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Phytoplasmas:

For a long time researched as plant viruses that caused plant yellowing diseases. Discovered in 1967 by Doi *et*. all.

Plant pathogenic bacteria Reside in plant phloem and insect-host tissues 200-800 nm cell size 530-1350 kbp size genomes Wall-less, polymorphic Gram positive Unculturable in artificial media



http://1.bp.blogspot.com/-63aX2TQr3AY/UNEcJHexG2I/AAAAAAAAQQ/GCejz-JbLV0/s1600/TEM_Phytoplasmas_Jana_Franova.jpg



Phytoplasma taxonomy

- Taxonomy is based on marker gene analysis: mainly of 16S rRNA gene
- Phytoplasmas belong to the phylum Firmicutes, class Mollicutes
- Phytoplasmas as unculturable microorganisms received 'Candidatus (Ca.)Phytoplasma' species status, that was given by the International Research Programme for Comparative Mycoplasmology in 2004
- 44 'Ca. Phytoplasma' species are confirmed to date [1]

Phytoplasma dissemination

- Phytoplasmas in nature are spread by insects: mainly from the order Hemiptera (leafhoppers, planthoppers, psyllids)
- Also they can be transmitted by dodder (parasitic plant) and anthropogenically (grafting, germplasm)







Grafted pine tree

Phytoplasma research methods

- Molecular biology methods: PCR, RFLP, hybridization, qPCR, NGS, etc.
- Immunological: staining (DAPI, Diene's stain), ELISA
- Electron microscopy

Economical impact

Phytoplasmas are known to infect more than 1000 of plant species worldwide.

They are causing epidemics of the agricultural and the industrial plants, thus, they inflict a meaningful impact on economics.



Infected strawberry



Infected common oaks



Control of the phytoplasmas

- Strict quarantine measures
- Agrotechnical methods (physical, biological fencing, application of insect repellents, insecticides, disposal of infected plants, etc.)
- Germplasm thermotherapy
- Antibiotic treatment of plants (tetracycline)



Phytoplasmas infecting gymnosperms

- Pine [2, 5, 12]
- Spruce [2, 3]
- Cycad [7, 8]
- Larch [6]
- Juniper [4]
- Cypress [9]



https://commons.wikimedia.org/wiki/File:BlankMap-World-2009.PNG

Conifers important industrial plants

- The wood of the conifer trees, is an economically important Lithuanian and international export commodity, and is highly valued in the industry
- Conifers are an important item for the landscaping and the gardening
- Phytoplasma infection damages trees and can make them vulnerable to other pathogens and climatic stress







https://www.google.com/url?sa=i&url=https%3A%2F%2Fgardentabs.com%2Fpine-treelandscaping%2F&psig=AOvVaw3CCWWi4JHCkYmJLFDr3yIO&ust=1603314803730000&so urce=images&cd=vfe&ved=0CAIQjRxqFwoTCJC32JyLxOwCFQAAAAAdAAAABAE



Phytoplasma infected mountain pine

'Ca. Phytoplasma pini' in Europe

Croatia [12] Czech Republic [11] Germany [10] Lithuania [2, 12] Poland [11] Spain [10]

Insect-vector unknown!



www.freeworldmaps.net

'Ca. Phytoplasma pini' in Lithuania

'Ca. Phytoplasma pini' was detected in Scots pine (*Pinus sylvestris* L.) [13] and mountain pine (*Pinus mugo* Turra) [2] in Lithuania



https://pt.wikipedia.org/wiki/Transporte_ferrovi%C3%A1rio_na_Litu%C3%A2nia#/media/Ficheiro:Un-lithuania.png

The infestation magnitude of the Curonian Spit mountain pines (*P. mugo*) infected with *'Ca*. Phytoplasma pini' phytoplasmas can be as high as 80% [2]



Goal

 Our goal was to survey mountain pines of the Curonian Spit for the insects-hosts of mountain pine proliferation decline (MPPD) phytoplasma.

Diseased mountain pines (P. mugo)



Symptomatic mountain pine trees exhibiting symptoms of dwarfed needles, proliferation, decline, infected with MPPD phytoplasma





Collected insects

Order Coleoptera
Family Carabidae
Calodromius spilotus III.
Family Curculionidae
Brachyderes incanus L.
Pissodes pini L.
Pissodes validirostis Gyll.
Strophosoma capitatum (De Geer, 1775)
Pissodes piceae (Illiger, 1807)
Pissodes piniphilus (Herbst, 1797)
Family Ptinidae
Ptinus subpilosus Strm.

Order Hemiptera Family Myridae Lygus rugulipennis Poppius, 1911 Family Lygaeidae Gastrodes (Gastrodes) grossipes DeGeer, 1773 Suborder Sternorrhyncha **Family Aphididae** Anoecia (Anoecia) corni (Fabricius, 1775) Cinara sp. Cinara (Cinara) pini (Linnaeus, 1758) Cinara (Schizolachnus) pineti (Fabricius, 1781) Cinara (Cinara) piniphila (Ratzeburg, 1844) Cinara (Cinara) pinihabitans (Mordvilko, 1895) Cinara (Cinara) pinea (Mordvilko, 1895)



Detection and identification of phytoplasmas

- For this work we have collected and tested more than 1000 insect samples
- Phytoplasma 16S rDNA amplicons were amplified from samples of: *Cinara* sp., *Cinara* (*Cinara*) *pini*, *Cinara* (*Schizolachnus*) *pineti*, *Cinara* (*Cinara*) *piniphila*
- 1200 bp size 16S rDNA amplicons from infected mountain pines and aphids were sequenced and used in the virtual restriction fragment length analysis (RFLP), and for RFLP group affiliation using iPhyClassifier online tool

'Ca. Phytoplasma pini' insects-hosts



(A)





(A) Cinara (Cinara) pini
(B) Cinara (Schizolachnus) pineti
(C) Cinara (Cinara) piniphila

(C)

Virtual RFLP

Identification of MPPD phytoplasma 16SrXXI-A phytoplasma subgroup strain based on virtual RFLP analysis of 1.2 kb of 16S rDNA sequence. 16S rDNA amplicons were derived from *C*. *(C.) pini* and *P. mugo* samples. MW – DNA size marker Φx174/HaeIII

*i*PhyClassifier affiliation: The phytoplasmas detected in *Cinara* sp., *Cinara* (*Cinara*) *pini*, *Cinara* (*Schizolachnus*) *pineti*, *Cinara* (*Cinara*) *piniphila* and *P. mugo* samples are variants of 16SrXXI-A phytoplasma subgroup





C. (C.) pini

Conclusions

- 'Ca. Phytoplasma pini' (16SrXXI-A phytoplasma subgroup) for the first time was found in C. (C.) pini, C. (C.) piniphila and C. (S.) pineti insects in Lithuania and worldwide.
- RFLP analysis showed that the PCR-RFLP profile of the positive insect samples was consistent with that of the 'Ca. Phytoplasma pini' from infected pine trees. These results suggest that C. (C.) pini, C. (C.) piniphila and C. (S.) pineti may be potential insect vectors of MPPD phytoplasma.

Acknowledgements, funding

- Acknowledgments: We are very grateful to Dr Rasa Jomantiene and Dr. Povilas Ivinskis for their insight and constructive critics that helped improve our work.
- Funding: This research was funded by the Research Council of Lithuania, grant No. MIP-51/2013.







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Thank you

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