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A BODIPY-based fluorescent sensor for amino acids bearing thiol

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A BODIPY-based fluorescent sensor for amino acids bearing thiol

1. Introduction
2. Objective
3. Molecular Design
4. Synthesis
5. Photophysics
6. Sensing
7. Conclusions

Sensing of biomolecules

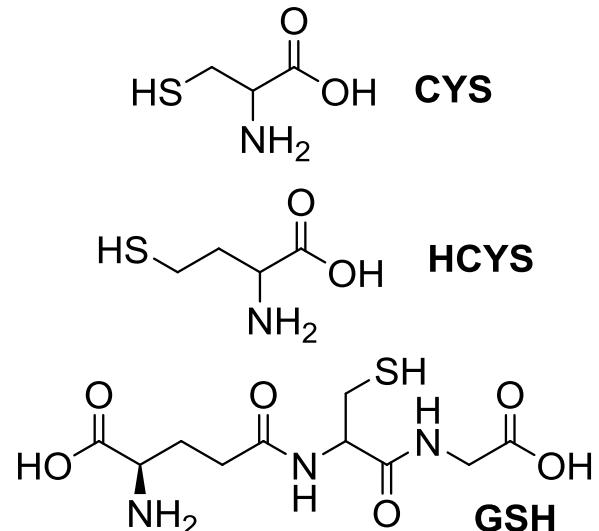
Visualization of biochemical events

Early detection of diseases

Unravel mechanism of physiological anomalies

One of the most seeking targets are **amino acids** (AA) like those bearing thiols:

Cysteine (CYS), Homocysteine (HCYS) and Glutathione (GSH)



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Fluorescent chemosensors

Ligh emission as a tool to transform chemical information (AA presence or concentration) into analitical information (fluorescence signal)

Design:



High affinity for the target analyte

Induce spectroscopic changes into the dye photophyscs

Upon target binding, reverse the aforementioned change into the spectroscopic signatures

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ON-OFF switches

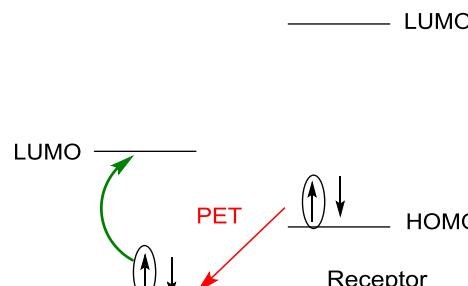
OFF state



ON state

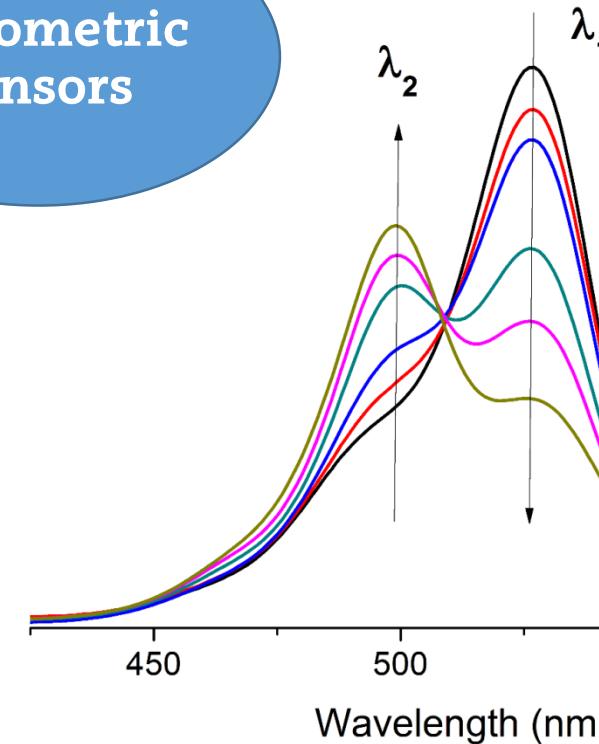


Receptor induced photoinduced electron transfer (PET)



Target binding disallows PET

Ratiometric sensors

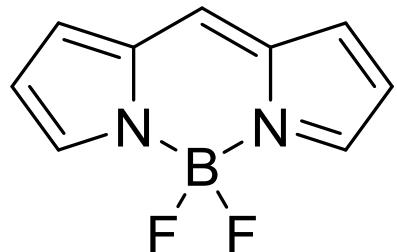


Binding of the target promotes spectral splitting
Different channels for detection (signal loss at λ_1
or signal growing at λ_2)

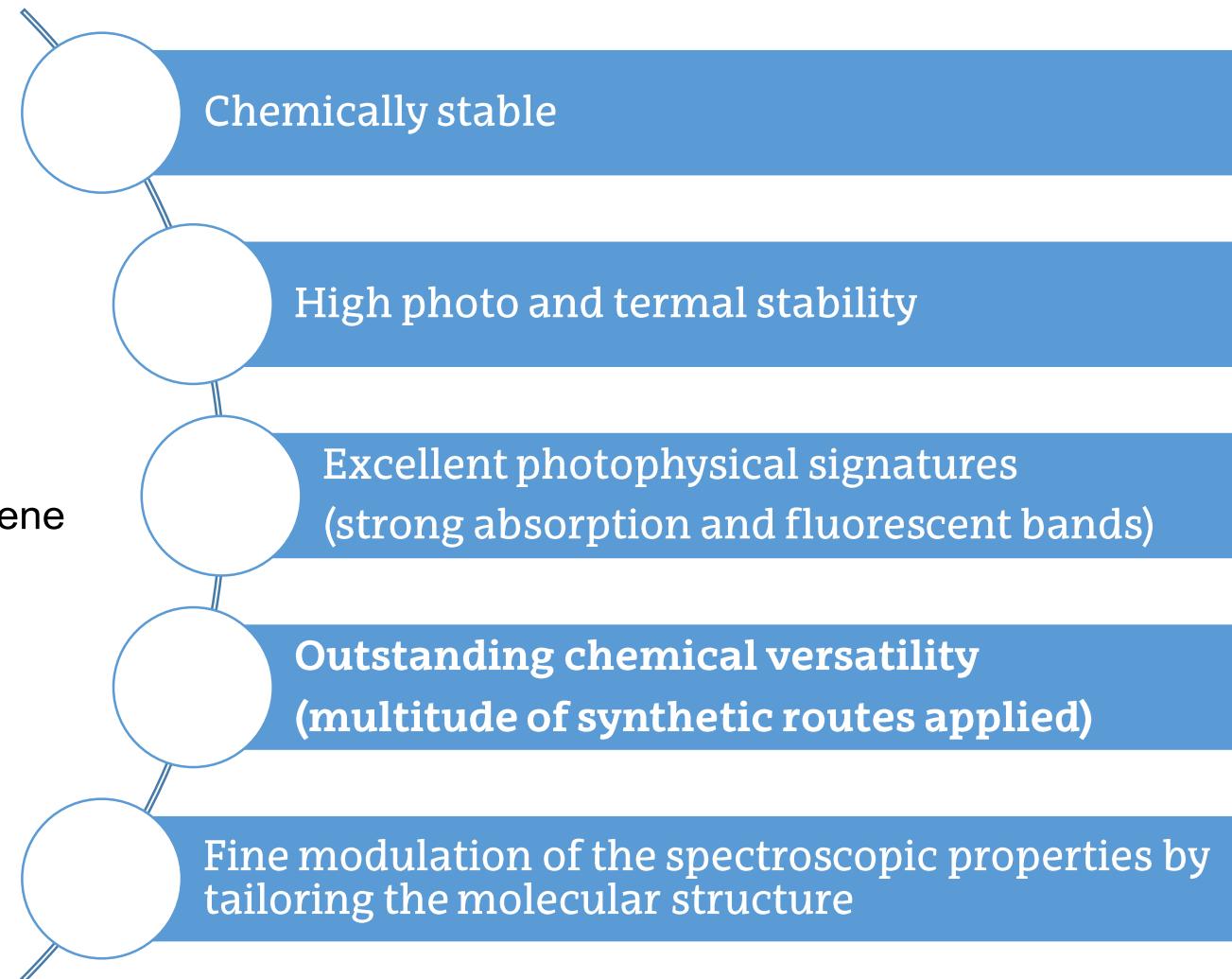
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BODIPY chromophore suitable molecular scaffold as fluorophore:



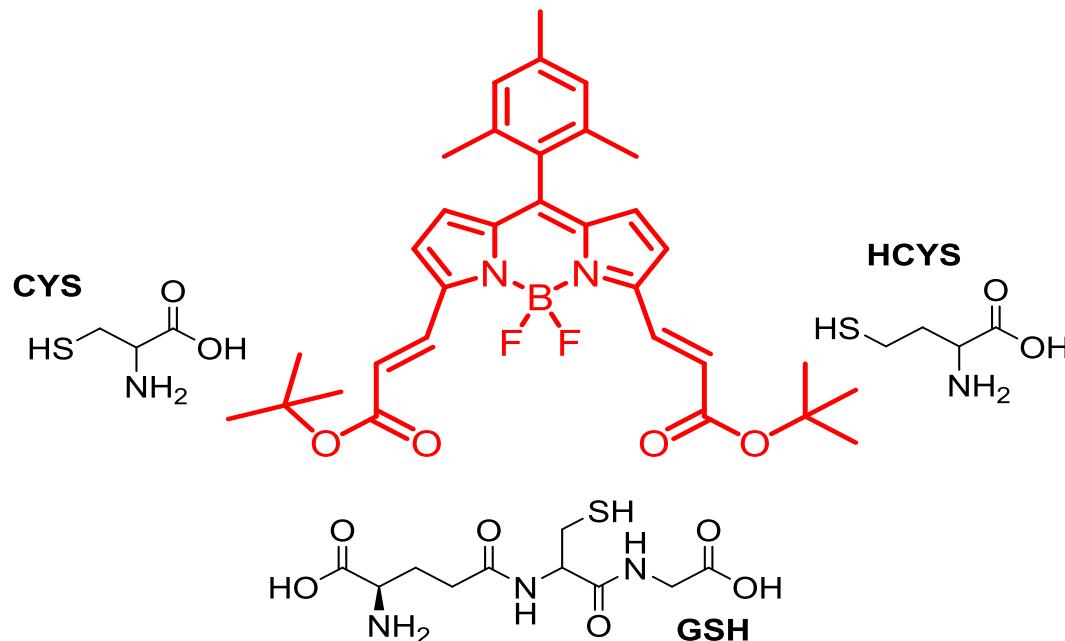
4,4-difluoroboron-3a,4a-diaza-s-indacene
BODIPY



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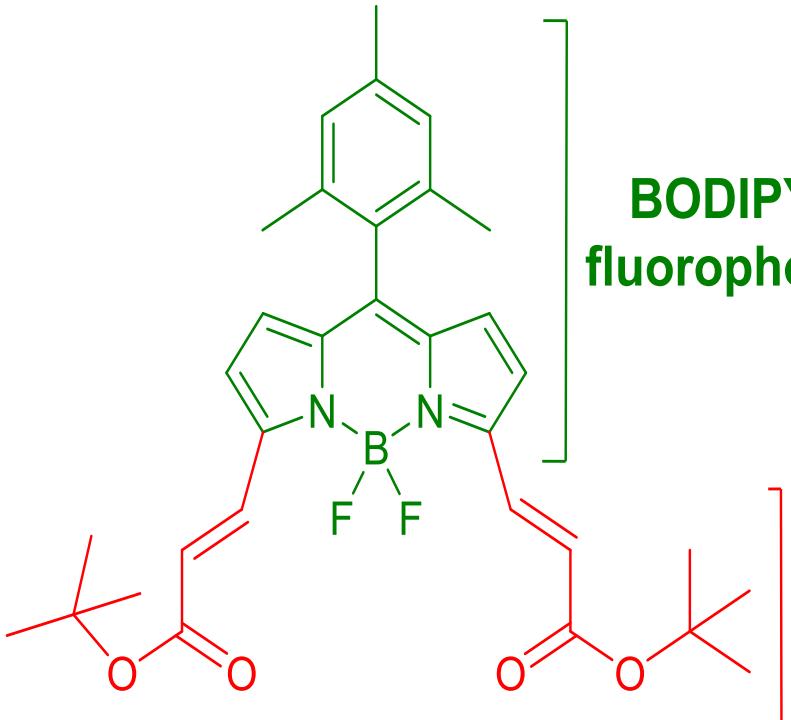
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Test the viability of the designed and synthesized BODIPY based ratiometric chemosensor for the detection of CYS, HCYS and GSH



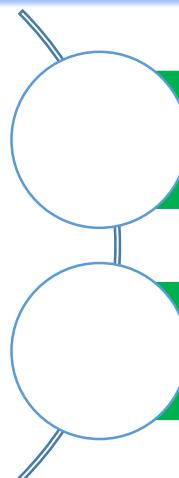
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**BODIPY
fluorophore**

**Amino acid
receptor**



Rigidized molecular structure to ensure high fluorescent response

Modulable spectroscopic signatures tailoring the molecular structure by the substitution pattern at the dipyrromethane core



Reaction sites (vinyl and carbonyl) for binding of the AA



Aromatic moiety at the α -pyrrolic position to extent the chromophoric π -system



Shift of the emission band to the red edge to increase the light penetration into tissues



Binding of the AA should alter the extention of the conjugated π -system and the photophysics

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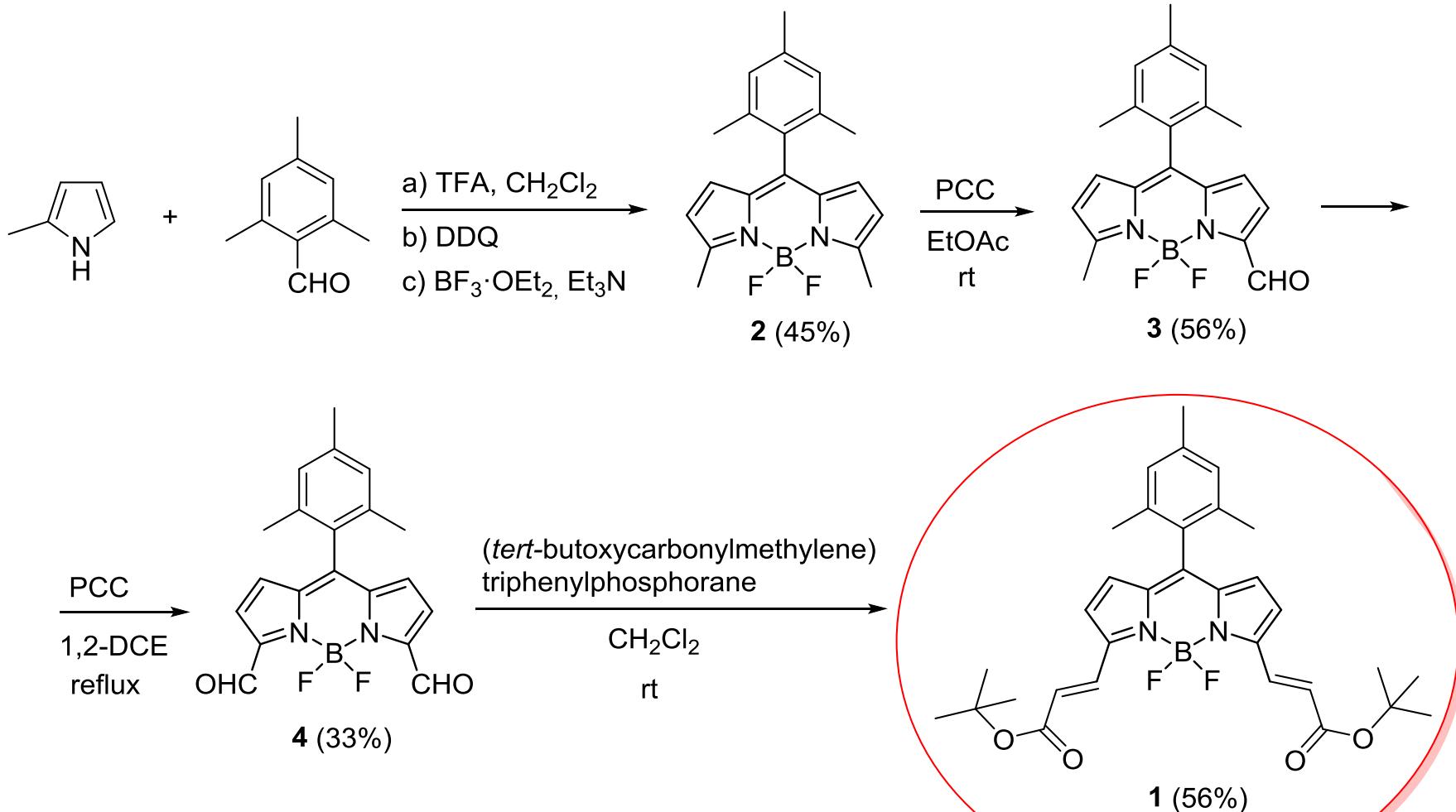
3. Molecular Design

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5. Photophysics

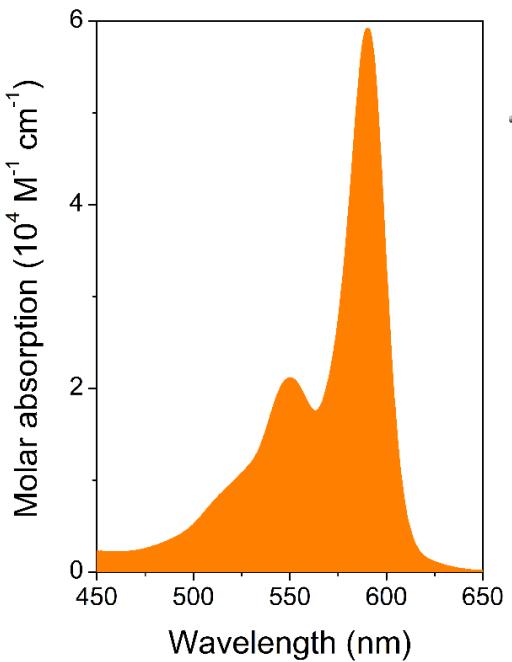
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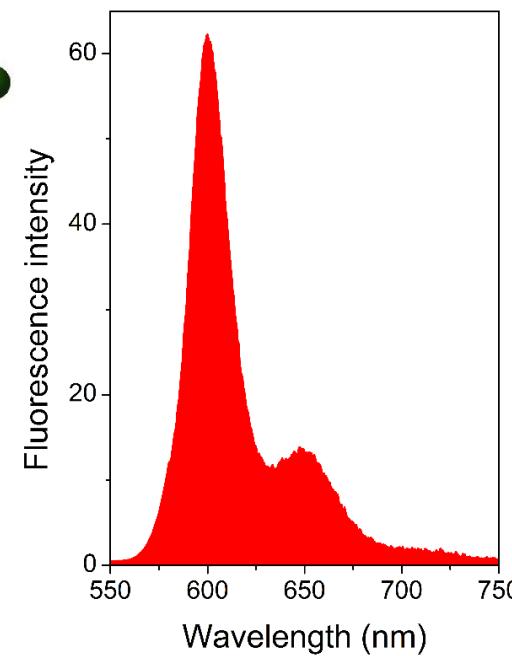
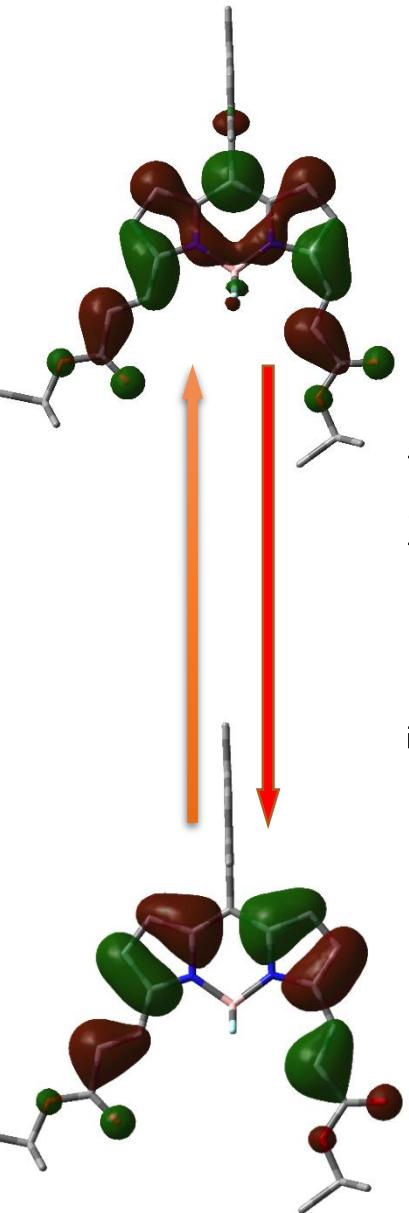


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$\lambda_{ab} = 589.5 \text{ nm}$
 $\epsilon_{\max} = 59000 \text{ M}^{-1} \text{ cm}^{-1}$
(ethanol)



$\lambda_{fl} = 598.5 \text{ nm}$
 $\phi_{fl} = 62\%$
 $\tau_{fl} = 5.42 \text{ ns}$
(ethanol)

The constrained and orthogonal 8-mesityl ensures rigid structure and avoids non-radiative relaxations.

The α -unsaturated diesters induce a pronounced bathochromic shift



High fluorescence at the red edge of the visible

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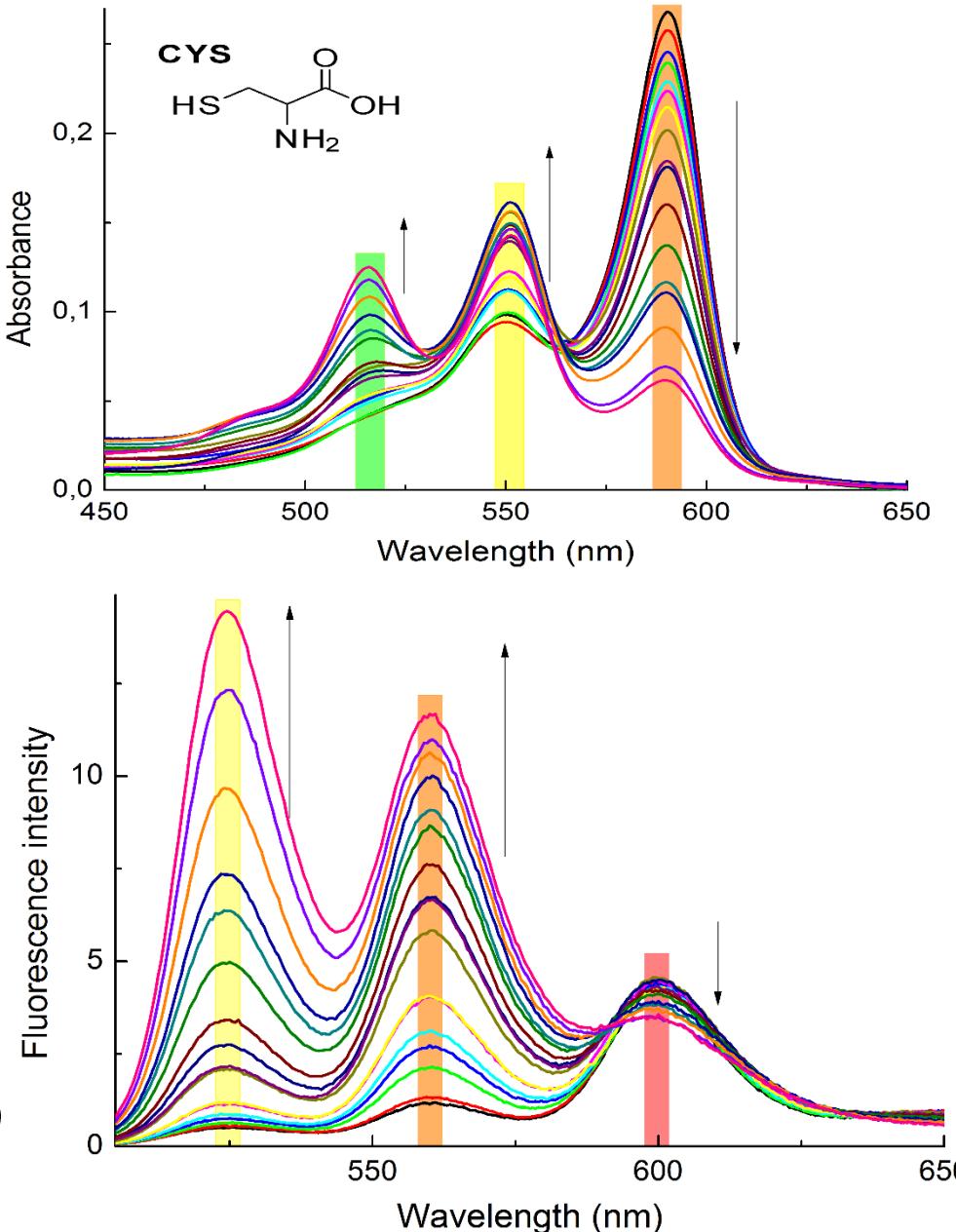
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CYSTEINE

AA

- 0
- 25 μ M
- 50 μ M
- 75 μ M
- 0,1 mM
- 0,15 mM
- 0,2 mM
- 0,25 mM
- 0,3 mM
- 0,4 mM
- 0,5 mM
- 0,7 mM
- 0,9 mM
- 1 mM
- 1,25 mM
- 1,5 mM
- 1,75 mM

EtOH/HEPES (1:1)
2 μ M



Remarkable spectral band splitting of the BODIPY induced by the presence of CYS in the media.

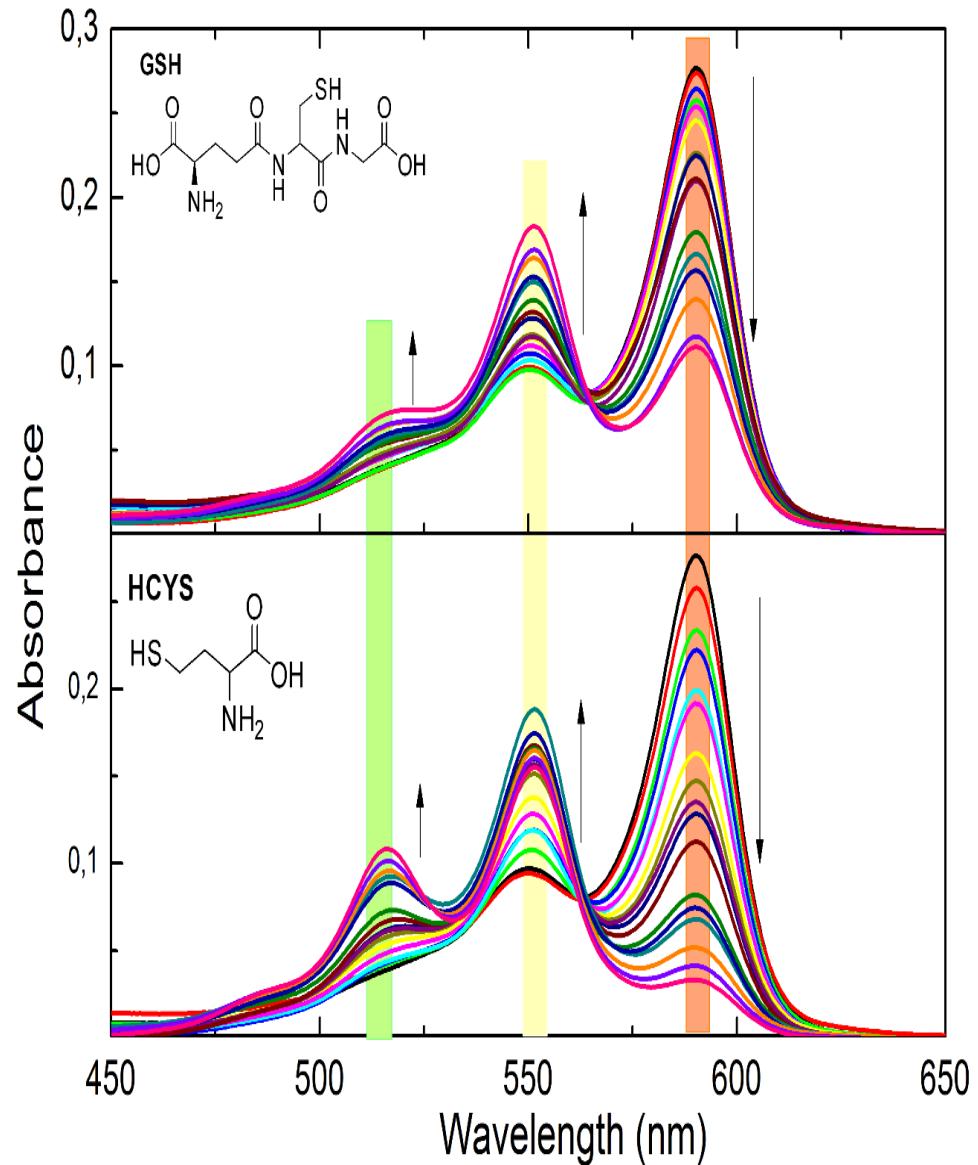
Especially clear in fluorescence, with new and strong hypsochromically shifted emissions.

Similar trends in the rest of AAs.

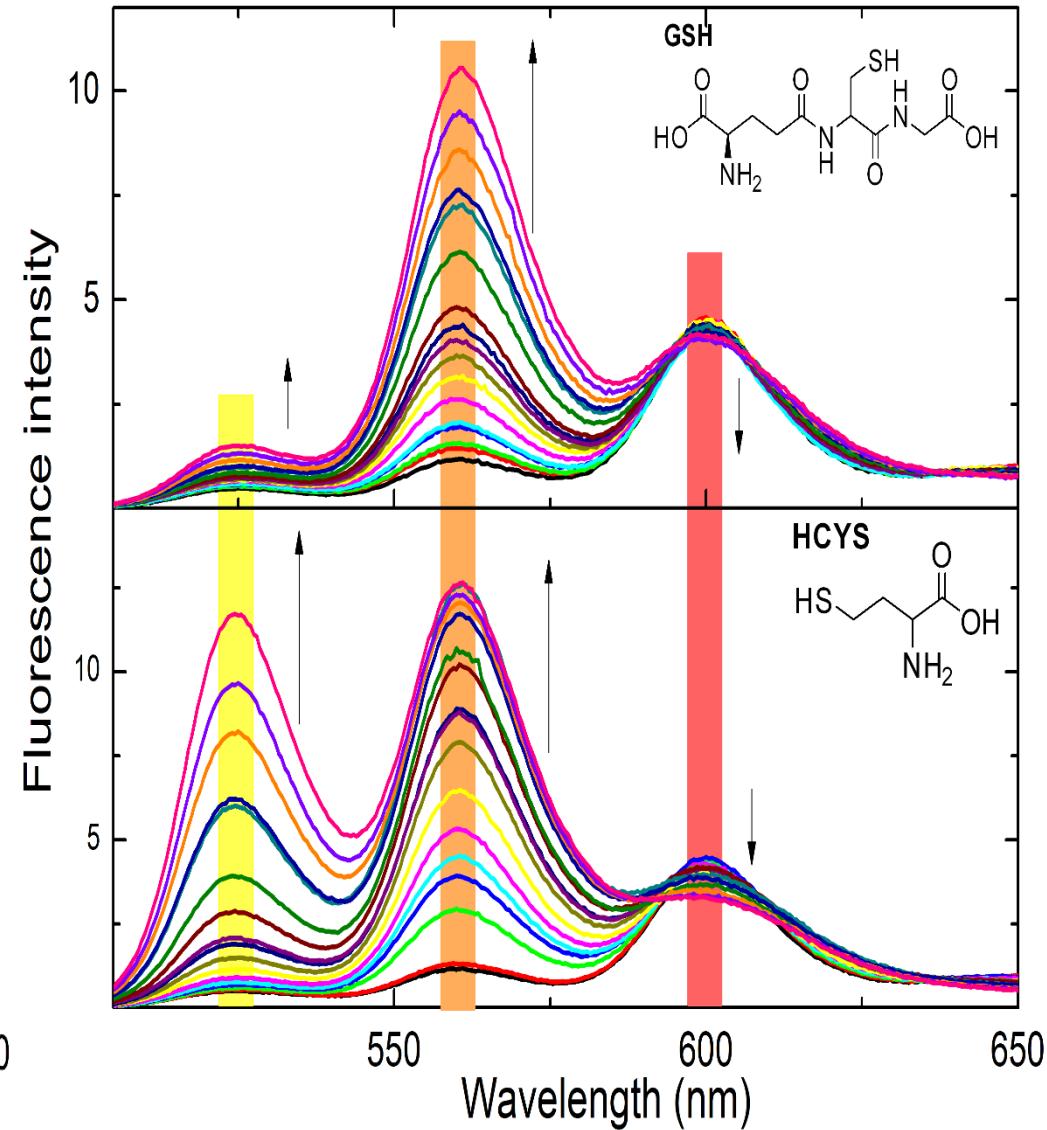
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GLUTATHIONE



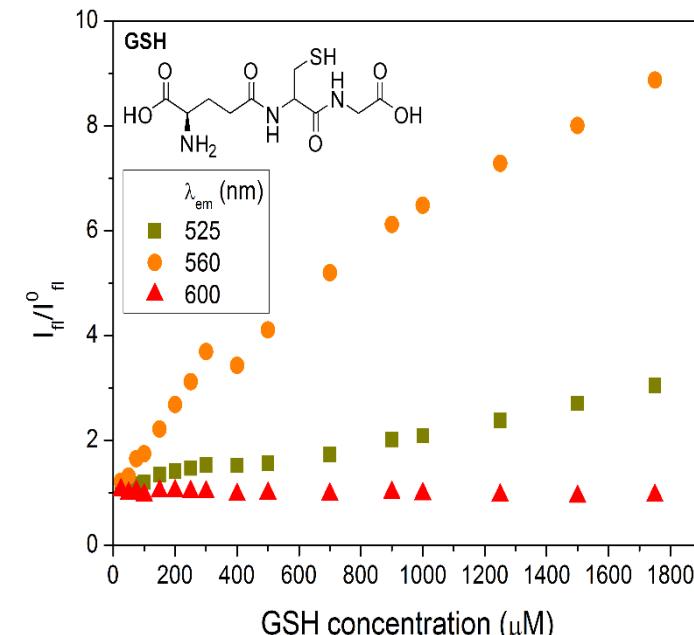
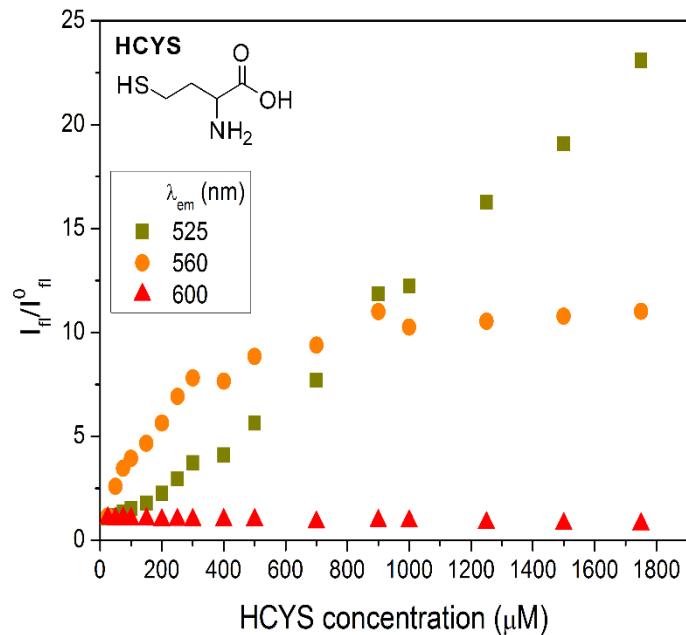
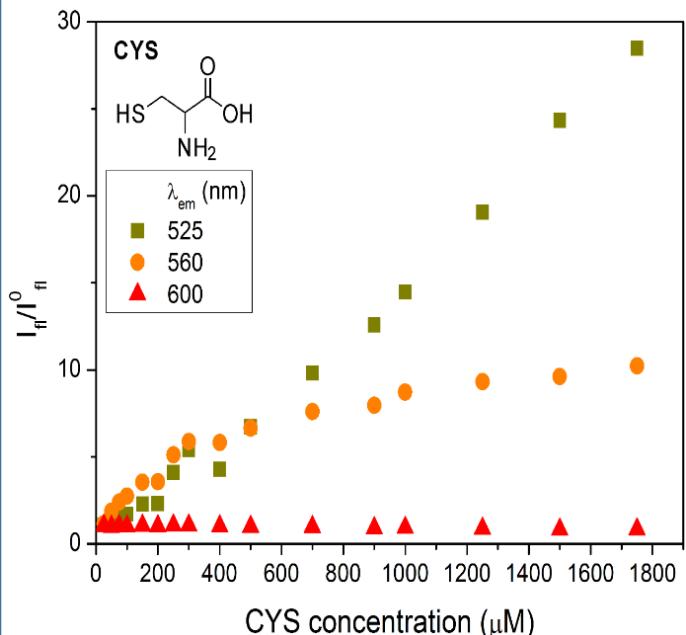
HOMOCYSTEINE



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DETECTION CHANNELS



Up to 3 **detection channels** to monitor and quantify the presence of AA:

- The fluorescence loss at 600 nm
- The fluorescence growing at 525 nm and 560 nm

These last two channels are the most recommended ones

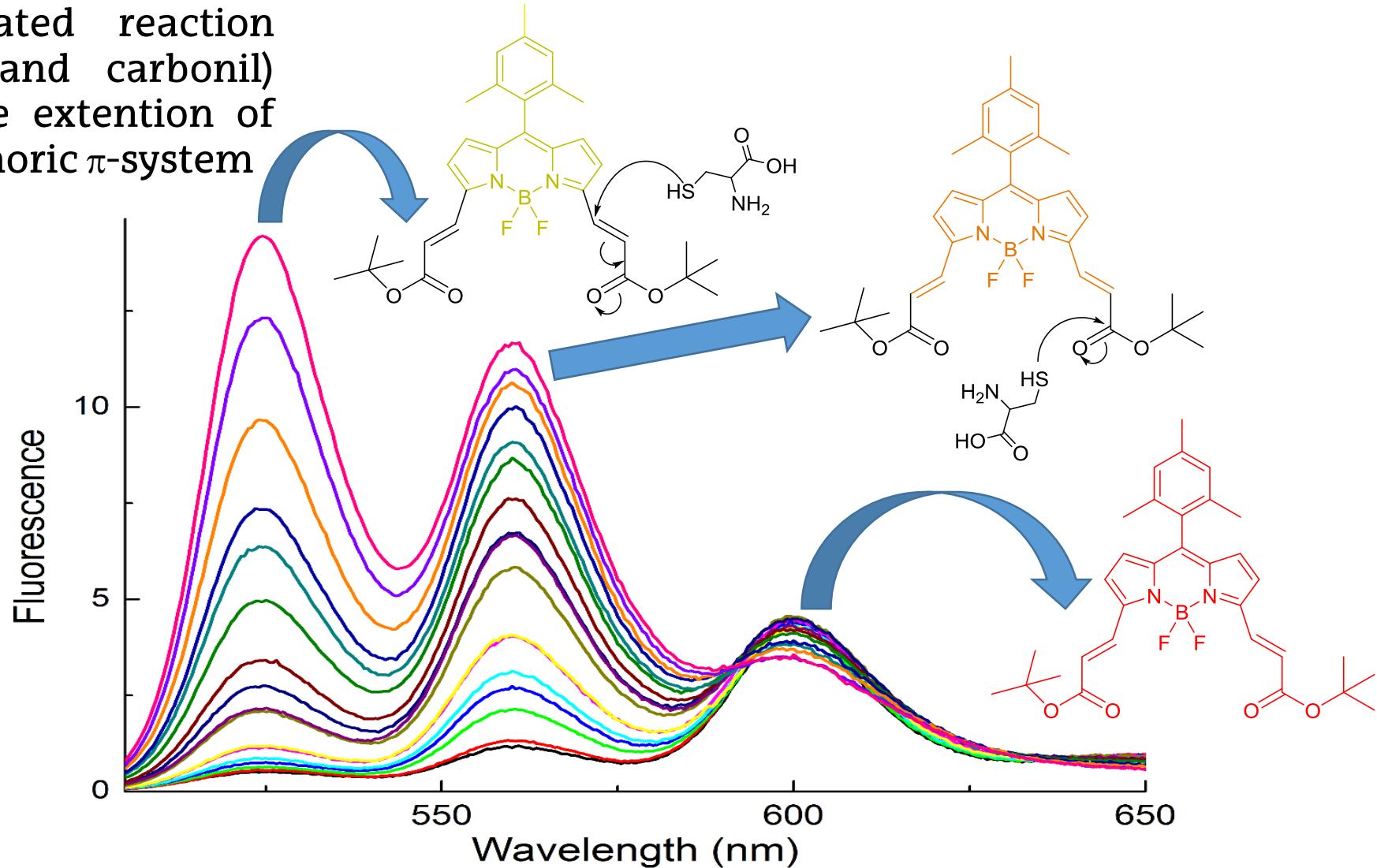
The sensor is **less sensitive** to GSH rather than to CYS and HCYS

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PROPOSED SENSING MECHANISM

The binding of the AA with the unsaturated reaction sites (vinyl and carbonil) decreases the extention of the chromophoric π -system



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The designed BODIPY-based chemosensor for **thiolated AAs** allows:

- **Easy and versatile detection**, just by the emission color.
- **Quantification** of the AA concentration by up to three detection channels.
- **Sensitive detection down to micromolar.**

