Second iCBSM 2017

Understanding the covalent inhibition of Clavulanate against β -lactamases (TEM-1 and KPC-2) with QM/MM screening methods.

<u>Rubén Fritz^{1*}</u>, Jans Alzate-Morales¹, Marc Van der Kamp², Jim Spencer³ and Adrian Mulholland²

¹Doctorado en Ciencias Aplicadas, Centro de Bioinformática y Simulación Molecular (CBSM), Facultad de Ingeniería, Universidad de Talca, Talca, Chile. ²Centre for Computational Chemistry, University of Bristol, Bristol, UK.

³Department of Molecular Bioscience, School of Cellular and Molecular Medicine, University of Bristol, Bristol, UK.

* rfritz@utalca.cl

Abstract

β-lactamases are a primary cause of bacterial resistance to β-lactam antibiotics for many important human pathogens (particularly Gram-negative bacteria) [1]. Inhibitors of β-lactamase have been implemented as a dual therapy with antibiotics, but there are only four inhibitors clinically approved and resistance to these compounds is also rising [2]. For β-lactam inhibitors, after acylation, the opening of five-membered ring leads to the formation of a transient imine intermediate then it rearranges several times to form a trans or cis final enamine inhibition-products. Slow hydrolysis of this product by the enzyme leads to an inhibited β-lactamase [2]. A computational study of reaction mechanism for the first step on the deacylation of the inhibitor clavulanate with TEM-1 (inhibited) and KPC-2 (hydrolyzed) enzymes using QM/MM Umbrella Sampling with DFTB method is presented [3]. 2D free energies surfaces for the reactions were calculated using the weighted histogram analysis method (WHAM) and the minimum energy path (MEP) was identified; where the highest point along the MEP is taken as the transition state giving the activation free energy " ΔG_{calc} ". Our results show that TEM-1 and KPC-2 have an approximate 5 kcal/mol difference in ΔG_{calc} . Such results are in good agreement with inhibition experimental data for two enzymes in which KPC-2 is less inhibited by clavulanate than TEM-1. We hope our methodology can assist the design and development of covalent inhibitors through a computational screening of inhibitory activity of other molecules.



Irreversible Inhibition

Irreversible Inhibition

KPC-2	0.027	19.72	18.76 ± 0.6
TEM-1	0.0017	21.36	23.85 ± 1.25
ΔΔG		-1.65	-5.08

Methodology



QM/MM MD Umbrella Sampling.

1) Complexes model creation (KPC-2 y TEM-1). 2) Adduct parametrization. 3) Molecular Dynamics (MD). 4) QM/MM MD, 6 replicas of 50 ps. 5) US of reaction coordinates. 5.1) Distance. $d_1 = CC(Ser70) - O(DW).$ $d_2 = O(Glu 166) - H(DW)$. $d_3 = H(DW) - O(DW).$ 5.2) Reaction coordinates definition. $R_x = (d_2 - d_3)$. Proton Transfer. $R_y = d_1$. Nucleophilic Attack. 5.3) Initial coordinates for Minimun Energy Path from (MEP) of Meropenem. 5.4) 2ps QM/MM MD simulation per windows, 0.1 Å . 5.5) Scan of the surface: R_v from 1.4 to4.0 Å. R_x from -0.8 to 1.1 Å. 5.6) 3 Umbrella Sampling QM/MM MD replicas. 6) Free Energy Estimation.

Potential of Mean Force (PMF) with WHAM.
 Estimation of Transition State (TS) and "ΔG_{calc}".



Conclusiones.

The activation free energies calculated for TEDC with TEM-1 and KPC-2 are in the same magnitud order as the experimental values for the inactivation constant.

These computational methodology aloud us to discriminate the inhibitory activity of CLA against KPC-2 and TEM-1 β -lactamase.

Perspectives.

Calculate others covalent complexes during CLA inhibition.

These protocol could potentially be use as a *In-silico* screening for inhibitory activity of covalent inhibitors.

Expansion to others protein-ligands systems of medical interest.

Acknowledgments.

R.F. thanks The Royal Society of Chemistry (RSC) for economic support through Researcher Mobility Grant (R. Fritz 16 Round 1). J.A-M. and R.F. thanks financial support from project FONDECYT No. 1140618.

References:

[1] Hermann, J. C., Pradon, J., Harvey, J. N., & Mulholland, A. J. *J. Phys. Chem. A* (2009), *113*(43), 11984–11994.
[2] Drawz, S. M., Papp-Wallace, K. M., & Bonomo, R. a. *Antimicrob. Agents Chemother* (2014), *58*(4), 1835–1846.
[3] Chudyk, E. I., Limb, M. a L., Jones, C., Spencer, J., van der Kamp, M. W., & Mulholland, A. J. *Chem Commun*, (2014), *50*, 14736–14739.

y Simulación Molecular CHILE	