

1 *Conference Proceedings Paper*

2 **Functionalized chitosan nanofibers with enhanced**
3 **antimicrobial activity for burn wound healing**
4 **applications**

5 **Alexandru Anisie^{*}, Irina Rosca, Luminita Marin¹**

6 ¹“Petru Poni” Institute of Macromolecular Chemistry, Grigore Ghica Voda Alley, Iasi, Romania

7 Correspondence: *anisie.alexandru@icmpp.ro

8 **Keywords:** keyword 1; keyword 2; keyword 3 (List three to ten pertinent keywords specific to the
9 article; yet reasonably common within the subject discipline.)

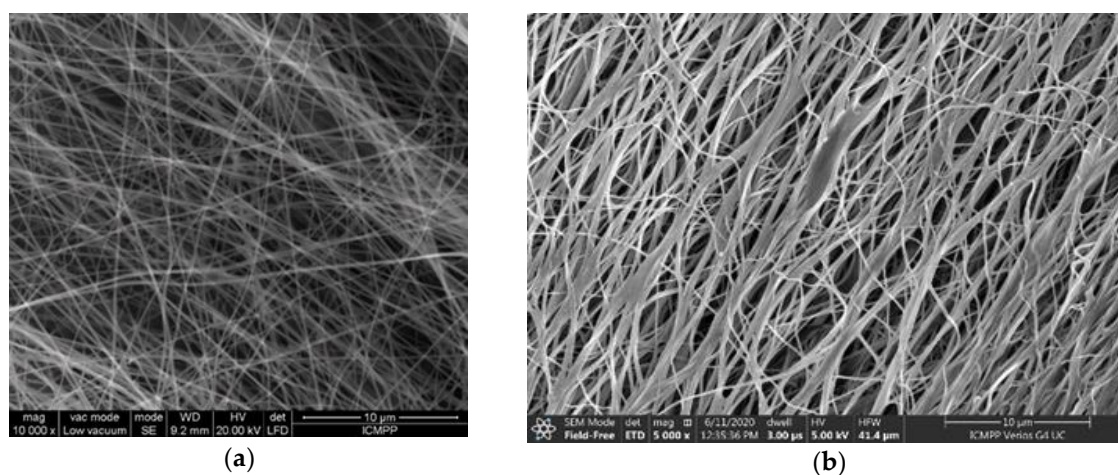
10

11 The electrospinning, a facile, ecological and efficient technique from production cost
12 view, was applied to yield chitosan (CS) nanofibers with sub-micrometric diameter which
13 preserved the intrinsic properties of chitosan such as biocompatibility, lack of toxicity and
14 good therapeutics activity (anti-microbial, anti-fungus, anti-tumor, anti-viral and anti-
15 cholesterolemic activity) with potential for a large variety of applications [1-6].

16 The aim of this study was to prepare chitosan-based nanofibers functionalized with
17 2-formylphenylboronic acid by the imination reaction in heterogenous medium, in order to
18 obtain biodegradable, biocompatible and antimicrobial bandages for burn wound healing
19 applications. The aldehyde has been chosen due to its antifungal and antibiofilm properties
20 demonstrated when it was combined with chitosan [7].

21 The preparation of the proposed fibers was realized in 3 steps. First, CS/PEO fibers
22 were electrospun from a blend solution of CS/PEO (weight ratio of 2/1) in 80% acetic acid
23 using an Inovenso electrospinning apparatus with a rotary collector, when applied the
24 following parameters: voltage equal with 7 kV, tip to collector distance 10 cm, flow rate 0.4
25 ml/h, collector rotation speed 800 RPM, the process being realized at room conditions. The
26 obtained material was neutralized using an aqueous solution of 5% NaOH to remove the
27 residual acetic acid and then it was washed with ultra-pure water to remove the PEO, in order
28 to obtain pure chitosan nanofibers. Further, the chitosan nanofibers were reacted with 2-
29 folmylphenylboronic acid in different conditions to obtain a series of materials with different

30 substitution degrees. The as obtained imine functionalized fibers were morphologically
31 characterized by scanning electron microscopy and polarized optical microscopy. The
32 imination reaction and the substitution degree were monitored by FT-IR and ^1H -RMN
33 spectroscopy. The presence of the imine units was also evidenced by thermo-gravimetical
34 analysis, by variation of the degradation temperature. The water adsorption capacity was
35 investigated by dynamic vapor sorption (DVS) technique and the antimicrobial activity was
36 screened against different bacterial and fungal strains. It was established that the substitutuion
37 degree influence the water sorption capacity of the fibers and the antimicrobial activity, the
38 best results being obtained against *staphylococcus aureus*, *candida albicans* and *aspergillus*
39 *brasiliensis*. It was concluded that as prepared materials keep a high potential for wound
40 healing applications.
41



42 **Figure 1.** SEM images of: (a) Chitosan/PEO nanofibers (b) Iminoboronate chitosan nanofibers

43 **References**

- 45 1. Ohkawa, K., Cha, D., Kim, H., Nishida, A., & Yamamoto, H. (2004). Electrospinning of Chitosan. *Macromolecular Rapid Communications* 25(18): 1600–1605.
- 46 2. Jeon, Y. J., Kim, S. K. (2000). Production of chitoooligosaccharides using ultrafiltration membrane reactor and their antibacterial activity. *Carbohydrate Polymers* 41: 133–141.
- 47 3. Jeon, Y. J., Kim, S. K. (2002) Antitumor activity of chisan oligosaccharides produced in an ultra-filtration
48 membrane reactor system. *Journal of Microbiology and Biotechnology* 12: 503–507.
- 49 4. Hirano, S., Nagao, N. (1989). Effects of chitosan, pectic acid, lysozyme and chitinase on the growth of
50 several phytopathogens. *Agricultural and Biological Chemistry* 53: 3065–3066.
- 51 5. Chirkov, S. N. (2002). The Antiviral Activity of Chitosan (Review). *Applied Biochemistry and Microbiology*
52 38(1): 1–8.
53
54

- 55 6. Sugano, M., Yoshida, K., Hashimoto, M., Enomoto, K., Hirano, S. (1992). Hypocholesterolemic activity of
56 partially hydrolyzed chitosan in rats. In: Brine, C. J., Sandford, P. A., Zikakis, J. P. (Ed.) Advances in chitin
57 and chitosan. Elsevier, London, pp. 472–478
- 58 7. Ailincăi D., Marin L., Morariu S., Mares M., Bostanaru A. C., Pinteala M., Simionescu B. C., M. Barboiu
59 (2016). Dual crosslinked iminoboronate-chitosan hydrogels with strong antifungal activity against Candida
60 planktonic yeasts and biofilms, Carbohydrate Polymers 152: 306-316.

61
62 **Acknowledgements**

63 This work was supported by the Romanian National Authority for Scientific Research MEN – UEFISCDI
64 (grant number PN-III-P1-1.2-PCCDI2017-0569, no. 10PCCDI/2018).



© 2020 by the authors; licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons by Attribution (CC-BY) license (<http://creativecommons.org/licenses/by/4.0/>).