angiosperms, being found in several distinct habitats and vegetation types, mainly in tropical and subtropical areas of America and Africa. Despite its notable economic importance, as well as its abundant occurrence in northeastern Brazil, few reports have been made about its metabolites and its pharmacological activities. The genus *Euphorbia* L. is globally distributed, but specially abundant in arid and semi-arid

2. Results and Discussion

Subfractions 47-51 obtained from CC showed the same profile on TLC, and therefore were gathered. The ^1H NMR spectrum of the subfractions 47-51 sample revealed a profile similar to polyphenols, and after comparison to literature data, was coded as "Ep-1".

On Ep-1 ^1H NMR spectrum, four singlets were observed in the region of aromatic hydrogens, in δ_{H} 7.43, 7.03, 6.56 and 6.49, suggesting that Ep-1 was a phenolic compound. The singlet in δ_{H} 7.43, integrating for two hydrogens, suggested the presence of ellagic acid. Other singlets observed in the ^1H NMR spectrum were attributed to the galloil (δ_{H} 7.03) and hexahydroxydiphenol (δ_{H} 6.56 and 6.49) groups. Characteristic signals of an osidic unit were observed in the region of δ_{H} 3.7 to 4.6, as well as a characteristic doublet of anomeric hydrogen in δ_{H} 6.20, associated to one unit of glucose. Therefore, the signals also suggested the presence of tannin on the sample.

The ^{13}C -APT NMR spectrum (100 MHz, DMSO-d 6) confirmed the presence of the ellagic acid in the sample by characteristic shifts at δ_{C}

3. Materials and Methods

Aerial parts of *E. phosphorea* were collected from Serra Branca, Paraíba, Brazil. The botanical identification was carried out by Doctor José Iranildo Miranda de Melo from Department of Biology of Universidade Estadual da Paraíba (UEPB).

Column chromatography (CC) was performed using sephadex LH-20 as stationary phase and methanol as mobile phase. Thin layer chromatography (TLC) was performed on silica gel GF₂₅₄. ^1H and ^{13}C -APT NMR spectra were acquired with a Bruker AscendTM 400 (^1H , 400

regions, where its occurrence is mostly as succulent xerophytes. As reported by previous studies, constituents from species of the genus *euphorbia*, such as tannins and flavonoids, have shown relevant activity in wound healing, conferring the genus therapeutical potential. This study conducted phytochemical investigations on the species *Euphorbia phosphorea*, obtained from the semi-arid region of Paraíba.

159.4, attributed to carbonyls and at δ_{C} 140.4 and 148.4 assigned to aromatic oxygenated carbons (C-2 / C-7 and C-3 / C-8, respectively). The spectrum of Ep-1 also revealed the presence of characteristic carbonyl ester signals at δ_{C} 164.9, 166.9 and 167.3, as well as oxygenated aromatic carbons in δ_{C} 144.1 to 145.7. Chemical shifts in 62.3 to 77.6 reinforced the presence of the galloyl and hexahydroxydiphenolic groups and the glucose unit present in the tannin structure. By comparing ^1H and ^{13}C NMR data with those described in the literature, it was possible to suggest that the other substance in the sample was 1-O-galloyl-3,6- (R) -hexydroxydiphenyl- β -glucopyranose (corilagine).

Analyzing the data obtained by low resolution HPLC-ESI-MS operating in the negative mode, it was possible to observe peaks with retention time in approximately 26.2 min and 38.9 min, corresponding respectively to ions with m/z 633 $[\text{M}-\text{H}]^-$ for corilagine and m/z 301 $[\text{M}-\text{H}]^-$ for ellagic acid, confirming the presence of these compounds in the sample.

MHz; ^{13}C , 100 MHz) spectrometer. Chemical shift (δ) values are given in ppm, with TMS as internal standard. HPLC-ESI-MS was performed with a Shimadzu Prominence 20A system consisting of a CBM-20A controller, LC-20 AD parallel-flow pumps, a DGU-20A degasser and a SIL-20A auto sampler. The HPLC system was coupled to an AmazonX Ion-Trap mass spectrometer (Bruker Daltonics, Bremen, Germany). The ESI-MS acquisition parameters were capillary voltage 4.5 kV, negative ESI mode; end-plate offset 500 V; nebulizer gas 35

psi; dry gas (N₂) with a flow rate of 8.0 L · min⁻¹ and temperature of 300 °C.

The fresh material was dried in a circulating air oven, pulverized and macerated with ethanol, yielding an ethanol extract. The obtained extract was then partitioned with the organic solvents hexane, dichloromethane, ethyl acetate and n-butanol to obtain the respective soluble fractions.

The ethyl acetate fraction was submitted to sephadex LH-20 column chromatography with methanol as eluent. The 88 subfractions obtained were then analyzed in thin layer chromatography and gathered according to their retention factor similarity.

4. Conclusions

Applying chromatographic and spectroscopic techniques, the phytochemical study of *Euphorbia phosphorea* led to the obtaining of ellagic acid and corilagin, contributing to the phytochemical knowledge of the genus *Euphorbia*.

Acknowledgments

The authors would like to thank to CAPES, CNPq, FINEP, IPeFarM and UFPB..

Conflicts of Interest

The authors declare no conflict of interest.

References

1. JAHAN, N.; KHALIL-UR-REHMAN; ALI, S.; BAHATTI, I. A. Antimicrobial potential of gemmo-modified extracts of *Terminalia arjuna* and *Euphorbia tirucalli*. **International Journal of Agriculture and Biology**, v. 13, n. 6, p. 1001-1005, 2011.
2. LUO, W.; ZHAO, M.; YANG, B.; SHEN, G.; RAO, G. Identification of bioactive compounds in *Phyllanthus emblica* L. fruit and their free radical scavenging activities. **Food Chemistry**, v. 114, n. 2, p. 499-504, 2009.
3. NAWWAR, M. A. M.; HUSSEIN, S. A. M.; MERFORT, I. NMR spectral analysis of polyphenols from *Punica granatum*. **Phytochemistry**, v. 36, n. 3, p. 793-798, 1994.
4. SHI, Q-W.; SU, X-H.; KIYOTA, H. Chemical and pharmacological research of the plants in genus *Euphorbia*. **Chemical reviews**, v. 108, n. 10, p. 4295-4327, 2008.
5. TRINDADE, M. J. de S.; LAMEIRA, O, A. Espécies úteis da família Euphorbiaceae no Brasil. **Revista Cubana de Plantas Medicinales**, v. 19, n. 1, p. 292-309, 2014.
6. VASAS, A.; HOHMANN, J. *Euphorbia* diterpenes: isolation, structure, biological activity, and synthesis (2008–2012). **Chemical Reviews**, v. 114, n. 17, p. 8579-8612, 2014.
7. WU, Q-C.; TANG, Y-P.; DING, A.; YOU, F.; ZHANG, L.; DUAN, J. C-NMR data three important diterpenes isolated from *Euphorbia* species. **Molecules**, v. 14, p. 4454-4475, 2009.
8. ZHU, M.; DONG, X.; GUO, M.; Phenolic profiling of *Duchesnea indica* combining macroporous resin chromatography (MRC) with HPLC-ESI-MS/MS and ESI/IT-MS. **Molecules**, v. 20, n. 12, p. 2246311475, 2015.
9. Goyal, M., Nagori, B. P., Sasmal, D. 2012. Wound healing activity of latex of *Euphorbia caducifolia*. **Journal of ethnopharmacology**, 144, 3, 786-790, 2012.