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Introduction

Inflammation of the mammary gland in cattle is known as mastitis, it is one of the most prevalent and expensive diseases of dairy herds worldwide, and antibiotic therapy is the main tool for its treatment (Hogeveen et al., 2011). *Staphylococcus aureus* is responsible for most cases of subclinical mastitis in dairy cows (Mestorino and Errecalde, 2012). This microorganism is characterized by the ability to develop resistance against various traditional antimicrobials, which represents a serious problem in the dairy production chain (Basanisi et al., 2017). Therefore, research for the development of new drugs that fight this disease is of utmost importance. Plants are promising sources of new biological agents with antibacterial action, in addition to having the advantage of not inducing resistance even after prolonged exposure (Gomes et al., 2016; Domadia et al., 2007). Strawberry (*Fragaria x ananassa*) is a berry with an important amount of metabolites, mainly anthocyanins, phenolic compounds and flavonoids (Basu et al., 2016; Aaby et al., 2007; Giamperi et al., 2012); The beneficial potential that these compounds present in strawberry fruit have on human, animal and plant health has been demonstrated (Giamperi et al., 2015; Giamperi et al., 2013; Nohynek, et al., 2006; Puupponen et al., 2001). However, the strawberry is a non-climacteric fruit, with a short shelf life, so much of what is harvested that does not meet the characteristics of export quality (Extra quality), is destined for the national fresh market (first quality), to industrialization (first and / or second quality) or to waste. In this sense, the objective of this work was to evaluate the antimicrobial activity of 3 quality strawberry fruits against multidrug-resistant *S. aureus* caused by bovine mastitis, as a proposal for a viable alternative to add value to the strawberry that does not reach quality of export (first and second quality).

Materials and methods

Biological material

The multi-resistant strains of *S. aureus* AMC 9* and AMC 23** were isolated from milk samples from cows with mastitis in the Ciénege de Chapala region, Michoacán, Mexico, and strain ATCC 27543 as reference. For the extraction of anthocyanins, strawberry fruits (*F. x ananassa*) of export quality (MEX Extra) (Driscoll's®), first quality (MEX1) and second quality (MEX2) purchased at the local market of Sahuayo, Michoacán were used. The fruits were lyophilized and stored in a dark desiccator at room temperature (23 ± 5 °C) until processing.

*AMC 9: Resistant to ampicillin, cephalothin, cefotaxime, cefuroxime, dicloxacillin, erythromycin, cefepime, gentamicin, penicillin.

**AMC 23: Resistant to ampicillin, cefotaxime, cefuroxime, erythromycin, cefepime, penicillin, trimethoprim-sulfamethoxazole, tetracycline.

Anthocyanin extraction from strawberry fruits

The technique described by Abdel-Aal and Hucl (1999) was used, which consisted of taking 1 g of sample and 5 mL of acidified ethanol (ethanol and 1N HCl; 85:15 v / v) and it was macerated in a mortar. Subsequently, the solutions were vigorously stirred in Vortex and the pH was adjusted to 1 with 1N hydrochloric acid. Then, the solutions were stirred (LSE® orbital shaker) at 250 rpm for 16 h, at room temperature. After the time, the solutions were centrifuged at 6,000 rpm for 15 min and the supernatant was recovered, which was titrated to 25 mL with acidified ethanol. The concentration of cyanidine 3-glycoside (Cy3G) was determined by colorimetry according to the technique described by the author. The samples were stored at -20 °C until use.

Minimum Inhibitory Concentration (MIC) and Bactericidal (MBC)

The MIC of the total extracts was determined for the pathogenic strains evaluated, using the microdilution method with some modifications (Seleshe et al., 2017). In 96-well microplates with 100 µl of Mueller-Hinton broth (Sigma-Aldrich®), 20 µl of bacterial inoculum adjusted to 10⁸ CFU ml⁻¹ was added. The extracts evaluated were serially diluted starting from a concentration of 100 µg Cy3G ml⁻¹, to obtain 10 dilutions (100, 50, 25, 12.5, 6.3, 3.2, 1.5, 0.8, 0.4 and 0.2 µg ml⁻¹) and 50 were added µl to each well, respectively. The microplates were incubated at 37 °C with constant agitation at 150 rpm for 16-18 h. Sterility control (Mueller-Hinton broth), growth control (broth + inoculum) and antibiotic control (Trimethoprim-Sulfamethoxazol®, 10 µg / ml) were included. After incubation, the turbidity of the samples was measured to determine the MIC by spectrophotometry (BioTek PowerWave HT, USA). Additionally, 10 µl of aqueous tetrazolium bromide solution (MTT) (3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide, 1 mg ml⁻¹, Sigma-Aldrich®) was added to the wells as a growth indicator and then incubated for 2 h. The color change to blue-purple indicates bacterial growth. The lowest concentration of the wells without coloration was considered the minimum inhibitory concentration. For the determination of the MBC, 100 µl of each well without color change was plated on Mueller-Hinton agar (Sigma-Aldrich®) and incubated at 37 °C for 24 hours. The lowest concentration that did not produce growth after this subculture was considered the CMB. All trials were carried out in triplicate.

Statistical analysis

Each experiment was carried out in triplicate. An analysis of variance and the comparison of Tukey means was performed using the SAS statistical software (version 9.0, Cary, NC, USA) and a difference of p < 0.05 was considered significant.

Results and Discussion

The concentration of cyanidine 3-glycoside (mg eq 100 g⁻¹ fresh fruit (ff)) of the anthocyanin extracts of strawberry fruits of 3 qualities was determined, the results are shown in Table 1. The differences in the concentration of anthocyanins in the quality of the fruit evaluated are significant, being the first quality strawberry extract (MEX1) which had the highest concentration of the cyanidin 3-glycoside anthocyanin pigment, followed by the second quality fruits (MEX2) and finally the extra quality fruits (MEX Extra). The concentrations shown by the strawberry fruits evaluated agree with Karaaslan and Yaman (2017), which obtained values of 40.2 ± 0.5 mg cyanidine 3-glycoside 100 g⁻¹. Likewise, the results are consistent with those reported by the USDA (2019), with values up to 22 mg Cy3G 100 g⁻¹.

These differences in the amount of anthocyanins between strawberry fruit quality evaluated can be explained by the degree of ripeness of the fruit at the time of processing. According to Williner (2003), the highest anthocyanin content is observed in fully ripe strawberry fruits, while some phenolic compounds such as ellagic acid decrease their concentration.

Table 1. Cyanidine 3-glycoside (mg 100 g⁻¹) concentration of anthocyanin of strawberry fruit extracts of 3 different quality.

Strawberry fruit quality	Cyanidine 3-glycoside concentration (mg 100 g ⁻¹)
MEX Extra	23.7 ± 1.28 c
MEX 1	43.9 ± 0.5 a
MEX 2	31.1 ± 0.53 b

The values indicate the average of 3 replicas ± standard deviation. Different letters indicate significant differences between treatments (n = 3, p ≤ 0.05).

The 3 types of strawberry showed an obvious inhibitory potential against all the pathogens evaluated (MIC, Table 2). Anthocyanin extracts of second quality strawberry fruits (MEX2) showed greater inhibitory capacity against all strains (6.3 to 12.5 µg ml⁻¹) compared to extra and first quality fruits (MEX Extra and MEX1), in which the MIC was 25 µg ml⁻¹ for all strains, with the exception of the reference strain ATCC 27543, which obtained inhibition at a concentration of 12.5 µg ml⁻¹ of the first quality strawberry extract (MEX1). Regarding the Minimum Bactericidal Concentrations (MBC) (Table 2), the results showed differences between the 3 qualities of fruits compared to the strains evaluated. Again, all the strains evaluated were more susceptible to second quality or waste strawberry extract (MEX2) with CMB values of 25 µg ml⁻¹, followed by extra quality strawberry extract (MEX Extra) with values of 50 µg ml⁻¹; and, finally, the strawberry extract of first quality (MEX1), which obtained CMB of 100 µg ml⁻¹ with the exception of the multi-resistant strain AMC 23, with CMB value of 50 µg ml⁻¹.

Table 2. Minimal Inhibitory Concentrations (MIC) and Minimal Bactericidal Concentrations (MBC) of strawberry fruit extracts of 3 different quality against multiresistant *S. aureus* bovine mastitis isolates.

Strain	Strawberry fruit quality			
	MEX Extra	MEX 1	MEX 2	
ATCC 27543	MIC	25 ± 0.52 b	12.5 ± 0.52 a	12.5 ± 0.41 a
	MBC	50	100	25
AMC 9	MIC	25 ± 0.41 b	25 ± 0.52 b	6.3 ± 0.55 a
	MBC	50	100	25
AMC 23	MIC	25 ± 0.98 b	25 ± 1.05 b	12.5 ± 0.75 a
	MBC	50	50	25

MIC and MBC expressed in µg ml⁻¹. The values indicate the average of 3 replicas ± standard deviation. Different letters indicate significant differences between treatments (n = 3, p ≤ 0.05).

Anthocyanins have also been the focus of various investigations, mainly for their antioxidant activity (Viskeli et al., 2009; Giamperi et al., 2012; Lila, 2004). However, the antimicrobial activity of anthocyanin pigments present in various fruits, flowers and plants has gained vital interest in the search for antibacterial compounds. Pertuzzatti et al. (2016) observed the potential antimicrobial activity of cranberry extracts (*Vaccinium ashei*) from different cultivars against *E. coli* (WCC, 20-35 mg ml⁻¹; CMB, 30-45 mg ml⁻¹) and attribute to the anthocyanins found (cyanidine, petunidine, malvidin and delphinidine, in its glycosylated form) this action. They attribute this fact to the partial hydrophobicity of the phenolic compounds and anthocyanins present, which allow destabilizing the cell wall of lipopolysaccharides in the case of Gram (-) bacteria. Hafidh et al. (2011) found antimicrobial activity of anthocyanin extracts of red cabbage (*Brassica oleracea*) against clinical isolates of methicillin-resistant *S. aureus* (MRSA) with MIC values 10 to 40 times higher than those found in the present investigation. The author mentions that the mixture of phenolic compounds present in extracts of purple cabbage has high antioxidant activity, which acts as a protective agent for cells. This synergistic effect of the phenolic compounds, in particular of the mixture of anthocyanins present, can explain the antimicrobial potential, suggesting as a mechanism of action the alteration in cell permeability, causing deformation in its structure and damage in functionality.

Conclusions

The multiresistant strains of *S. aureus* that cause bovine mastitis are susceptible to strawberry anthocyanin extracts, in addition to a quality-dependent effect of the fruit evaluated, the second quality strawberry fruit extract (MEX2) being observed, which has greater inhibitory and bactericidal activity.

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