

Patrícia Calado^{1,2}, Catarina Dias^{1,2}, Ana M. Matos^{1,2,3}, Maria T. Blásquez-Sánchez^{1,2}, Philipp Dätwyler⁴, Beat Ernst⁴, M. Paula Macedo³, Amélia P. Rauter^{1,2,*}

¹Centro de Química e Bioquímica, Faculdade de Ciências, Universidade de Lisboa, Ed. C8, Campo Grande, 1749-016 Lisboa, Portugal

²Centro de Química Estrutural, Faculdade de Ciências, Universidade de Lisboa, Ed. C8, Campo Grande, 1749-016 Lisboa, Portugal

³Centro de Estudos de Doenças Crónicas (CEDOC), Chronic Disease Research Center, NOVA Medical School/Faculdade de Ciências Médicas, Universidade Nova de Lisboa, Lisboa, Portugal

⁴Department of Pharmaceutical Sciences, Pharmazentrum, University of Basel, Klingelbergstrasse 50, CH-4056 Basel, Switzerland

The state of the art:

Stilbenes, particularly resveratrol, are amongst the best documented and emblematic neuroprotective natural products. In addition to the well-known antioxidant and anti-inflammatory activities, this compound inhibits A β oligomeric cytotoxicity and reduces neuronal cell death [1, 2]. On the other hand, rosmarinic acid, the active principle of the neuroprotective plant *Salvia sclareoides*, prevents amyloid aggregation, reducing also a number of other events underlying AD pathology [3-5]. Interestingly, methyl caffeate itself, a sub structural unit of rosmarinic acid, reduces significantly A β oligomeric cytotoxicity and promotes disaggregation of A β oligomers [5].

The problem:

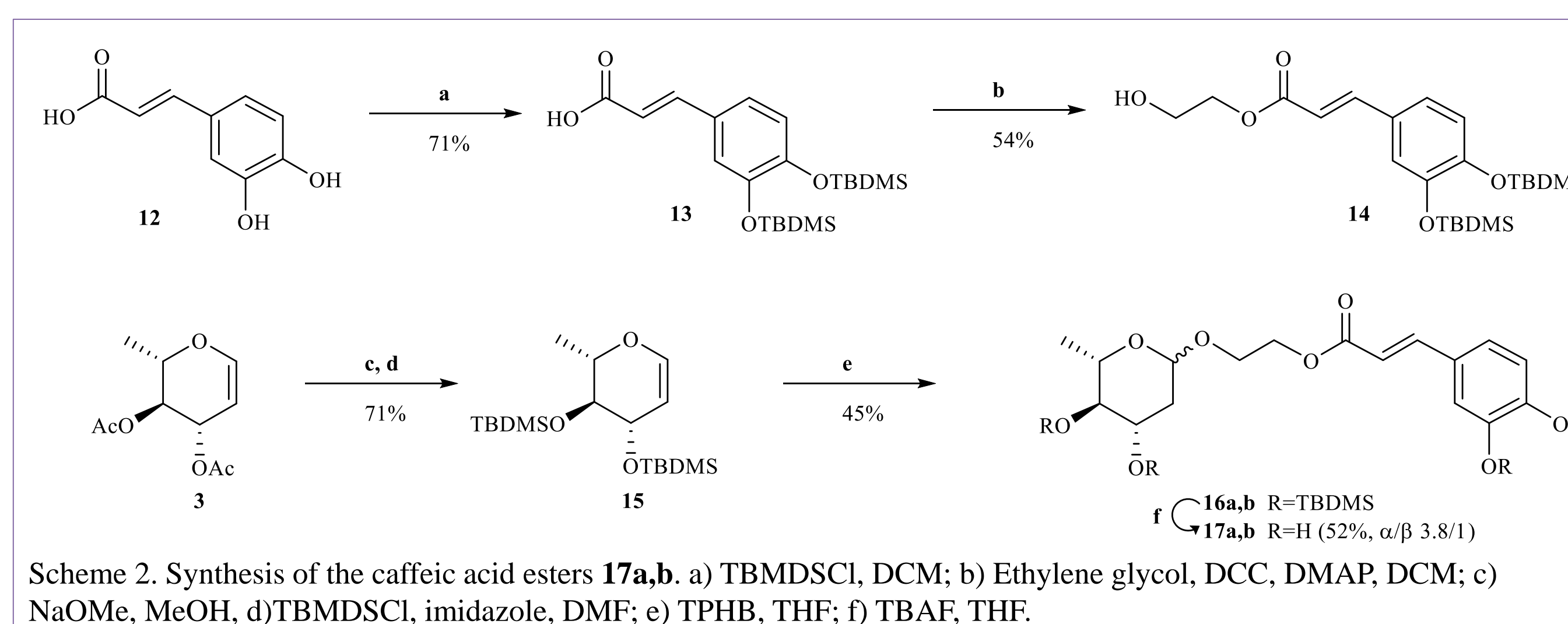
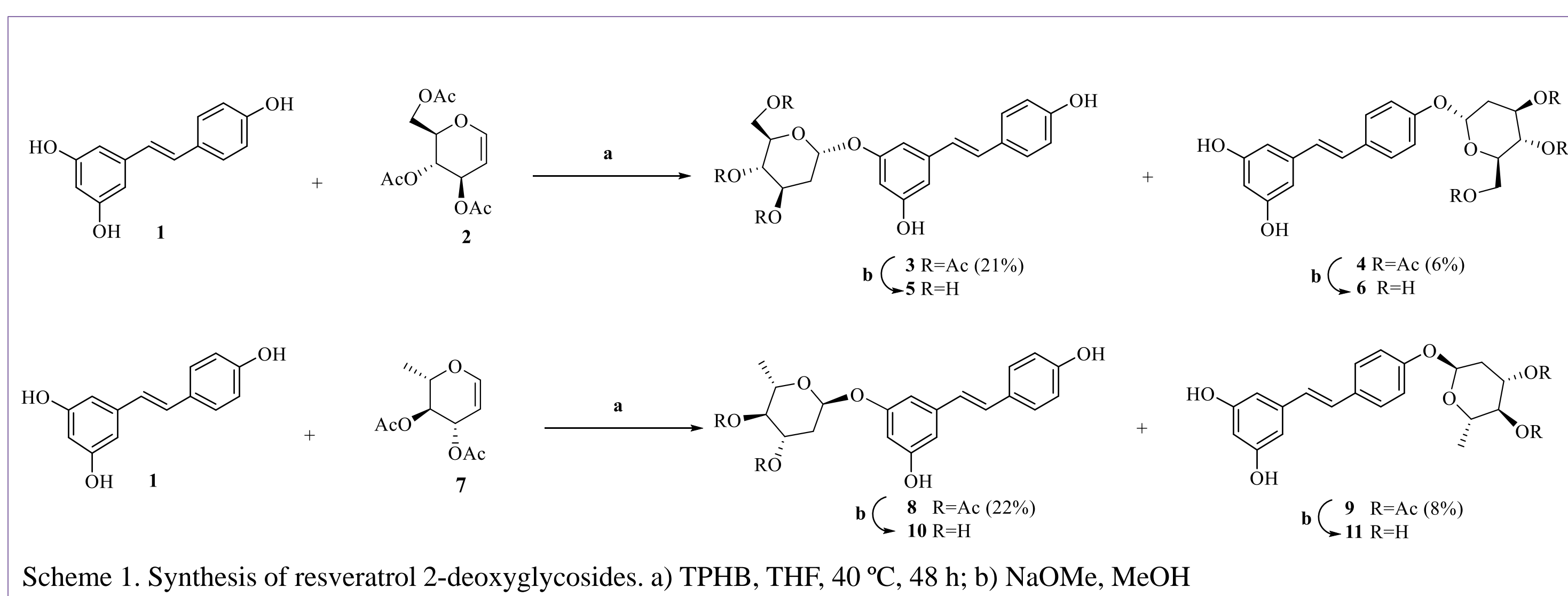
Resveratrol bioavailability in humans is less than 1% [6, 7]. Also, methyl caffeate has low bioavailability [5]. These compounds' water-insolubility limits their further pharmacological exploitation.

The solution:

These findings encouraged us to investigate neuroprotective principles' glycosylation, using glycals as glycosyl donors to afford new resveratrol and caffeic acid ester 2-deoxyglycosides, envisioning more effective and bioavailable compounds.

The synthesis:

Resveratrol 3-*O*- and 4'-*O*-glycosides **3**, **4**, **8** and **9** were prepared by reaction of resveratrol with the appropriate acetyl protected glycal (**2** and **7**, respectively), in the presence of triphenylphosphane hydrobromide (TPHB), a catalyst known to afford almost exclusively 2-deoxyglycosylation instead of the well known Ferrier products, obtained starting from acylglycals with Lewis and Brønsted catalysts. Separation of 3-*O*- and 4'-*O*-glycosides followed by Zemplén deprotection afforded glycosides **5**, **6**, **10** and **11** in 21%, 6%, 22% and 8%, respectively. On the other hand, for the synthesis of the caffeic acid ester, the aromatic hydroxy groups of caffeic acid were protected with a *tert*-butyldimethylsilyl (TBDMS) group, followed by Steglich esterification. With TPDPS or TMS protection, no further developments were achieved. Coupling of **15** with **14** was once again accessed with TPHB as catalyst, to give **17a/b** as an anomeric mixture α/β in ratio 3:1 in 52% yield.



The neuroprotective assays and drugability evaluation:

Hydrogen peroxide overproduction causing oxidative stress in neuroblastoma cells (SH-SY5Y) was used to access the neuroprotective effects of glycosides **5**, **6**, **10**, **11** and **17**.

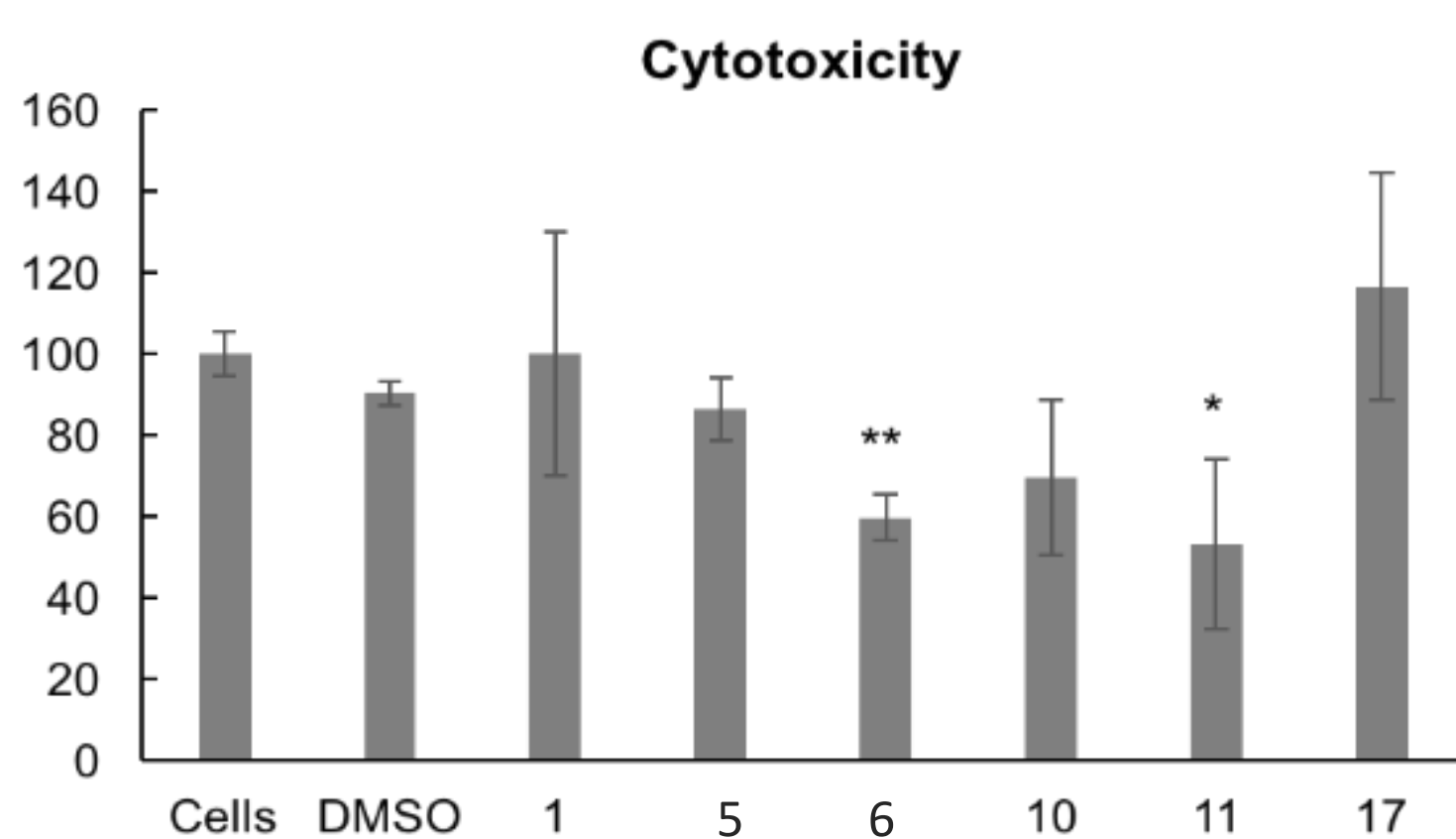
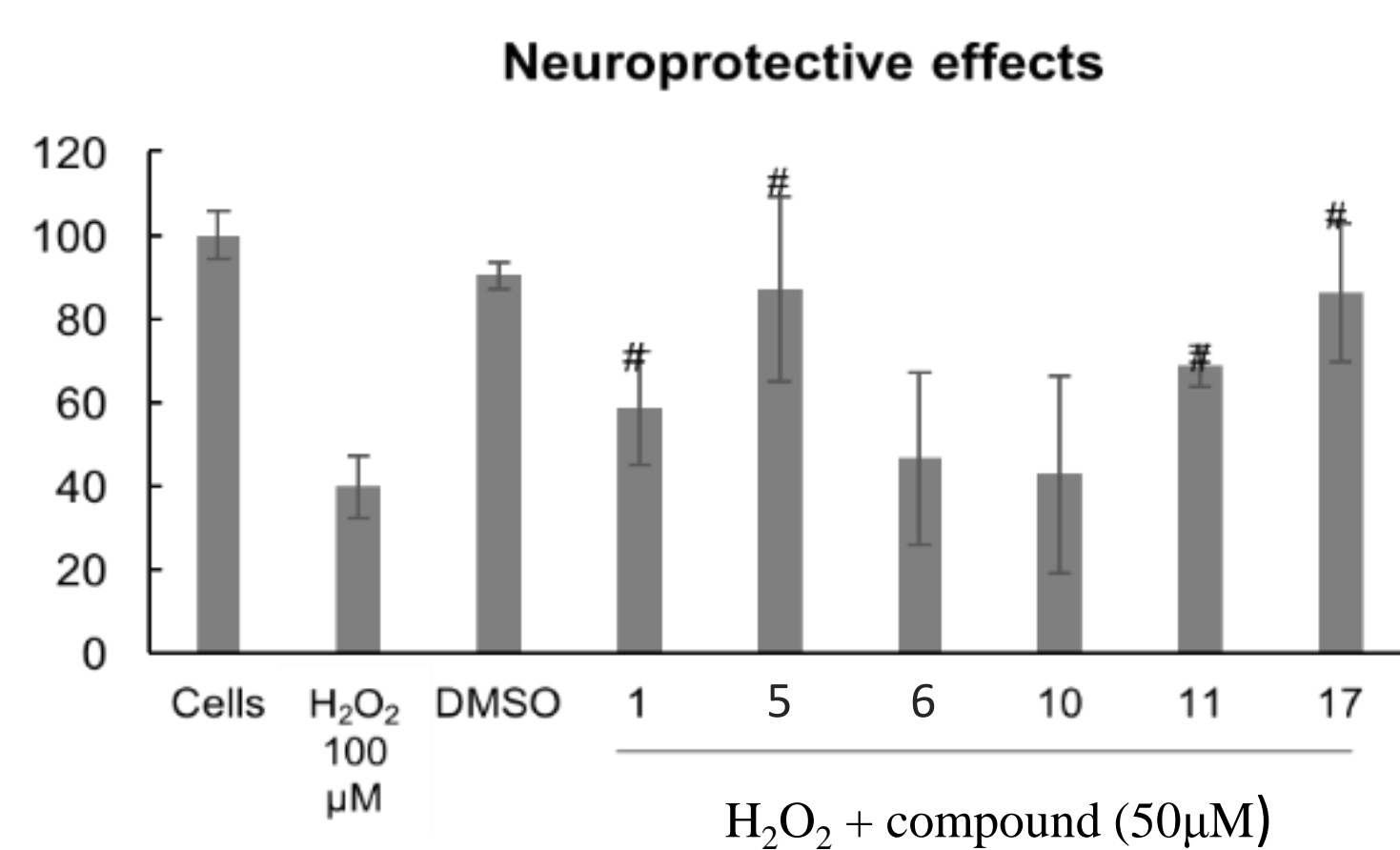


Figure 1. Neuroprotective and cytotoxic effects of synthesized compounds in neuroblastoma (SH-SY5Y) cells. * = significantly different when compared to cells control (p-value < 0.05); # = significantly different when compared to hydrogen peroxide controls (p-value < 0.05). Results are expressed as the mean \pm SEM of at least three independent experiments.

Incubation of cells with 100 μ M of H₂O₂ led to a cell viability decrease of ca. 60%. Incubating the cells with both hydrogen peroxide (100 μ M) and resveratrol glycosides **5** and **11** resulted in a statistically significant increase in the percentage of viable cells (figure 1). Caffeic acid ester **17** showed the best neuroprotection activity, maintaining cellular viability similar to that of the control. By accessing glycosides' toxicity in neuroblastoma cells, only compounds **6** and **11** led to a statistically different loss of viability at 50 μ M, remaining above 50% in all cases. The toxicity of the two most promising compounds was also assessed in Caco-2 and in HepG2 cell lines, and they were not toxic at all concentrations tested (0.1 - 100 μ M). When evaluating the drugability of the glycosides, Log D_{7.4} values indicated a moderate lipophilicity, essential to their bioavailability and blood-brain barrier penetration. Indeed, Log D values close to 2 have been established as ideal for BBB penetration [8, 9]. The best permeability results were shown by 2,6-dideoxy-*arabino*-hexopyranosides **10** and **11** (L-series).

Table 1. Physicochemical properties and intestinal wall permeability of lead candidates.

Compound	HBA	HBD	log D _{7.4}	PAMPA (-log Pe)/cms ⁻¹
1	0	3	n.d. ^[a]	3.97 \pm 0.02
5	2	5	1.748 \pm 0.074	10.00 \pm 0.00
6	2	5	1.747 \pm 0.097	6.73 \pm 0.18
10	2	4	2.013 \pm 0.061	5.56 \pm 0.13
11	2	4	2.813 \pm 0.112	4.82 \pm 0.02
17 a/b	3	4	n.d. ^[a]	10.00 \pm 0.00

^[a]Not detected due to poor ionization; HBA - Hydrogen bond acceptor; HBD - Hydrogen bond donor; logD_{7.4} - distribution coefficient at pH 7.4.

Conclusion:

Our results show a good improvement in compounds' efficiency. Indeed, resveratrol glycosides **5** and **11** were more effective at protecting neuronal cells from peroxide-induced cytotoxicity than resveratrol itself. In addition, caffeic acid ester **17** showed the best neuroprotection activity. Coefficient partition measurements demonstrated the moderate lipophilicity of resveratrol glycosides, with log D values typical of CNS drug and ideal for BBB penetration, while passive permeation assessed by PAMPA revealed that our glycosides from the L series were more effective to permeate the intestinal barrier than our glycosides from the D series.

Acknowledgements:

The European Union for the support of the project entitled "Diagnostic and Drug Discovery Initiative for Alzheimer's Diseases" (D3i4AD), FP7-PEOPLE-2013-IAPP, GA 612347

References:

- [1] S. D. Rege, T. Geetha, G. D. Griffin, T. L. Broderick, J. R. Babu. *Front. Aging Neurosci.* **6**, 218 (2014); [2] C. Rivière, T. Richard, L. Quentin, S. Krisa, J. M. Ménilon, and J. P. Monti. *Bioorg. Med. Chem.* **15**, 1160 (2007); [3] T. Alkam, T. Nitta, A. Mizoguchi, H. Itoh, A. Nabeshima, T. Behav. *Brain Res.* **180**(2), 139 (2007); [4] C. Airoldi, E. Sironi, C. Dias, F. Marcelo, A. Martins, A. P. Rauter, F. Nicotra, J. Jimenez-Barbero, *Chem. Asian J.* **8**, 596 (2013); [5] T. Walle. *Ann. NY Acad. Sci.* **1215**, 9 (2011); [6] E. Wenzel, V. Somoza. *Mol. Nutr. Food Res.* **49**, 472 (2005); [7] F. Marcelo, C. Dias, A. Martins, P. J. Madeira, J. Jorge, M. H. Florêncio, F. J. Cañada, Eurico J. Cabrita, J. Jiménez-Barbero, A. P. Rauter. *Chem. Eur. J.* **19**(21), 6641 (2013), [8] L. Di, E. H. Kerns, Chapter 5 - Lipophilicity, In *Drug-Like Properties* (2nd Edition), Academic Press, Boston, pp.39-50 (2016); [9] Z. Rankovic, *J. Med. Chem.* **58** (6), 2584 (2015).



5th International Electronic Conference
 on Medicinal Chemistry
 1-30 November 2019

