



Effects of process parameters on the color quality of anthocyanin-based colorants from conventional and microwave-assisted aqueous extraction of sweet potato (*Ipomoea batatas* L.) leaf varieties

Jin Mark DG. Pagulayan, Aprille Suzette V. Mendoza, Fredelyn M. Gascon,
Jan Carlo C. Aningat, Abigail S. Rustia, Casiana Blanca J. Villarino

*Presentation for the 1st International Electronic Conference on Food Science and Functional Foods
November 10-25, 2020*

Rationale

- Color is a significant trait of sensory quality evaluation and a key attribute that affects the perception of an individual to foods. Colorants are to make food products more appealing and desirable to consumers
- Natural colorants are more preferred because of the detrimental issues on safety and health by synthetic colorants.

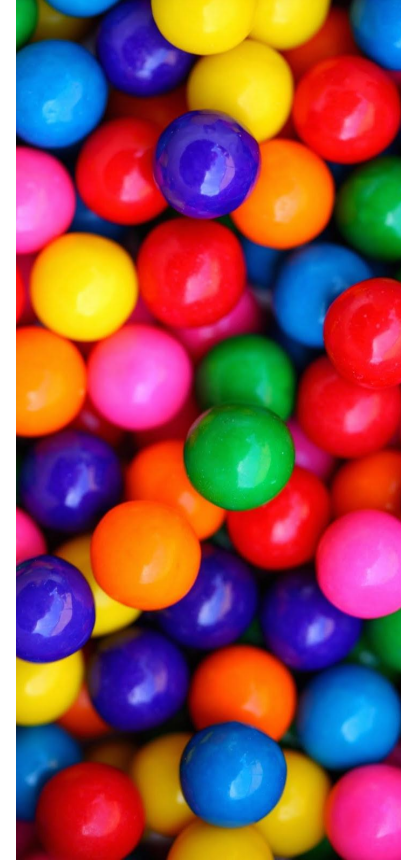


Image source: https://blog.kulikulifoods.com/wp-content/uploads/2015/03/1334760_75811556.jpeg

<https://1nuc2h2s2zs3uw82f35tl0ki-wpengine.netdna-ssl.com/wp-content/uploads/foodcoloring-340x340.jpg>

Rationale

- **Anthocyanins**

- Approved natural food colorant which is also the most important water-soluble natural pigments among group of flavonoids
- known for their excellent antioxidant properties which can protect the human body from oxidative stress thereby decreasing the risk of chronic disease such as aging, diabetes, cardiovascular diseases, and cancer
- A wide range of color hues such as red, violet, purple and blue in plants like fruits, flowers, leaves, and tubers can be produced by anthocyanins at varying pH
- Applied to foods and beverages such as yogurt, jellies, juices, and wine
- pH, temperature, light, oxygen, and metal ions affect the stability of anthocyanins

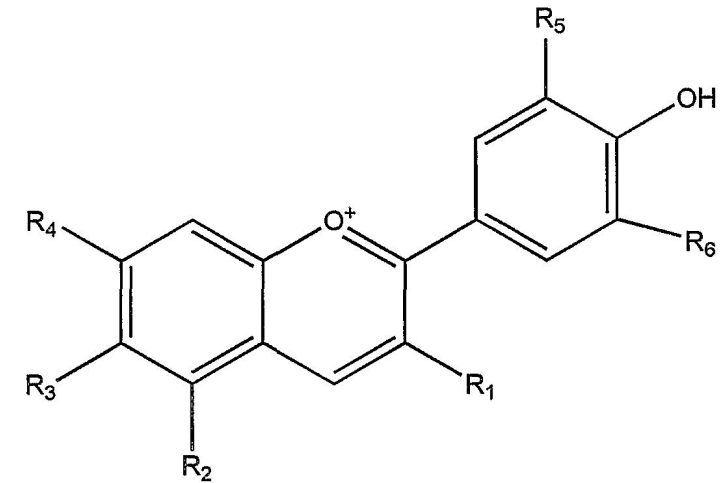
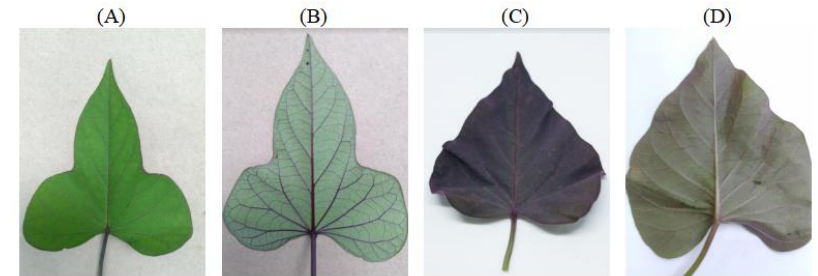


Image source: <https://www.pathway27.eu/files/53/0000000153.png>

<https://nutritionyoucanuse.com/wp-content/uploads/2018/01/canstockphoto53806795.jpg>

Rationale

- One of the potential sources of anthocyanins is sweet potato (*Ipomoea batatas* L.), the seventh most important food crop and the second root and tuber crop grown next to cassava.
- In comparison with the other parts of sweet potato, the leaves contain significantly higher amounts of anthocyanins
- The anthocyanins in sweet potato leaf (SPL) have high potential that they can be used as natural colorants in foods
- They have superior shelf life and equal stability in light and heat as those in red cabbage

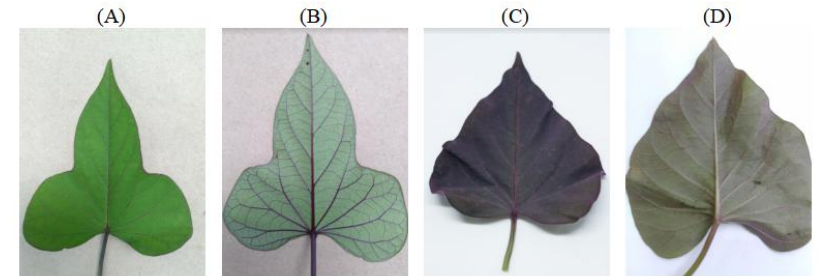


Objective of the Study

To evaluate the effects of process parameters (time and raw material weight [RMW]) of conventional and microwave-assisted aqueous extraction on the color quality (i.e. lightness [L^*], chroma [C^*] and hue [H°]) of anthocyanin –based colorants of red and Inubi sweet potato (*Ipomoea batatas* L.) leaves

Materials and Methods

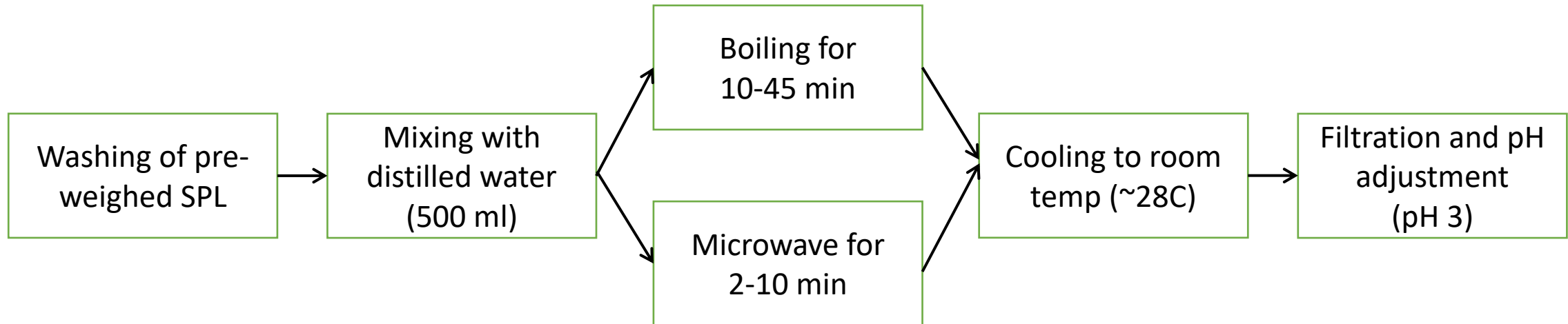
- Inubi and red SPL variety were used
- Red and violet pigments can be observed throughout the leaf blade and veins of red SPL while those of Inubi SPL can only be seen on the veins.
- Inubi SPL were obtained from Sapang Multipurpose Cooperative, Moncada, Tarlac while red SPL was procured from Fresh-Q Industries, Brgy. Sulucan, Angat, Bulacan.
- Leaves were procured at the age of 3 months and transported to the laboratory where it was stored inside the chiller ($\sim 4^{\circ}\text{C}$) prior to analysis.



Materials and Methods

Extraction Methods

- Conventional (boiling) and Microwave-assisted



Materials and Methods

Table 2.1. Actual values of process parameters of the 14 Runs in the central composite design.

Standard Order	Block	Point Type	X ₁ , RMW(g per 500 ml d'H ₂ O)	X ₂ , BT (min) ^a	X ₂ , MT (min) ^b
1	Block 1 (Day 1)	Factorial	50.00	10.00	2.00
2		Factorial	125.00	10.00	2.00
3		Factorial	50.00	45.00	8.00
4		Factorial	125.00	45.00	8.00
5		Center	87.50	27.50	5.00
6		Center	87.50	27.50	5.00
7		Center	87.50	27.50	5.00
8	Block 2 (Day 2)	Axial	34.47	27.50	5.00
9		Axial	140.53	27.50	5.00
10		Axial	87.50	2.75	0.76
11		Axial	87.50	52.25	9.24
12		Center	87.50	27.50	5.00
13		Center	87.50	27.50	5.00
14		Center	87.50	27.50	5.00

^aBT- Boiling time for conventional extraction

^bMT- Microwave extraction time for microwave-assisted extraction (MAE)

Materials and Methods

Table 2.2. Central composite experimental design showing independent variables with actual and coded values.

Factor	Parameter	Units	Actual Values		Coded Values	
			Minimum	Maximum	Minimum	Maximum
X_1	Raw material weight(RMW)	g per 500 mL water	50	125	-1	1
X_2^a	Boiling time (BT)	min	10	45	-1	1
X_2^b	Microwave extraction time (MT)	min	2	8	-1	1

^aFor conventional extraction

^bFor microwave-assisted extraction (MAE)

Materials and Methods

Modelling of Responses

- Different RMW-BT and RMW-MT combinations were generated using Design-Expert® Application V.7.0.0 (DX7, Stat-Ease Inc.).
- Central composite rotatable response surface methodology (RSM) design

$$Y = \beta_0 + \sum_{i=1}^n \beta_i X_i + \sum_{i=1}^n \beta_{ii} X_i^2 + \sum \sum_{i < j=1}^n \beta_{ij} X_i X_j, \quad (1)$$

Materials and Methods

Color Measurement

- Using a bench spectrophotometer (ColorFlexEZ, Hunter Associates Laboratory Inc., Virginia, USA)

$$\text{Chroma}(C^*) = \sqrt{a^{*2} + b^{*2}}, \quad (2)$$

$$\text{Hue}(H^\circ) = \arctan\left(\frac{b^*}{a^*}\right), \quad (3)$$

Materials and Methods

Color Measurement

- Using a bench spectrophotometer (ColorFlexEZ, Hunter Associates Laboratory Inc., Virginia, USA)

$$\text{Chroma}(C^*) = \sqrt{a^{*2} + b^{*2}}, \quad (2)$$

$$\text{Hue}(H^\circ) = \arctan\left(\frac{b^*}{a^*}\right), \quad (3)$$

Results and Discussion

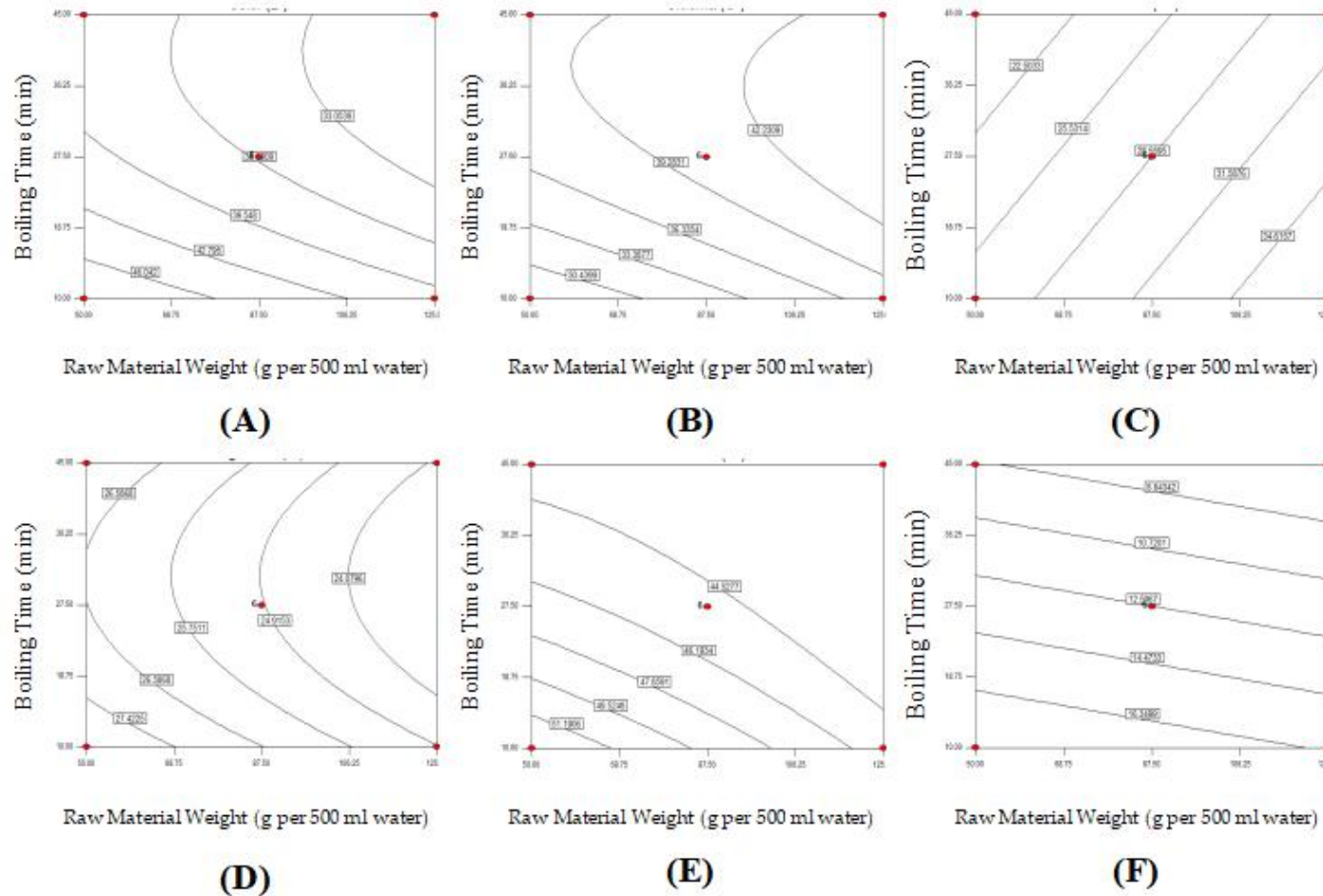


Figure 3.1. Contour plot showing the effect of raw material weight and boiling time on the color of *Imubi*(A-C) and red (D-F) SPLC: (From left to right) lightness, chroma and hue of SPLC.

Results and Discussion

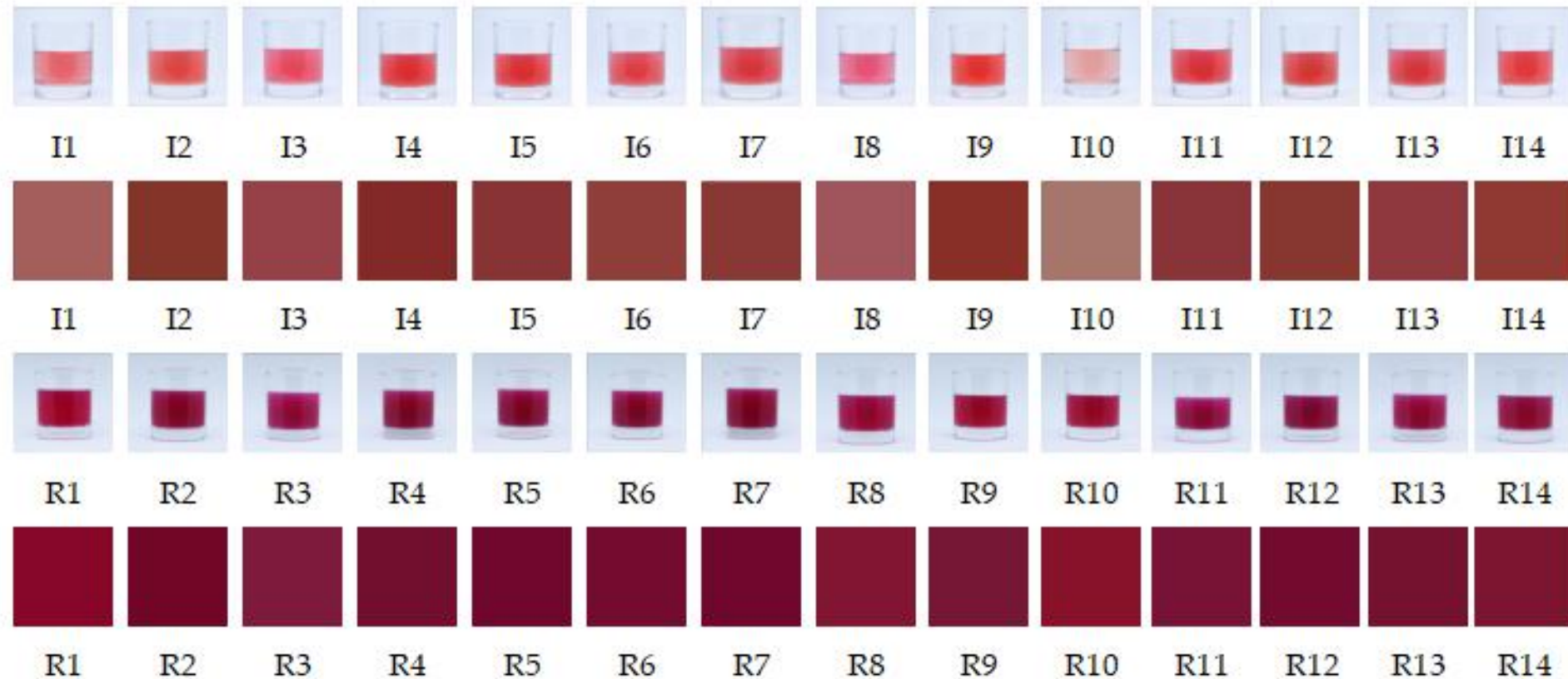


Figure 3.2. Actual images and color swatches of SPLC samples extracted through conventional method arranged according to standard number: (I1-I14) Inubi SPLC and (R1-R14) red SPLC.

Results and Discussion

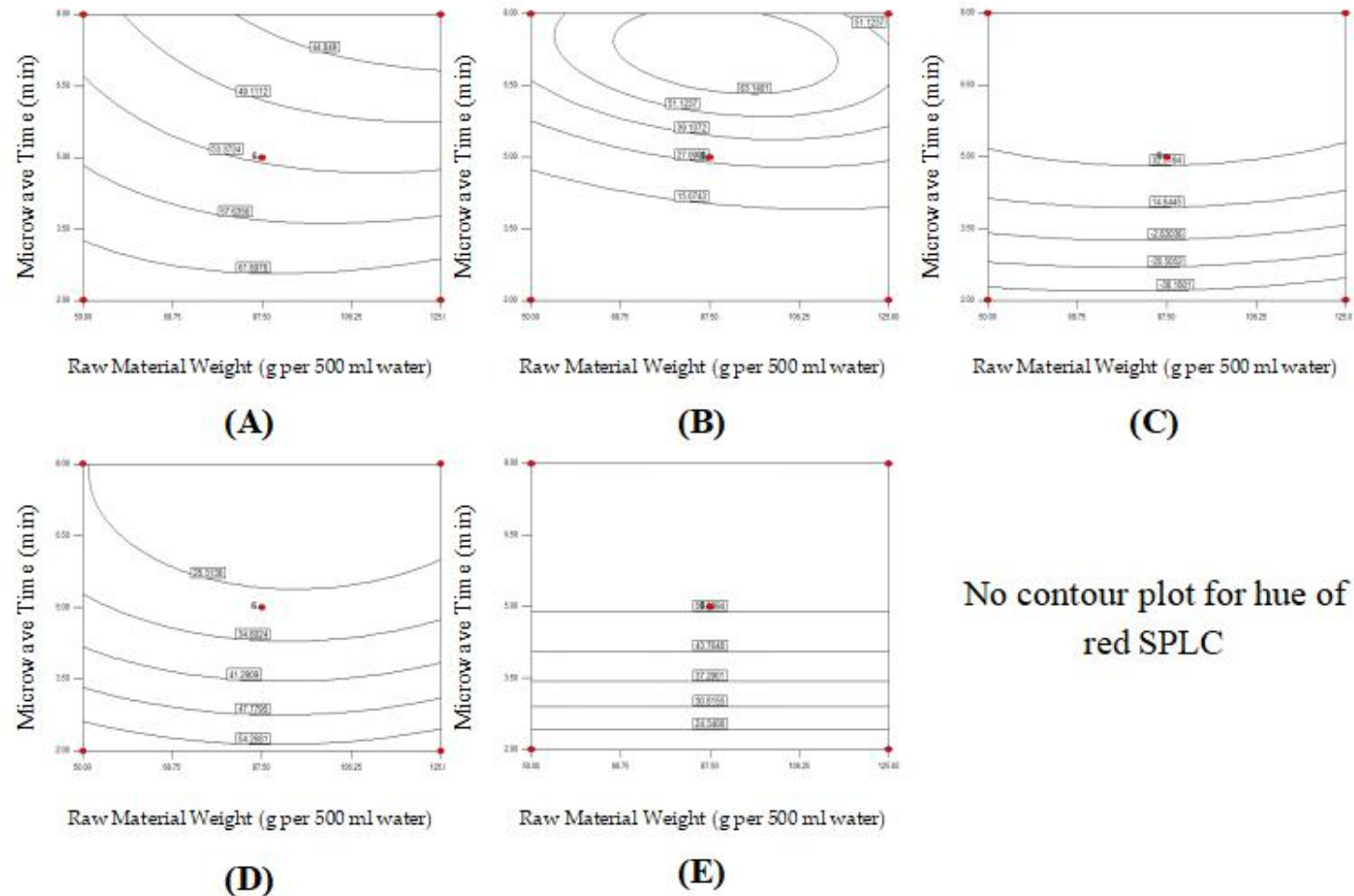


Figure 3.3. Contour plot showing the effect of raw material weight and microwave time on the color of *Inubi*(A-C) and red (D-E) SPLC: (From left to right) lightness, chroma and hue of SPLC.

Results and Discussion

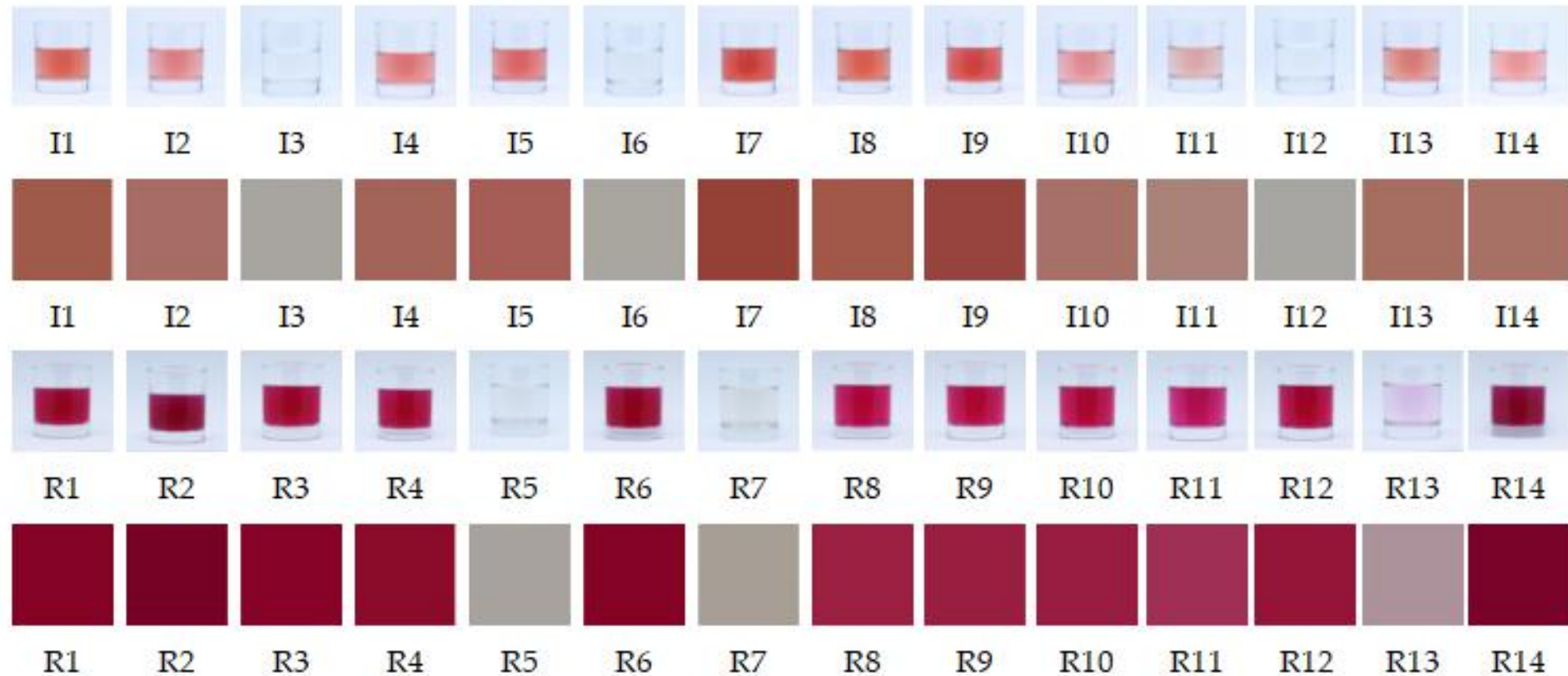


Figure 3.4. Actual images and color swatches of SPLC samples extracted through MAE arranged according to standard number: (I1-I14) Inubi SPLC and (R1-R14) red SPLC.

Results and Discussion

Table 3.3. Summary of verification experiment results.

<u>Response^e</u>	Conventional Extraction		Microwave Assisted Extraction	
	<u>Inubi SPLC-A¹</u>	<u>Inubi SPLC-B²</u>	<u>Inubi SPLC-A⁴</u>	<u>Inubi SPLC-B⁵</u>
Lightness (L^*)	35.04±1.93 ^b	39.97±1.89	81.23±0.01 ^{ab}	88.01±0.01 ^{ab}
Chroma (C^*)	41.34±0.56 ^b	40.12±1.91	15.46±0.01	8.65±0.01 ^{ab}
Hue (H°)	26.54±3.71	33.38±0.88 ^b	-82.14±0.05 ^{ab}	75.15±0.06 ^{ab}
	Red SPLC-A ²	Red SPLC-B ³	Red SPLC-A ⁴	Red SPLC-B ⁶
Lightness (L^*)	23.41±0.81	23.90±0.16 ^b	39.70±3.71	25.00±0.95
Chroma (C^*)	46.57±0.20	45.08±0.36 ^b	46.60±4.99	49.90±0.79
Hue (H°)	15.09±0.57	8.20±1.36 ^b	5.60±1.47 ^b	18.60±1.64

¹Conditions: 75 g of SPL per 500 mL distilled water boiled for 20 min

²Conditions: 125 g of SPL per 500 mL distilled water boiled for 10 min

³Conditions: 50 g of SPL per 500 mL distilled water boiled for 30 min

⁴Conditions: 34.5 g of SPL per 500 mL distilled water extracted through MAE for 5 min

⁵Conditions 50 g of SPL per 500 mL distilled water extracted through MAE for 2 min

⁶Conditions 125 g of SPL per 500 mL distilled water extracted through MAE for 8 min

^aSignificantly different from the predicted value based on prediction interval.

^bSignificantly different from the predicted value based on confidence interval.

^eExpressed as mean ± standard deviation.

Conclusion and Recommendation

- Boiling time/microwave time and raw material weight affected the color quality of Inubi and red SPLC
- RMW and BT and MT generally had a significant ($p < 0.05$) effect on the color (L^* , C^* , H°) of both SPLC varieties.
- A more in-depth analysis on the possible effects of process parameters on the extraction of anthocyanin-based colorants from sweet potato leaves
- Useful on the possible optimization as well as exploring more on the nature of colorants found in SPLC
- It is recommended that the effect of process parameters of other extraction methods on the quality of SPLC and other possible sources of anthocyanin-based colorants to be explored.

References

1. Stich, E. Food Color and Coloring Food: Quality, Differentiation, and Regulatory Requirements in the European Union and the United States. In Handbook on natural pigments in food and beverages industrial applications for improving food color, 1st ed.; Carle, R., & Schweiggert, R. M., Eds.; Woodhead Publishing: Duxford, UK, 2016; pp. 3-27.
2. Garber, L. L., Hyatt, E. M., & Nafees, L. The Effects of Analogous Food Color on Perceived Flavor: A Factorial Investigation. *J. Food Prod. Mark.* 2016, 22(4), 486-500. doi:10.1080/10454446.2015.1072866
3. Burrows, G. & Hornero-Mendoza, D. Carotenoids and colour in fruit and vegetables. In *Phytochemistry of Fruits and Vegetables*, Tomas Barberan, F.A., Robins, R.J. Eds.; Oxford University Press: Oxford, UK, 1997.
4. Zhang, J. S., Sokhansaj, S., Wu, R. Fang, W., & Yang, P. A Transformation Technique from RGB Signals to the Munsell System for Colour Analysis of Tobacco Leaves. *Comput. Electron. Agric.* 1998, 19, 155-166.
5. Shi, Z.; Francis, J. F. & Daun, H. Quantitative Comparison of Stability of Anthocyanins from Brassica oleracea and Tradescantia pallida in Non-sugar Drink Model and Protein Model Systems. *J. Food Sci.* 1992, 57, 768-770.
6. Hine, T. *The Total Package: The Evolution and Secret Meanings of Boxes, Bottles, Cans, and Tubes*, 1st ed.; Little Brown & Co: New York, USA, 1995.
7. Marmion, D. M. *Handbook of U.S. Colorants: Foods, Drugs, Cosmetics, and Medical Devices*, 1st ed.; Wiley Publishing; New York, USA, 1991.
8. Pasiadis, I., Asimakopoulos, A., & Thomaidis, N. Food colours for bakery products, snack foods, dry soup mixes, and seasonings. In *Colour Additives for Foods and Beverages*, 1st ed.; Scotter, M. J. Ed.; Woodhead Publishing: Duxford, UK, 2015; pp. 211-226.
9. Carochio, M., Morales, P., & Ferreira, I. C. F. R. Natural Food Additives: Quo Vadis?. *Trends Food Sci. Technol.* 2015, 45(2), 284–295. doi: 10.1016/j.tifs.2015.06.007
10. Weiss, B. Synthetic Food Colors and Neurobehavioral Hazards: The View from Environmental Health Research. *Environ. Health Perspect.* 2011, 120(1), 1-5. doi:10.1289/ehp.1103827
11. Potera, C. The Artificial Food Dye Blues. *Environ. Health Perspect.* 2010. 118, 428-430. doi: 10.1289/ehp.118-a428
12. Leong, H. Y., Show, P. L., Lim, M. H., Ooi, C. W., & Ling, T. C. Natural Red Pigments from Plants and Their Health Benefits: A Review. *Food Rev. Int.* 2018, 34(5), 463-482. doi: 10.1080/87559129.2017.1326935
13. Timberlake, C. F., & Henry, B. S. Plant Pigments as Natural Food Colours. *Endeavour* 1986, 10(1), 31-36.
14. Shipp, J., & Abdel-Aal, E. S. M. Food Applications and Physiological Effects of Anthocyanins as Functional Food Ingredients. *Open Food Sci. J.* 2010, 4(1), 7-22. doi: 10.2174/1874256401004010007
15. Sigurdson, G. T., Tang, P., & Giusti, M. M. Natural Colorants: Food Colorants from Natural Sources. *Annu. Rev. Food Sci. Technol.* 2017, 28(8), 261–280. doi: 10.1146/annurev-food-030216-025923
16. Takeoka, G. & Dao, L. Anthocyanin. In *Methods of Analysis for Functional Foods and Nutraceuticals*, 1st ed.; Hurst, W. J. Eds.; Taylor & Francis Inc.: Florida, USA, 2002; pp. 219-241.

References

17. Huang, M., & Ferraro, T. Phenolic Compounds in Food and Cancer Prevention. ACS Symposium Series Phenolic Compounds in Food and Their Effects on Health II, 1992, 8-34. doi:10.1021/bk-1992-0507.ch002
18. Shimozono, H., Kobori, M., Shinmoto, H., & Tsushida, T. Suppression of the Melanogenesis of Mouse Melanoma B16 Cells by Sweet Potato Extract. Nippon Shokuhin Kagaku Kogaku Kaishi 1996, 43(3), 313-317. doi:10.3136/nskkk.43.313
19. Tsai, P., Mcintosh, J., Pearce, P., Camden, B., & Jordan, B. R. Anthocyanin and Antioxidant Capacity in Roselle (Hibiscus Sabdariffa L.) Extract. Food Res. Int. 2002, 35(4), 351-356. doi:10.1016/s0963-9969(01)00129-6
20. Duangmal, K., Saicheua, B., & Sueeprasan, S. Colour evaluation of freeze-dried roselle extract as a natural food colorant in a model system of a drink. LWT 2008, 41(8), 1437-1445. doi:10.1016/j.lwt.2007.08.014
21. Wrolstad, R. E. Anthocyanins. In Natural Food Colorants: Science and Technology, 1st ed.; Lauro, G. J. and Francis, F. J. Eds.; Taylor & Francis Inc.: Florida, USA, 2000.
22. Horbowicz, M., Kosson, R., Grzesiuk, A., Debski, H. Anthocyanins of Fruits and Vegetables – Their Occurrence, Analysis and Role in Human Nutrition. Veg. Crops Res. Bull. 2008, 68, 5-22.
23. Khoo, H. E., Azlan, A., Tang, S. T., & Lim, S. M. Anthocyanidins and Anthocyanins: Colored Pigments as Food, Pharmaceutical Ingredients, and the Potential Health Benefits. Food Nutr. Res. 2017, 61(1), 1361779. doi 10.1080/16546628.2017.1361779
24. Laleh, G. H., Frydoonfar, H., Heidary, R. et al. The Effect of Light, Temperature, pH and Species on Stability of Anthocyanin Pigments in Four Berberis Species. Pakistan J. Nutr. 2006, 5(1), 90-92. doi:10.3923/pjn.2006.90.92
25. He, J., & Giusti, M. M. Anthocyanins: Natural Colorants with Health-promoting Properties. Annu. Rev. Food Sci. Technol. 2010, 1(1), 163–187. doi: 10.1146/annurev.food.080708.100754
26. Ahmed, M., Akter, M. S., Lee, J. C., & Eun, J. B. Encapsulation by Spray Drying of Bioactive Components, Physicochemical and Morphological Properties from Purple Sweet Potato. LWT 2010, 43(9), 1307-1312. doi: 10.1016/j.lwt.2010.05.014
27. Cortez, R., Luna-Vital, D. A., Margulis, D., & Gonzalez de Mejia, E. Natural Pigments: Stabilization Methods of Anthocyanins for Food Applications. Compr. Rev. Food Sci. Food Saf. 2016, 16(1), 180–198. doi: 10.1111/1541-4337.12244
28. Francis, F., & Markakis, P. C. Food Colorants: Anthocyanins. Crit Rev Food Sci Nutr 1989, 28(4), 273-314. doi:10.1080/10408398909527503
29. Stintzing, F. C., & Carle, R. Functional Properties of Anthocyanins and Betalains in Plants, Food, and in Human Nutrition. Trends Food Sci Technol 2004, 15(1), 19-38. doi: 10.1016/j.tifs.2003.07.004
30. Henry, B.S. Natural Food Colours. In Natural Food Colorants, 2nd ed.; Hendry, G. A. F. and Houghton, J. D. Eds.; Springer US: New York, USA, 1996, pp. 39-78.
31. Ioannou, I., Hafsa, I., Hamdi, S., Charbonnel, C., & Ghoul, M. Review of the Effects of Food Processing and Formulation on Flavonol and Anthocyanin Behaviour. J Food Eng 2012, 111(2), 208–217. doi: 10.1016/j.jfoodeng.2012.02.006
32. Jackman, R. L., Yada, R. Y., Tung, M. A., & Speers, R. A. Anthocyanins As Food Colorants a Review. J Food Biochem 1987, 11(3), 201-247. doi:10.1111/j.1745-4514.1987.tb00123.x

References

33. Bakowska-Barczak, A. Acylated Anthocyanins as Stable, Natural Food Colorants – A Review. *Polish J Food Nutr Sci* 2005, 14, 107–116.
34. Giusti, M. M., & Wrolstad, R. E. Acylated Anthocyanins from Edible Sources and Their Applications in Food Systems. *Biochem Eng J* 2003, 14(3), 217–225. doi: 10.1016/S1369-703X(02)00221-8
35. Otake, K., Terahara, N., Saito, N., Toki, K., & Honda, T. Chemical Structures of Two Anthocyanins from Purple Sweet Potato, *Ipomoea batatas*. *Phytochemistry* 1992, 31(6), 2127-2130. doi:10.1016/0031-9422(92)80378-r
36. Ghosh, Dilip & Konishi, Tetsuya. Anthocyanins and Anthocyanin-rich Extracts: Role in Diabetes and Eye Function. *Asia Pac J Clin Nutr* 2007, 16(2), 200-208. doi: 10.6133/apjcn.2007.16.2.01
37. Islam, S. Sweetpotato (*Ipomoea batatas* L.) Leaf: Its Potential Effect on Human Health and Nutrition. *J Food Sci* 2006, 71(2), R13-R21. doi:10.1111/j.1365-2621.2006.tb08912.x
38. Lebot, V. *Tropical Root and Tuber Crops: Cassava, Sweet Potato, Yams and Aroids*, 2nd ed.; CABI: Wallingford, UK, 2019.
39. Miyazaki, T., Tsuzuki, W., & Suzuki, T. Composition and Structure of Anthocyanins in the Periderm and Flesh of Sweet Potatoes. *Engei Gakkai Zasshi* 1991, 60(1), 217-224. doi : 10.2503/jjshs.60.217
40. Shi, Z., Bassa, I., Gabriel, S., & Francis, F. Anthocyanin Pigments of Sweet Potatoes--*Ipomoea batatas*. *J Food Sci* 1992, 57(3), 755-757. doi: 10.1111/j.1365-2621.1992.tb08088.x
41. Sun, H., Mu, T., Xi, L., Zhang, M., & Jingwang, C. Sweet potato (*Ipomoea batatas* L.) Leaves as Nutritional and Functional Foods. *Food Chem* 2014, 156, 380–389. doi: 10.1016/j.foodchem.2014.01.079.
42. Thu, N.N., Sakurai, C., Uto, H., Lien, D.T.K., Yamamoto, S., Ohmori, R., et al. The Polyphenol Content and Antioxidant Activities of the Main Edible Vegetables in Northern Vietnam. *J Nutr Sci Vitaminol* 2004, 50(3), 203–210. doi: 10.3177/jnsv.50.203
43. Truong, V., Mcfeeters, R., Thompson, R., Dean, L., & Shofran, B. Phenolic Acid Content and Composition in Leaves and Roots of Common Commercial Sweetpotato (*Ipomea batatas* L.) Cultivars in the United States. *J Food Sci* 2007, 72(6), C343-C349. doi:10.1111/j.1750-3841.2007.00415.x
44. Xu, W., Liu, L., Hu, B., Sun, Y., Ye, H., Ma, D., & Zeng, X. TPC in the Leaves of 116 Sweet Potato (*Ipomoea batatas* L.) Varieties and Pushu 53 Leaf Extracts. *J Food Compost Anal* 2010, 23(6), 599-604. doi: 10.1016/j.jfca.2009.12.008
45. Yoshimoto, M., Yahara, S., Okuno, S., Islam, M. S., Ishiguro, K., & Yamakawa, O. Antimutagenicity of Mono-, Di-, and Tricaffeoylquinic Acid Derivatives Isolated from Sweetpotato (*Ipomoea batatas* L.) Leaf. *Biosci Biotechnol Biochem* 2002, 66(11), 2336-2341. doi: 10.1271/bbb.66.2336
46. Islam, M.S., Yoshimoto, M., Terahara, N., & Yamakawa, O. Anthocyanin Compositions in Sweetpotato (*Ipomoea batatas* L.) leaves. *Biosci Biotechnol Biochem* 2002, 66(11), 2483–2486. doi: 10.1271/bbb.66.2483
47. Villareal, R. L., Tsou, S. C., Chiu, S. C., & Lai, S. H. (1979). Use of Sweet Potato (*Ipomoea batatas*) Leaf Tips as Vegetables III. Organoleptic Evaluation. *Exp Agric* 1979, 15(02), 123. doi:10.1017/s0014479700000508
48. Otake, K. Characteristics of Food Color Pigments Derived from Ayamurasaki, Proceedings of International Workshop on Sweetpotato Production System Toward the 21st Century, KNAES, Miyazaki, Japan, Dec 9- 10, 1997; Labonte D. R., Yamashita, M., Mochida, H., Eds.; National Agricultural Research Center for Kyushu Okinawa Region. pp. 303–309.
49. Otake, K., Hatanak, A., Kajiwara, T., Muroi, T., Nishiyama, T., Yamakawa, O., Terahara, N., Yamaguchi, M. Evaluation Method and Breeding of Purple Sweetpotato “Yamagawamurasaki” (*Ipomoea batatas*) for Raw Material of Food Colorants. *Nippon Shokuhin Kogyo Gakkaishi* 1994, 41:287–93.

References

50. Philippine Rural Development Project. Available online: [http://www.drive.daprdp.net/iplan/pcip/PCIP%20Tarlac%20Sweetpotato%2020150522%20\(1\)%20\(1\).pdf](http://www.drive.daprdp.net/iplan/pcip/PCIP%20Tarlac%20Sweetpotato%2020150522%20(1)%20(1).pdf) (Accessed 5 May 2018).
51. Philippine Statistics Authority (PSA). Available online: <https://psa.gov.ph/sites/default/files/Major%20Vegetables%20and%20Rootcrops%20Q4%20Bulletin%2C%20October-December%202017.pdf> (Accessed 5 May 2018).
52. Cevallos-Casals, B. A., & Cisneros-Zevallos, L. Stability of Anthocyanin-based Aqueous Extracts of Andean Purple Corn and Red-fleshed Sweet Potato Compared to Synthetic and Natural Colorants. *Food Chem* 2004, 86(1), 69-77. doi:10.1016/j.foodchem.2003.08.011
53. He, X. L., Li, X. L., Lv, Y. P., & He, Q. Composition and Color Stability of Anthocyanin-based Extract from Purple Sweet Potato. *Food Sci Technol* 2015, 35(3), 468-473. doi: 10.1590/1678-457X.6687
54. Rumbaoa, R. G. O., Cornago, D. F., & Geronimo, I. M. Phenolic Content and Antioxidant Capacity of Philippine Sweet Potato (*Ipomoea batatas*) Varieties. *Food Chem* 2009, 113(4), 1133-1138. doi: 10.1016/j.foodchem.2008.08.088
55. Song, J., Li, D., Liu, C., & Zhang, Y. Optimized Microwave-assisted Extraction of Total Phenolics (TP) from *Ipomoea batatas* Leaves and its Antioxidant Activity. *Innov Food Sci Emerg Technol* 2011, 12(3), 282-287. doi: 10.1016/j.ifset.2011.03.001
56. Liao, W. C., Lai, Y.-C., Yuan, M.-C., Hsu, Y.-L., & Chan, C.-F. Antioxidative Activity of Water Extract of Sweet Potato Leaves in Taiwan. *Food Chem* 2011, 127(3), 1224-1228. doi: 10.1016/j.foodchem.2011.01.131
57. Kong, J.-M., Chia, L.-S., Goh, N.-K., Chia, T.-F., & Brouillard, R. Review Analysis and Biological Activities of Anthocyanins. *Phytochemistry* 2003, 64(5), 923-933. doi: 10.1016/S0031-9422(03)00438-2
58. Bhuyan, D. J., Van Vuong, Q., Chalmers, A. C., van Altena, I. A., Bowyer, M. C., & Scarlett, C. J. (2015). Microwave-assisted extraction of *Eucalyptus robusta* leaf for the optimal yield of total phenolic compounds. *Ind Crops Prod* 2015, 69, 290-299. doi: 10.1016/j.indcrop.2015.02.044
59. Binasoy, J. P. Effects of Pasteurization on the Total Phenolic Content, Radical Scavenging Activity, and Physicochemical Characteristics of Sweet Potato (*Ipomoea batatas*) Leaves and Tips Beverage. Bachelor's Thesis, University of the Philippines Diliman, Quezon City, Philippines, March 2011.
60. Kala, H. K., Mehta, R., Sen, K. K., Tandey, R., & Mandal, V. Critical Analysis of Research Trends and Issues in Microwave Assisted Extraction of Phenolics: Have We Really Done Enough. *Trends Analyt Chem* 2016, 85, 140-152. doi: 10.1016/j.trac.2016.09.007
61. Myers, R. H., Montgomery, D. C., & Anderson-Cook, C. M. *Response Surface Methodology: Process and Product Optimization Using Designed Experiments*, 3rd ed.; John Wiley & Sons, INC.; New Jersey, USA, 2009.
62. Villarino, C. B., Jayasena, V., Coorey, R., Chakrabarti-Bell, S., & Johnson, S. Optimization of Formulation and Process of Australian Sweet Lupin (ASL)-Wheat Bread. *LWT* 2015, 61(2), 359-367. doi: 10.1016/j.lwt.2014.11.029
63. McGuire, R. G. Reporting of Objective Color Measurements. *Hortic Sci* 1992, 27 (12), 1254-1255.
64. McLellan, M. R., Lind, L. R., & Kime, R. W. Hue Angle Determinations and Statistical Analysis for Multi-quadrant Hunter L,a,b Data. *J Food Qual* 1995, 18(3), 235-240.

References

- 65.Reyes, L. F., & Cisneros-Zevallos, L. Degradation Kinetics and Colour of Anthocyanins in Aqueous Extracts of Purple- and Red-flesh Potatoes (*Solanum tuberosum* L.). *Food Chem* 2007, 100(3), 885-894. doi: 10.1016/j.foodchem.2005.11.002
- 66.Sadilova, E., Carle, R., & Stintzing, F. C. Thermal Degradation of Anthocyanins and its Impact on Color and In Vitro Antioxidant Capacity. *Mol Nutr Food Res* 2007, 51(12),1461-1471. doi: 10.1002/mnfr.200700179
- 67.Sadilova, E., Stintzing, F., & Carle, R. Thermal Degradation of Acylated and Nonacylated Anthocyanins. *Food Chem Toxicol* 2006 , 71(8), 504-512. doi: 10.1111/j.1750-3841.2006.00148.x
- 68.Loypimai, P., Moongngarm, A., & Chottanom, P. Thermal and pH Degradation Kinetics of Anthocyanins in Natural Food Colorant Prepared from Black Rice Bran. *J Food Sci Technol* 2016, 53(1), 461-470. doi: 10.1007/s13197-015-2002-1
- 69.Mori, K., Goto-Yamamoto, N., Kitayama, M., & Hashizume, K. Loss of Anthocyanins in Red-wine Grape Under High Temperature. *J Exp Bot* 2007, 58(8), 1935-1945. doi: 10.1093/jxb/erm055
- 70.Heredia, F., Francia-Aricha, E., Rivas-Gonzalo, J., Vicario, I., & Santos-Buelga, C. Chromatic Characterization of Anthocyanins from Red Grapes—I. pH Effect. *Food Chem* 1998, 63(4), 491-498. doi:10.1016/s0308-8146(98)00051-x