



## Qualitative phenolic profile of *Rumex tunetanus* flowers and stems using ultra-high-performance liquid chromatography coupled to quadrupole-time-of-flight mass spectrometry and their antioxidant activity

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*Received: / Accepted: / Published:*

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### Abstract:

The present study was designed to investigate the bioactive compound in *Rumextunetanus* extracts (polygonaceae), a plant growing in GarâaSejnane region (NW-Tunisia). Hydro-methanol extracts of flowers and stems of *Rumextunetanus* were analyzed by RP-UHPLC-ESI-QTOF-MS in the negative mode to identify the maximum of bioactive compounds. Applied the aforementioned method, a total of 60 bioactive compounds were characterized for the first time in *Rumextunetanus* between them, 18 photochemical were firstly identified in the Polygonaceae family in negative ionization mode. Quantification of the identified compounds revealed that quercetin-3-O-glucuronide and (-)-epicatechingallate were the most abundant phenolic compounds in flowers and stems, respectively. Moreover, positive correlations were found between the antioxidant activity measured by DPPH and FRAP assays with the total phenolic compounds ( $r = 0.98$ ;  $r = 0.99$ , respectively) and the abundance of some phenolic subfamilies such as hydroxycinnamic acids, hydroxybenzoic acids, flavonols and flavones with  $r > 0.86$ . The compounds displaying significant ( $P < 0.01$ ) and good correlations with the antioxidant activity ( $r > 0.93$ ) were hydroxybenzoic acid, rutin, quercetin-3-O-glucuronide, quercetin-3-O-glucoside, quercetin and luteolin-7-O-rutinoside. In addition, the flowers and stems of *Rumex tunetanus* showed different bioactive compound profiles and significant antioxidant properties of extracts. These results highlight the potential of the RP-UHPLC-ESI-QTOF-MS and MS/MS system to identify untargeted metabolic profiling of *Rumex tunetanus*. Overall, these results contribute to the clear explanation of the past and current usage of genus *Rumex* in folk medicine. Future investigations are necessary to develop purified antioxidant extracts, with the application of more selective extraction techniques.

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**Keywords:** *Rumex tunetanus*, Phenolic compounds, Metabolic profiling, Antioxidant activity, RP-UHPLC-ESI-QTOF-MS.

## 1. Introduction

In response to the increased popularity and greater demand for medicinal plants, a recent study by world health organization claimed 80% dependency of world population on ethnomedicines[1].

In fact, the importance of medicinal plants in solving the health care problems of the world is gaining increasing attention and it is growing phenomenally at the international level. However, the pharmacological evaluation of substances from plants is an established method for the identification of new compounds which can leads to the development of novel and safe medicinal agents in order to increase the dangerous side effects of synthetic molecules [2].The genus *Rumex* represents one of the most important genera of Polygonaceae family, including approximately 200 species widely distributed in North American, European, African and Ausian countries [3].Since very old times, differents species of *rumex* genus have been used for relief of symptoms of diseases. Much interest in *Rumex* species emanates from their long use in folk medicines as well as their pharmacological properties pharmacological activities such as anti-inflammatory, antidiuretic, antitumor, analgesic, antifungal and anti-viral activities [1].Large number of medicinal plants have been investigated for their antioxidant properties .Polyphenols are considered among the most important antioxidants in humain diet, and their presence in plant can protect consumers against oxidative

stress , cardiovascular and chronic diseases [4].In fact, *Rumex* genus emerged as a good source of natural antioxidants. The major types of phenolic antioxidants found in *Rumex* genus include phenolic acids and their derivatives, namely gallic acid, dihydroxybenzoic acid, hydroxybenzoic acid and vanillic acid, and flavonoids, namely flavan-3-ols(catechin, epicatechin and epicatechin gallate), flavonols (rutin, quercetin-3-O-glucoside, quercetin-3-O-glucuronide, quercetin and isoramnetin), as well as condensed tannins as B-type procyanidin dimer and A-type procyanidin trimer[5]. Tunisia flora is known for its diversity of medicinal plant among them *Rumex tunetanus*. This plant was growing in the wet marshes of the Sejenane plain. *Rumex tunetanus* was never seen after its discovery in 1888, and more than 120 years after the discovery of this *rumex*, we find a large population of *Rumex tunetanus* in december 2009[6]. Therefore, as potential bioactive markers, the total phenolic content (TPC) and antioxidant capacity of flowers and stems of *rumex tunetanus* were firstly evaluated. Secondly, their phenolic profiles were extensively studied by ultra-high performance liquid chromatography (UHPLC) coupled with two detection systems, DAD and quadrupole time-of-flight (QTOF)-MS using electrospray ionization in negative ionization mode.

## 2. Results and Discussion

### Total phenolic content (TPC) and antioxidant activity of the *Rumex tunetanus* flowers and stems

The determination of TPC of the extracts was performed by the Folin-Ciocalteu assay and calculated as gallic acid equivalents and is listed in Table 1. *Rumex tunetanus* flowers extract was found to contain higher total phenolic contents (146.20 mg GAE/g extract) as compared to *Rumex tunetanus* stems extract (118.88 mg GAE/g extract).

The extracts were evaluated for their antioxidant /radical scavenging activity by DPPH and FRAP. The results of percentage scavenging are depicted in Table 1. In general, both of these two analyses showed that the hydro-methanolic extract of *Rumex tunetanus* exhibited good antioxidant activities. According to the aforementioned results for TPC, the flowers showed higher antioxidant activity values than stems by the two assayed methods. In fact, our results have indicated that the DPPH value of *Rumex tunetanus* flowers and stems extracts are in accordance with those obtained by Yousef et al. [11] for *Rumex tuberosus* flowers (0,65mg/ml of extract) and stems (0,69 mg/ml of extract).

### Characterization strategy and fragmentation pattern study

The metabolic profiling of the aqueous-methanolic extracts of flowers and stems was performed by using RP-HPLC-DAD-QTOF-MS using ESI in the negative mode. This is the commonest ionization source and mode used to identify the phenolic and non-phenolic compounds in *Rumex tunetanus* flowers and stems. Using this methodology, our

characterization steps could be basically summarized by a targeted searching of previously, an untargeted analysis and a predictive study of unreported phenolic structures based on all the spectrometric data obtained by the detection techniques applied (Figure 1).

Thanks to the methodology proposed in the present study, a total of 60 metabolites (60 in flowers and 51 in stems (Table 2) have been tentatively identified, including sugars (1), organic acid (3), hydroxybenzoic acids (8), hydroxycinnamic acids (1), flavonoids (28) as well as tannins (19). These results exemplify that RP-HPLC-DAD-QTOF-MS is useful to detect and characterize novel chemical structures.

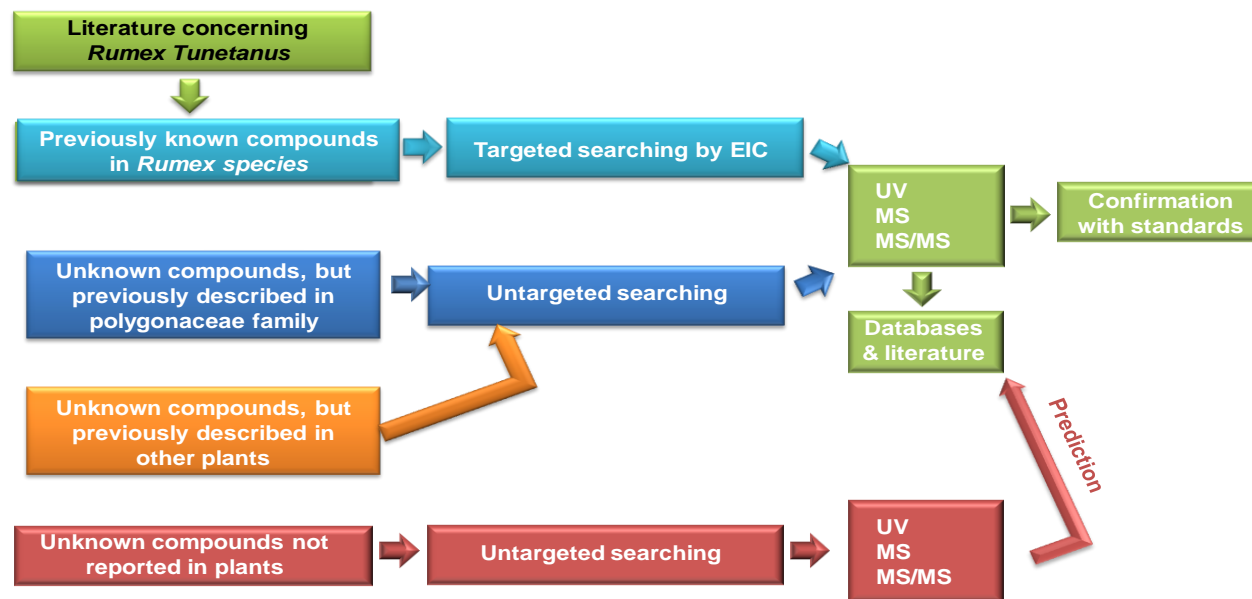
### Comparison between flowers and stems

Table 1 shows the qualitative differences between flowers and stems: presence (+) or absence (-) of the characterized compounds. In brief, more than 90% of the compounds were in common in both extracts. Moreover, as summary Figure 2 depicts these differences in terms of number of compounds found in each phenolic class. Interestingly, flavonoids and their glycoside derivatives were characterized in the negative ionization mode as a more widely spread phenolic group of *Rumex tunetanus* samples with 28 compounds. To the best of our knowledge, this is the first time that dihydroxybenzoic acid hexoside (8), syringic acid hexoside (12) and tri-*O*-methoxyellagic

acid (**51**) have been identified in Polygonaceae family.

**Table 1** Total phenolic content (TPC) (mg of GAE/g of the extract) determination and antioxidant capacities of the studied *Rumex tunetanus* flowers and stems parts as measured by DPPH (mg/ml of extract) and FRAP (mmol eq. FeSO<sub>4</sub>/g of the extract) antioxidant assays

Extract	TPC	DPPH	FRAP
Flowers	146.2 ± 1.86	0.72 ± 0.230	528.24 ± 32.36
Stems	118 ± 1.05	0.68 ± 0.230	307.04 ± 54.36



**Figure 1:** The strategy for deducing of the most possible structures of *Rumex tunetanus* bioactive compounds

Table 2. Compounds characterized by HPLC-DAD-QTOF-MS in *Rumex tunetanus* flowers and stems

N°	RT <sup>a</sup> (min)	Exp. <sup>a</sup> m/z for [M-H] <sup>-</sup>	Proposed compound	Presence in		N°	RT <sup>a</sup> (min)	Exp. <sup>a</sup> m/z for [M- H] <sup>-</sup>	Proposed compound	Presence in			
				Flowers	stems					Flowers	Stems		
<b>Sugars</b>													
1	1.52	341.1116	Sucrose	+	+	58	36.81	329.0673	Tricin II	+	+		
						60	39.70	329.0643	Tricin III	+	+		
<b>Organic Acid</b>													
2	1.59	191.0542	Quinic acid I*	+	+	<b>Flavonoids- flavanones</b>		50	30.69	287.0562	Eriodictyol*	+	-
4	1.82	191.0538	Quinic acid II	+	+	55	34.39	271.0605	Naringenin*	+	+		
5	1.97	191.0183	Citric acid*	+	+	<b>Flavonoids- flavonols</b>					+	+	
<b>Phenolic acid (Hydroxycinnamic acid and derivatives )</b>													
19	17.94	355.1028	Ferulic acid hexoside	+	+	16	17.03	289.0727	(+)-Catechin* <sup>c</sup>	+	+		
						23	19.53	289.0727	(-)-Epicatechin* <sup>c</sup>	+	+		
<b>Phenolic acid (Hydroxybenzoic acids and derivatives)</b>													
7	3.75	169.0137	Gallic acid*	+	+	47	27.37	287.0556	Fustin <sup>b</sup>	+	+		
8	4.43	315.0725	Dihydroxybenzoic acid hexoside <sup>b</sup>	+	-	<b>Flavonoids- isoflavones</b>							
9	8.88	153.0186	Dihydroxybenzoic acid hexoside <sup>b</sup>	+	+	57	36.57	299.0568	7-Methoxy 2'-hydroxy genistein (cajanin) <sup>b</sup>	+	+		
10	12.14	137.024	Hydroxybenzoic acid I*	+	+	59	37.38	359.078	5.7.4 -trihydroxy-6.3.5- trimethoxyisoflavone (Irigenin) <sup>b</sup>	+	+		
11	12.96	137.0236	Hydroxybenzoic acid II	+	+	<b>Flavonoids- glucoside</b>					+	+	
12	13.94	359.0976	Syringic acid hexoside <sup>b</sup>	+	+	46	27.25	591.1357	Flavonoid glucoside- HMG conjugate <sup>b</sup>	+	+		
25	20.43	167.0347	Vanillic acid*	+	-	<b>Condensed tannins and derivatives</b>					+	+	
51	31.38	343.0429	Tri-O-methoxyellagic acid <sup>b</sup>	+	+	13	15.82	577.1334	B-type procyanidin dimer (I) <sup>c</sup>	+	+		
<b>Flavonoids- flavonols</b>													
						14	16.34	577.1334	B-type procyanidin dimer (II) <sup>c</sup>	+	-		

31	23.39	615.0990	Quercetin-O-galloyl-hexosideb	+	+	22	18.71	577.1349	B-type procyanidin dimer (III)c	+	+	
32	23.60	609.1465	Quercetin-3-O-rutinoside (rutin)*c	+	+	24	19.81	577.136	B-type procyanidin dimer (IV)c	+	-	
34	24.11	463.0881	Quercetin-3-O-glucoside I*c	+	+	26	20.48	729.1472	B-type procyanidin dimer gallate I	+	-	
35	24.33	477.0656	Quercetin-3-O-glucuronide*	+	+	27	20.72	863.1813	A-type procyanidin trimer	+	+	
36	24.44	463.0884	Quercetin-3-O-glucoside Iic	+	-	28	21.09	729.1465	B-type procyanidin dimer gallate II	+	-	
39	25.31	607.1306	Quercetin-3-[6''-(3-hydroxy-3-methylglutaryl)] $\beta$ -hexosideb	+	+	29	21.47	577.1363	B-type procyanidin dimer (V)c	+	+	
40	25.56	599.1050	Quercetin dihydroxybenzoylhexoside b	+	+	30	21.63	729.1455	B-type procyanidin dimer gallate III	+	+	
43	26.44	505.099	Quercetin-3-O-hexosyl-6''-acetate	+	-	37	24.8	729.1464	B-type procyanidin dimer gallate IV	+	-	
45	27.71	625.1183	Quercetin-O-dihexoside	+	+	44	26.61	729.1473	B-type procyanidin dimer gallate V	+	+	
49	29.67	639.1339	Quercetin-O-feruloylhexosideb	+	+	48	28.13	881.1566	B-type procyanidin dimer digallateb	+	+	
52	31.88		Quercetin*c	+	+	<b>Hydrolysable tannins and derivatives</b>				+	+	
53	32.46	345.0608	Quercetin-3,6-dimethyl etherb	+	+	3	1.70	331.0648	Galloyl glucose I	+	+	
54	32.71	315.0514	Isorhamnetin*	+	+	6	2.72	331.0661	Galloyl glucose II	+	+	
<b>Flavonoids-flavones</b>						15	16.97	483.0816	Di-O-galloyl-glucose I	+	+	
	38	25.12	593.1532	Luteolin-7-O-rutinoside (Scolymoside)*b	+	+	17	17.63	453.1036	Hydroxy-methoxyphenyl-O-(O-galloyl)-hexose I <sup>b</sup>	+	+
	41	26.00	447.0943	Luteolin-7-O-glucoside (Cynaroside)*c	+	+	18	17.80	483.1122	Hydroxy-dimethoxyphenolgalloyl-glucose <sup>b</sup>	+	+
	42	26.18	329.0671	Tricin I	+	+	20	17.94	453.1031	Hydroxy-methoxyphenyl-O-(O-galloyl)-hexose II <sup>b</sup>	+	+
	56	35.75	285.0405	Luteolin*c	+	+	21	18.35	483.0763	Di-O-galloyl-glucose II	+	+

<sup>b</sup>Compounds described here for first time in family Polygonaceae. Several saccharide combinations and conjugation positions are reported in different plant families (see KNApSack, Reaxys or SciFinder databases)

<sup>c</sup>compounds were previously reported in several Rumex species.

<sup>a</sup>RT, retention time;; Exp, experimental

\*Identification confirmed by comparison with standards

### 3. Materials and Methods

#### Solvents and standards

The solvents used for extraction were ultrapure water and methanol. All solvents used for HPLC-MS analysis were delivered by J.T Baker (Phillipsburg -USA). The reagents used to measure the TPC (total phenol content) and the antioxidant capacity were: Folin-Ciocalteu; sodium carbonate ( $\text{Na}_2\text{CO}_3$ ); 2,2-diphenyl ,1-picrylhydrazyl (DPPH); 2,4,6-tri(2-pyridyl)-1,3,5-triazine (TPTZ) and ferric chloride ( $\text{FeCl}_3 \cdot 6\text{H}_2\text{O}$ ); gallic acid, acetic acid ( $\text{C}_2\text{H}_4\text{O}_2$ ); Ferric sulfate ( $\text{FeSO}_4$ ); hydrochloric acid (HCl); trihydrate sodium acetate ( $\text{C}_2\text{H}_3\text{NaO}_2 \cdot 3\text{H}_2\text{O}$ ); sodium acetate; ascorbic acid. The phenolic standards used in our work were bought from Sigma-Aldrich (Saint-Louis -Missouri). The degree of purity of the standards was around 95% (w/w).

#### Plant samples procurement and extraction

Flowers, stems and leaves from the *Rumex tunetanus* were collected from the Garâa Sejenane region (NW–Tunisia) in May 2014. The samples was harvested and transferred to the laboratory where they dried in the dark at room temperature 30 °C, and then they were finely ground prior to extraction.

Flowers and stems were extracted using methanol/water 80:20 (v/v) as described elsewhere [7]. The residue were filtered with a syringe filter (regenerated cellulose, 0.45µm pore size) and stored at –20 °C until future analysis. The extractions were repeated twice for each studied *rumex tunetanus* part.

#### Total phenol content and antioxidant capacity assays

The TPC of the extracts was determined in triplicate by the colorimetric assay using the Folin–Ciocalteu reagent as reported by Ammar et al.[8]. The DPPH assay was based on the method described by Gargouri et al.[9]. The FRAP assay was conducted following the method described by Ammar et al. [10].

#### Mass spectrometry-based analyses

The analyses were made with an Agilent 1200 series rapid resolution (Palo Alto, CA, USA). The system was coupled with a DAD and a 6540 Agilent ultra-high-definition (UHD) accurate-mass Q-TOF LC/MS, which was equipped with Agilent Dual Jet Stream electrospray ionization (Dual AJS ESI) interface. The detail of negative ionization modes was described by Abidi et al [7].

#### Databases

In addition of consulting literature, the following databases were used to retrieve chemical information: PubChem (<http://pubchem.ncbi.nlm.nih.gov>), ChemSpider (<http://www.chemspider.com>), SciFinder Scholar (<https://scifinder.cas.org>), Reaxys (<http://www.reaxys.com>), Phenol-Explorer ([www.phenol-explorer.eu](http://www.phenol-explorer.eu)) and KNApSAcK Core System ([http://kanaya.naist.jp/knapsack\\_jsp/top.html](http://kanaya.naist.jp/knapsack_jsp/top.html)). MassBank and Metlin Metabolite Database were used to check fragmentation patterns in some cases.



#### 4. Conclusions

Globally, these results exemplify the usefulness of RP-HPLC-DAD-QTOF-MS to perform characterization studies, such as that described here. In this way, 60 phenolic compounds were characterized in flowers and 51 in stems of *rumex tunetanus* which were classified into twelve compound families that possess potent antioxidant activities. Flavonols, flavanols, flavones, hydroxybenzoic acids and condensed tannins were the most representative groups in both samples by using the relative amounts of the phenolic compounds. In addition, the antioxidant properties of extracts were evaluated in terms of single electron transfer as DPPH and FRAP assay and *Rumex tunetanus* flowers exhibited significantly higher antioxidant activities than the stems. Besides, the isolation of pure compounds with pharmacological activities holds significance in contemporary and future research. Recently, the plant extract was also being used by the researchers to produce nanoparticles [2]. Therefore, there is huge room for research in these directions.

#### Conflicts of Interest

The authors declare no conflict of interest.

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