



Universidade do Minho
Escola de Engenharia



Optimization of the crosslinking process with glutaraldehyde vapor in PVA based electrospun membranes to wound dressings applications

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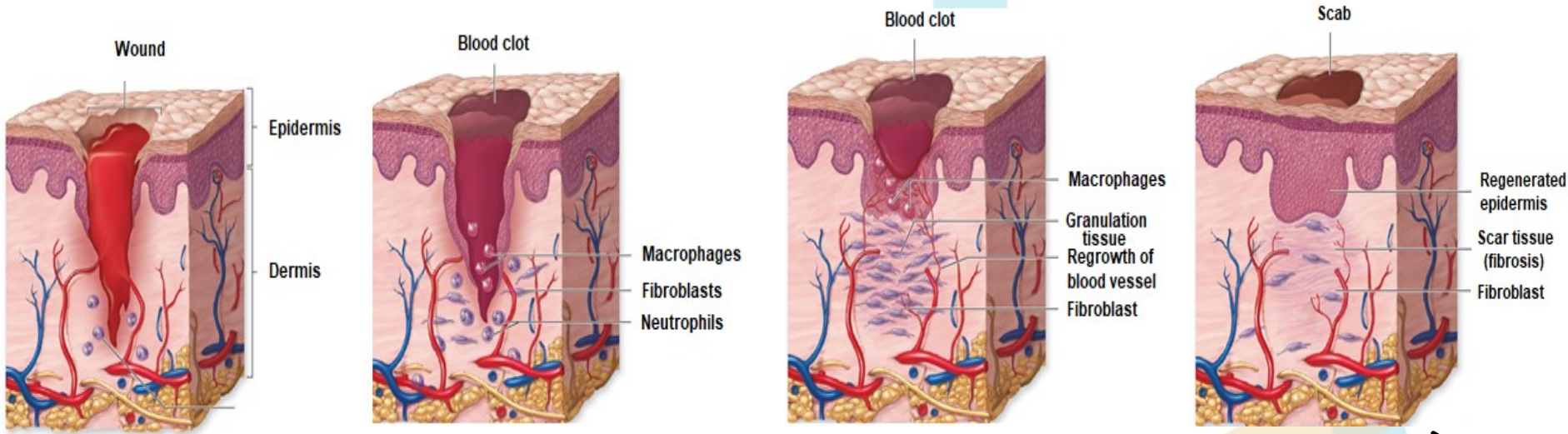


Summary

1. Introduction
2. Methodology
3. Results
4. Conclusions



Acute and Chronic Wounds



**Hemostasis
Phase**

**Inflammatory
Phase**

**Proliferation
Phase**

**Maturation
Phase**

ACUTE WOUNDS

Prolongated state

**CHRONIC
WOUNDS**

- defective cell matrix (high levels of proteases, ROS, etc.);
- high bacteria counts;
- moisture imbalance.



Why engineer new dressings?

Chronic wounds require expensive and time demanding multistep therapies to induce debridement, to fight infection and minimize inflammation.

Films



Not recommended to full thickness, infected or highly exuding wounds

Hydrocolloids



Can cause maceration. Not specified for wounds producing high levels of exudates. Not indicated for infected tissue.

Hydrogels



Not appropriate for wounds with high levels of exudates and infected tissue.

Alginates



May cause dryness and scabbing.

Effective dressings should:

- Allow gaseous exchanges;
- Create a moist environment;
- Be impermeable to microorganisms;
- Remove excess of exudates and prevent desiccation;
- Be non-toxic;
- Provide mechanical protection;
- Be cost-effective;
- Be easy to use.

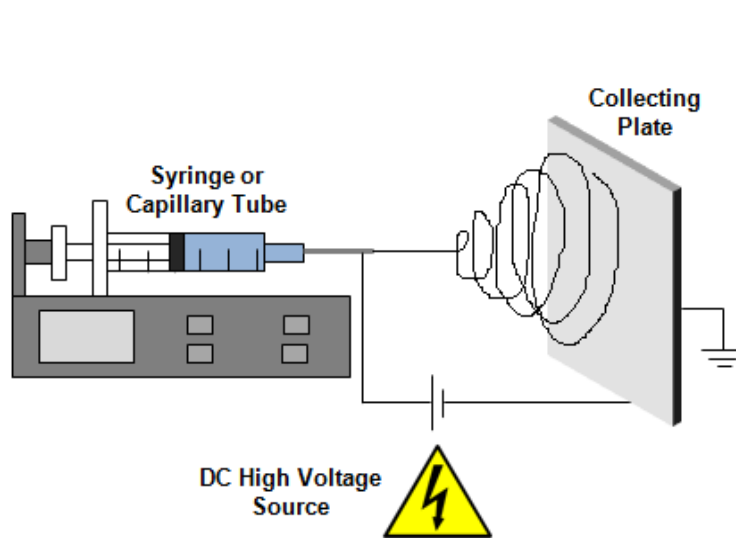


Limitations



Electrospinning technique

Principle: A jet of charged fluid of the dissolved or molten polymer is ejected out of a capillary tube after the electric potential overcomes the surface tension (formation of Taylor cone).



- ✓ Simple and low-cost production;
- ✓ Ultrafine fibers (nanoscale diameters);
- ✓ Similar morphologies to the natural extracellular matrix (ECM);
- ✓ Porosity control;
- ✓ Easy processability of both natural and synthetic polymers;
- ✓ Non-woven;
- ✓ Large surface-to-volume ratio;
- ✓ Allows the introduction of additives (e.g. antimicrobial agents).



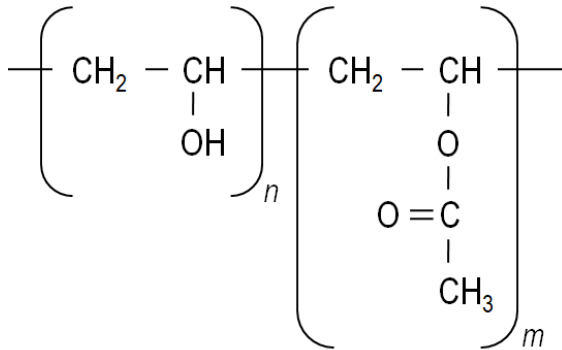
Dressing Production is influenced by:

Solution parameters (type of polymer, solvent, additives, concentration),
processing conditions (applied voltage, spinning distance and feed rate),
 and **surrounding environment** (temperature, humidity and air flow).



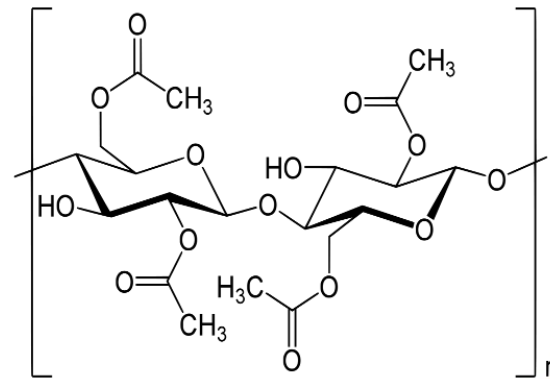
Biodegradable polymers

Poly(vinyl alcohol) (PVA)



- ✓ Food and Drug Administration (FDA)-approved polymer;
- ✓ Biocompatible;
- ✓ Biodegradable;
- ✓ Hydrophilic;
- ✓ Good transparency;
- ✓ Good film forming ability;
- ✓ Thermo-stability and chemical resistance.

Cellulose acetate (CA)



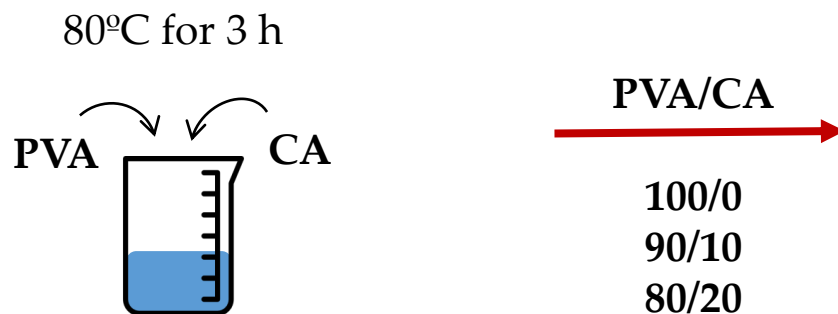
- ✓ Biodegradable;
- ✓ Good mechanical performance;
- ✓ High affinity to other polymers and biomolecules;
- ✓ Good hydrolytic stability;
- ✓ Relative low cost;
- ✓ Excellent chemical resistance;
- ✓ Ability to mimic the ECM to promote cell adhesion.



Methods

Production of meshes

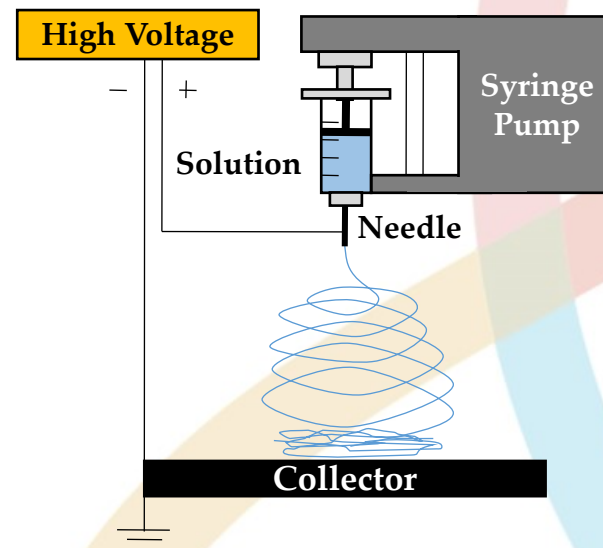
1. Polymeric Solutions



Concentration: **10% (w/v)**

Solvents: **75/25% (v/v) acetic acid/dH₂O**

2. Eletrospinning



The most stable electrospun nanofibers were reached at:

Voltage: 25 kV;

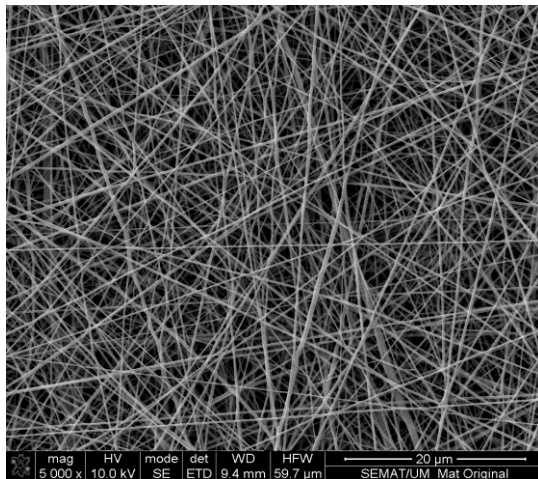
Feed rate: 0.8 mL/h;

Distance between syringe and collector: 18 cm.

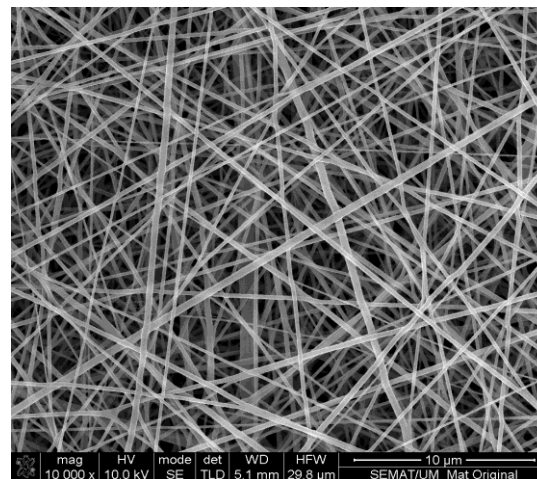


Mats' Morphology

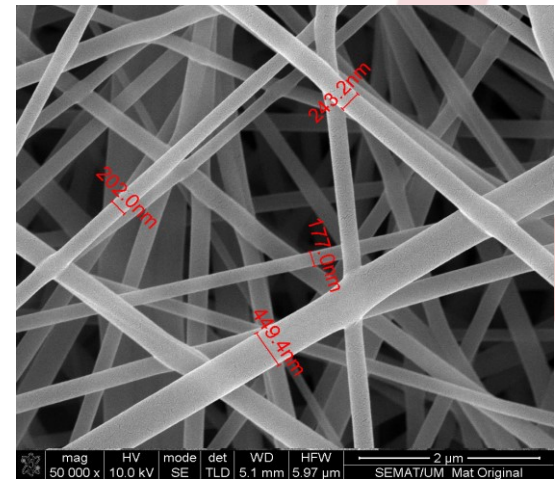
Original Mat: 80/20 PVA/CA; 75/25% (v/v) acetic acid/dH₂O



Mag 5 000 x



Mag 10 000 x



Mag 50 000 x

Average Fiber Diameter (Arithmetic) = 194 ± 51 nm

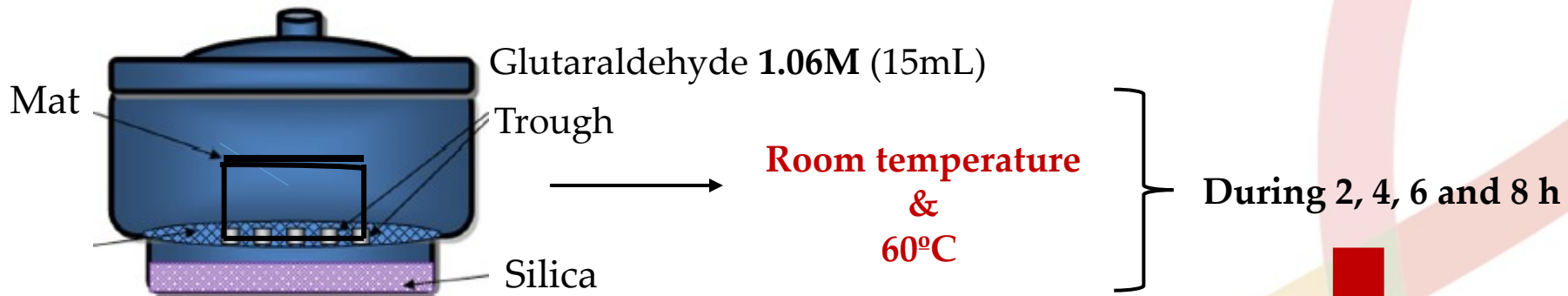
Data obtained using ImageJ (2 images of Mag 50 000 x – 100 measurements each)



Crosslinking Process

➤ 1st trial with 80/20 PVA/CA – Starting point:

Mats were initially dried for 72 h at 40°C



Temperature: 60°C
Glutaraldehyde exposure: 7 h
Drying temperature: 60°C for 24 h

2nd Trial

Crosslinking accomplished after 6 and 8 h at 60°C.
At 8 h mats became yellowish.

Successful crosslinking

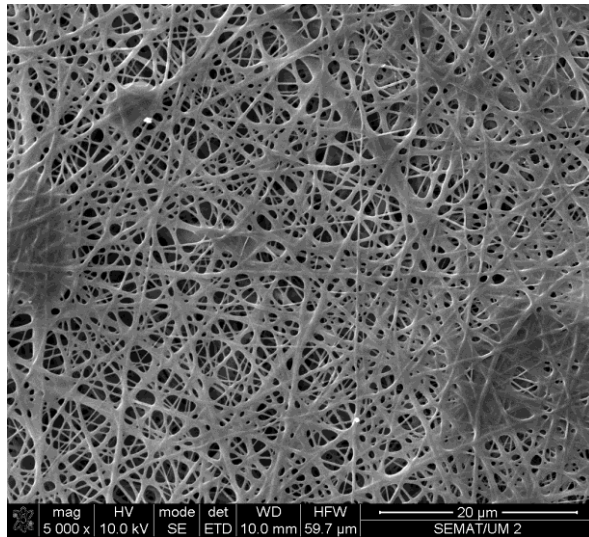


Mats become yellowish

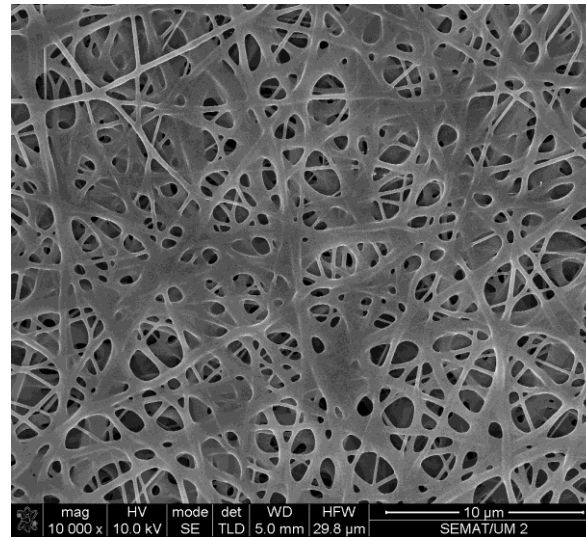


Mats morphology after crosslinking

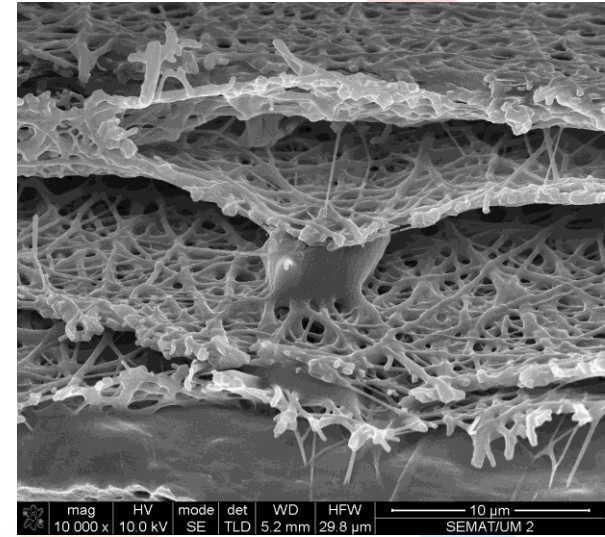
Crosslinking: 60°C; 7h; 15mL GA (**without washings**)



Mag 5 000 x



Mag 10 000 x



Mag 10 000 x

Average Fiber Diameter (Arithmetic) = 343 ± 340 nm

Data obtained using ImageJ (2 images of Mag 50 000 x – 100 measurements each)



Crosslinking Process

3rd Trial:

New Conditions:

Temperature: 60°C

Glutaraldehyde exposure: 7 h

Drying temperature: 45°C for 24 h

How to prevent the yellow color in the drying process?



Successful crosslinking and drying process



How to remove the glutaraldehyde efficiently?

Sonication

RT

Orbital shaker

37°C at 100 rpm

0.26M glycine solution

Orbital shaking at RT
during 30 min

SEM

~~Damaged
structure~~

15 and 30 minutes (100 ml of dH₂O changed every 5 minutes)

1st: Dry at 45°C and 40°C

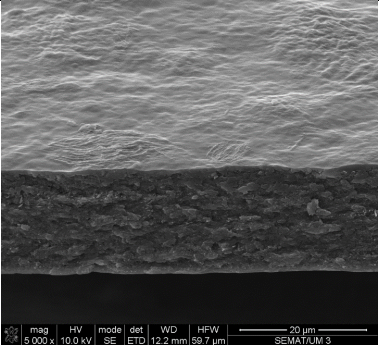
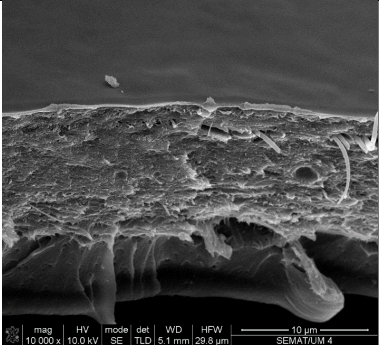
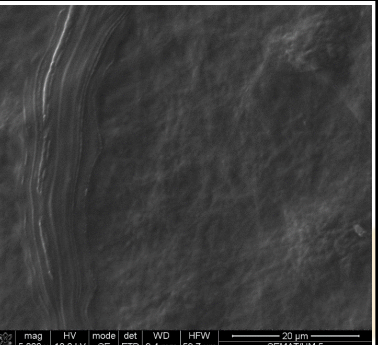
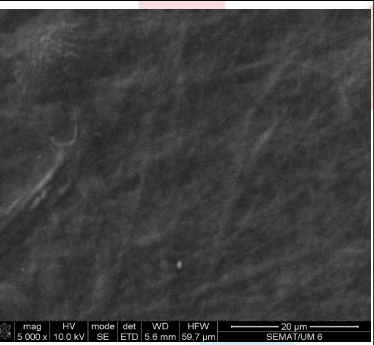


2nd: RT (to avoid the yellow coloration)



Removal of GA excess

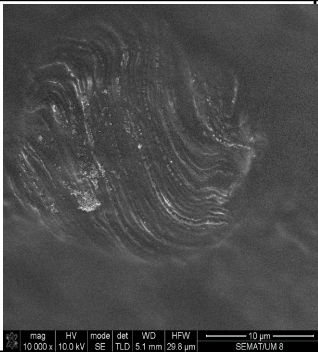
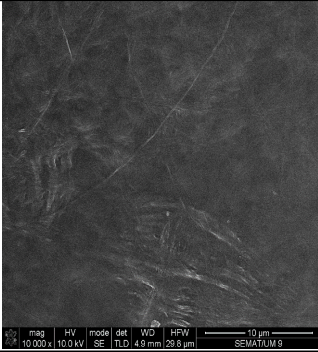
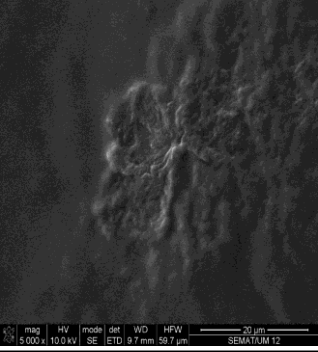
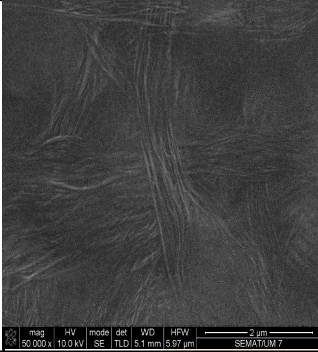
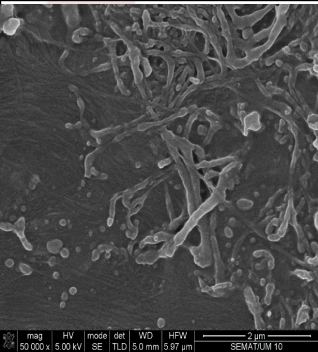
Washings

Process	Sonication	Sonication	Orbital shaker (100 rpm)	Orbital shaker (100 rpm)
Duration (min)*	15	30	15	30
Structure				
Temperature	RT	RT	37°C	37°C

*Every 5 minutes the water changed



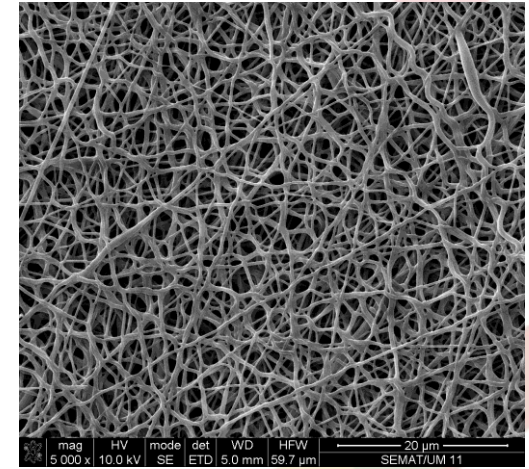
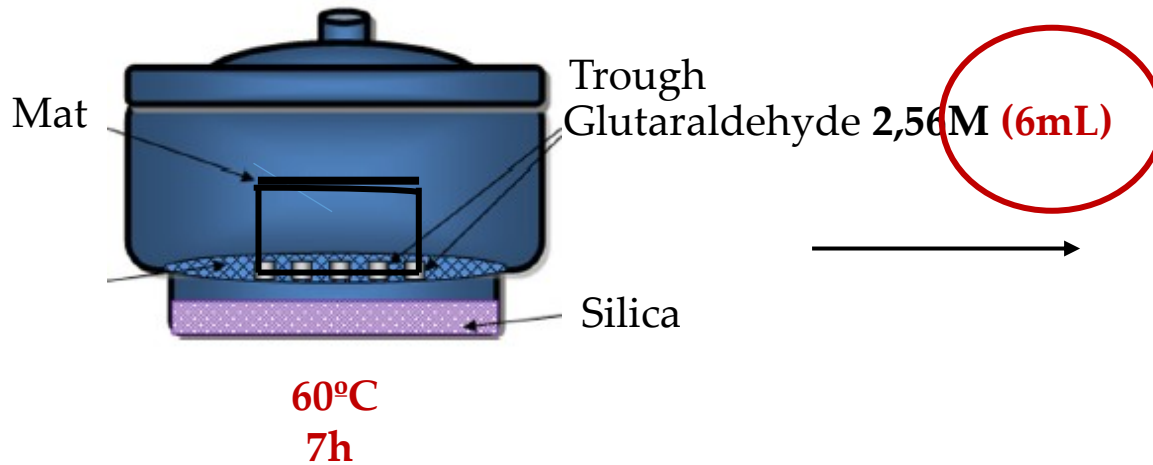
Results and Discussion

Process	Sonication	Orbital shaker (100 rpm)	Orbital shaker (100 rpm)	Orbital shaker (100 rpm)	Orbital shaker (100 rpm)
Duration (min)	30	15	30	30	180
Structure					
Concentration (%)	0.5	0.5	0.5	2.0	0.5
Temperature	RT	37°C	37°C	37°C	37°C

Washings with glycine



The new crosslinking approach method



Successful crosslinking



Without architectural change

Electrospun meshes without compromising their functions



Conclusion

After analyzing and experimenting the various crosslinking and removing excess GA processes, **the amount of GA applied seems to be the simplest and most effective way to attain an effective crosslinking** without harming the structure or turning the surface cytotoxic (due to excess GA).



Acknowledgments

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