Discovery of selective inhibitor leads by targeting an allosteric site in Insulin-Regulated Aminopeptidase (IRAP)

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The scope

Insulin-regulated aminopeptidase (IRAP) is a transmembrane, zinc-dependent aminopeptidase that performs a variety of important biological functions. IRAP, also known as oxytocinase, leucyl-cystinyl aminopeptidase and placental leucine aminopeptidase is implicated in the regulation of trafficking of glucose transporter type 4, the control of oxytocin levels in pregnancy, as well as the generation of antigenic peptides for cross-presentation. IRAP is also a specific binding site for angiotensin IV, which upon binding serves as a competitive inhibitor of the enzyme. <u>https://www.frontiersin.org/research-topics/12216/physiological-pathological-roles-and-pharmacology-of-insulin-regulated-aminopeptidase</u> IRAP (EC 3.4.11.3) belongs to the M1 family of aminopeptidases and shares high sequence and structural homology with the endoplasmic reticulum aminopeptidases ERAP1 and ERAP2. These enzymes have been shown to adopt conformations between "open" and "closed" states, which expose a large internal cavity for 🔧



peptide entrance and mediate efficient substrate processing. <u>https://www.frontiersin.org/articles/10.3389/fimmu.2017.00946/full</u>

Due to its involvement in several human disease states, IRAP is an emerging pharmaceutical target. With the aim to discover inhibitors of IRAP that do not disrupt the physiological function of other M1 enzymes, we pursuit the discovery of lead-like compounds that bind away from the highly homologous active site.

A recent, high-resolution crystal structure of "closed" **ERAP1** in complex with a potent phosphinic pseudopeptide inhibitor revealed **distinct druggable pockets** occupied by two molecules from the crystallization medium.

The idea



Garde binding Hinge domain **C-terminal domain**

The method

First we identified the corresponding pockets in the C-termial domain of IRAP (PDB ID: 5MJ6), which interestingly, share comparably low homology with those of ERAP1 and ERAP2. To discover high-affinity compounds that bind and disrupt the physiological processing of large substrates of IRAP, we employed structure-based virtual screening according to the following steps:

From the purchasable chemical space of the **ZINC-15** database, we selected a subset of lead-like compounds (MW of 200–350 and cLogP < 3.0), clean from reactive groups or PAINS (anodyne subset) and in-stock. A total of 2.6 million compounds were docked at the selected site of IRAP using **AutoDock VINA** in order to select the **top-ranked 1%** compounds with the highest affinity.

These compounds were also docked at the related pocket of

14 × 10⁶ purchasable compounds from ZINC

2.6 × 10⁶ lead-like subset selected for docking

 25×10^3 top-ranked

ligands for IRAP pocket

305 more selective

for IRAP versus ERAP1

33 obtained for

in vitro screening

We opted to **target pocket 1** as the largest pocket and a site that comprise residues from all three functional domains of the enzymes, next to the hinge domain.

The result

Screening of the compounds using the fluorigenic Leu-AMC substrate revealed 3 compounds with IC_{50} values < 100 μ M and characteristic inhibition plots shown.







ERAP1, so as to **disfavor compounds of equally high affinity** for the enzyme with the highest homology to IRAP. After visual investigation of residue-specific interactions with the topranked compounds, we selected 305 compounds for **molecular dynamics simulations** of 20 ns each (> 6 μ s in total). From the second half of the MDs we obtained 1,000 snapshots and carried out free energy calculations with the MM/GBSA and MM/PBSA methods, which guided the final selection of the most promising compounds for experimental investigation.

For their evaluation we employed two orthogonal functional assays: one using a small fluorigenic substrate (Leu-AMC), and one by following the degradation of oxytocin by IRAP using HPLC.

Interestingly, compounds that inhibit Leu-AMC with IC_{50} <100 μ M did not display similar inhibition of the natural substrate, oxytocin. In contrast, compound 26 that does not **inhibit Leu-AMC, exhibited 69% inhibition of the cyclic substrate** at 100 µM tested. From the dose-dependent inhibition of oxytocin degradation by IRAP, followed by HPLC in the presence of compound **26**, we estimated an IC_{50} value of 40 μ M (shown below).



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