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In Silico Identification of Protein Targets Associated to the Insecticide Activity of Eugenol Derivatives †

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Abstract: The control of insect pests and the need for increased food production due to the world population growth, together with the environmental issues associated to synthetic pesticides, has stimulated the development of new and "greener" alternatives, based on natural compounds. Eugenol is a natural compound that is the major component of clove oil. It has demonstrated antimicrobial and antioxidant activity, being also a powerful insecticide. Recently, new eugenol derivatives have been developed, with some molecules displaying increased insecticide activity. One of the difficulties associated to the rational development of new eugenol derivatives with enhanced insecticidal activity lies in the lack of knowledge of the specific protein target responsible for its activity and to the binding conformation of these molecules. Here, we report the application of an integrated molecular modelling—inverted virtual screening protocol of a collection of eugenol derivatives with confirmed insecticide activity against a molecular library of protein targets typically associated with the insecticide activity of natural compounds. The protocol included 6 different scoring functions from popular docking software alternatives. The results consistently show a marked preference for interaction of the eugenol derivatives with the odorant binding proteins (OBPs) in insect species. Interestingly, OBPs have been regarded as promising targets in the insect periphery nerve system for environmental-friendly approaches in insect pest management. The present results provide clues for the rational development of new eugenol derivatives as bioinsecticides targeting OBPs.

Keywords: biopesticides; Eugenol; Odorant binding proteins; inverted virtual screening

1. Introduction

The increase in population has caused a strain in agriculture due to rising demand and decreased land availability. Crops need to become resistant to damage and disease and the use of pesticides, fungicides and herbicides has allowed for crop protection and long-term storage[1,2]. However, when these chemicals are used extensively or incorrectly, they become hazardous to the environment and to the human health[3]. Thus, the search for new natural, safe, and ecofriendly alternatives, i.e., biopesticides, is being stimulated. Plants, animals, and bacteria produce metabolites that can exhibit

insecticidal activity. Using those metabolites to protect the crops may lead to higher specificity, lower toxicity and even a decrease in pest resistance[4].

Eugenol, the major component of clove oil, has established insecticidal and antimicrobial activity against a variety of targets [5–7]. Consequently, the search for new eugenol derivatives with higher efficiency was boosted to find additional alternatives to known insecticides. However, there is still significant work to be done to find out their correct binding conformation and most importantly, their exact target(s) and mechanism(s) of action. This is precisely where computational chemistry can provide valuable insight.

In this study, it is presented the application of an integrated molecular modelling—inverted virtual screening protocol for the identification of potential protein targets for a series of eugenol derivatives with confirmed insecticide activity. The protocol included the study of protein targets typically associated with the insecticide activity and included 6 different scoring functions from popular docking software alternatives.

2. Methods

A search on Scopus and was performed for papers describing virtual screening (VS) studies involving targets and molecules with insecticidal/herbicide activity. The selection criteria were relevance of the target and year of publication. In the eighteen studies found, fourteen targets were identified and are listed in Table 1.

Table 1. List of targets selected for the inverted virtual screening study.

Target	Organism	PDB Target	Resolution (Å)	Description	Ref.
Ecdysone receptor	*Heliothis virescens*	1R20	3.00	VS based on 1R20 bound to an agonist as a model for the development of a receptor-based pharmacophore model.	[8]
		1R1K	2.90	VS of 2 million compounds against 1R1K, an ecdysone receptor structure bound to its known ligand ponasterone A.	[9]
Chitinase	*Ostrinia furnacalis*	3WL1	1.77	Pharmacophore-based screening using two crystal structures of chitinases: 3WL1 bound to its reaction product and 3WQV bound to an inhibitor.	[10]
		3WQV	2.04		
beta-N-acetyl-D-hexosaminidase OfHex1		3NSN	2.10	VS of the ZINC database to identify OfHex1 inhibitors using 3NSN crystal structure bound to a known inhibitor.	[11]
		3OZP	2.00	VS of the ZINC database targeting 3OZP, a crystal structure of OfHex1 bound to an inhibitor.	[12]
N-Acetylglucosamine-1-phosphate uridyltransferase (GlmU)	*Xanthomonas oryzae*	2V0K	2.30	Homology model built for docking using 2V0K and 2VD4 as templates. 2V0K crystal structure is bound to its known ligand and 2VD4 is bound to a possible inhibitor.	[13]
		2VD4	1.90		
Acetylcholinesterase	*Aedes aegypti*	1QON	2.72	Search for new molecules with insecticidal activity against *Ae. Aegypti* using acetylcholinesterase crystal structures 1QON and 4EY6 as targets, both bound to possible inhibitors.	[14]
		4EY6	2.40		
	Drosophila melanogaster	1DX4	2.70	Homology 3D model built for VS using 1DX4 as template.	[15]

				1DX4 crystal structure is bound to a potent inhibitor.	
Polyphenol oxidase	*Ipomoea batatas*	1BUG	2.70	Docking simulations using the homologous polyphenol oxidase crystal structure of sweet potato in complex with phenylthiourea, a commonly used pesticide.	[16]
p-hydroxyphenylpyruvate dioxygenase	*Arabidopsis thaliana*	6ISD	2.40	Development of a receptor-ligand pharmacophore model based on the crystal structure 6ISD bound to a commonly used pesticide. The best model created was then used for VS studies.	[17]
Oxidoreductase	*Spinacia oleracea*	1YVE	1.65	Crystal structure of a plant oxyreductase, 1YVE bound to its cofactor, NADPH used in VS assays to find new inhibitors.	[18]
Voltage-gated sodium channel	*Periplaneta americana*	6A95	2.60	Crystallographic structure of a Voltage-gated sodium channel NavPaS bound to a pore blocker, tetrodotoxin (TTX)	[19]
Octopamine receptor	*Blattella germanica*	4N7C	1.75	Crystal structure of Bla g 4, an octopamine receptor, bound to tyramine.	[20]
Sterol carrier protein-2 (HaSCP-2)	*Helicoverpa armigera*	4UEI	Solution NMR	Structure-based VS of a database of commercially available compounds to find potential inhibitors of HaSCP-2. The residues Phe53, Thr128, and Gln131 were selected for the binding cavity.	[21]
Peptide deformylase	*Xanthomonas oryzae*	5CY8	2.38	Docking and VS of a library of 318 phytochemicals. 5CY8 crystal structure is bound to a possible inhibitor.	[22]
Alpha-esterase-7 (αE7)	*Lucilia cuprina*	5TYJ	1.75	Computational design of potent and selective covalent inhibitors of αE7. 5TYJ and 5TYP crystal structures are bound to inhibitors: (3-bromo-5-phenoxylphenyl)boronic acid and (3-bromo-4-methylphenyl)boronic acid respectively.	[23]
		5TYP	1.88		
Odorant Binding Protein	*Aedes aegypti*	5V13	1.84	Search for new molecules with insecticidal activity against *Ae. Aegypti* using a crystal structure of a mosquito juvenile hormone-binding protein, 5V13 bound to its natural hormone.	[14]
	Drosophila melanogaster	2GTE	1.40	2GTE crystal structure is bound to its natural ligand	[24]
	Anopheles gambiae	3N7H	1.60	QSAR and docking studies for the rational design of mosquito repellents using the crystal structure 3K1E bound to a polyethylene glycol molecule. 3N7H crystal structure is bound to a commonly used repellent.	[25]
	Aedes aegypti	3K1E	1.85		[25]

Eugenol and eleven derivatives (Figure 1 EU1-EU3e) were selected as new potential insecticides. These molecules have been previously synthesized and validated experimentally with good insecticidal activity.

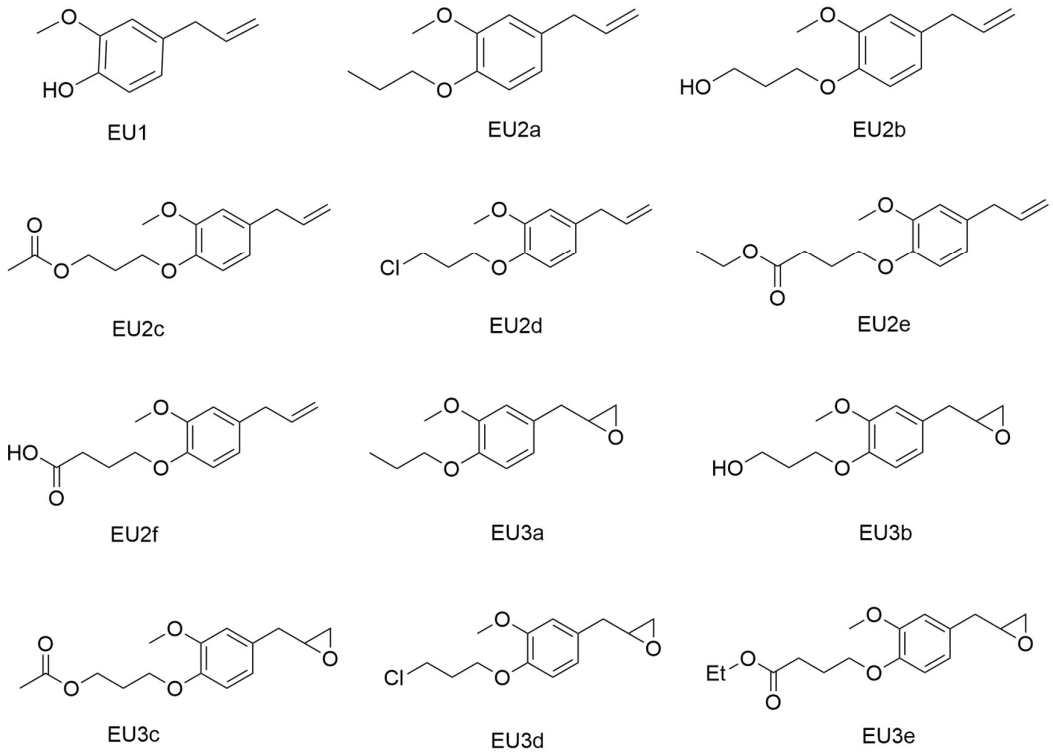

Figure 1. of eugenol and derivatives used in this study.

Each PDB structure was prepared for docking using the Autodock Vina plugin for Pymol[26]. Crystallographic waters and cofactors were removed. The ligands were extracted and saved in separate files to be used for the re-docking and as reference site for the docking coordinates. When there were no crystallographic ligands present, a selection based on the most important active site residues was made. Re-docking was used to evaluate the ability of the docking software to reproduce the geometry and orientation of the crystallographic pose as well as the quality of the docking protocol, and to optimize the docking protocol.

The docking programs/scoring functions used were GOLD[27] (PLP, ASP, ChemScore, and GoldScore scoring functions), AutoDock Vina[28] and LeDock[29]. With each docking program/scoring function the protocol was optimized for each protein target, to minimize the rmsd in the docking predictions of the reference ligand in redocking, by comparison with the crystallographic structure of the corresponding complex.

The optimized parameters for each program/scoring function were: Vina—docking box position, docking box dimension, exhaustiveness; LeDock—docking box position, docking box dimension; GOLD (PLP, ASP, ChemScore, GoldScore)—binding pocket center, docking region radius, search efficiency, number of runs. The final optimized conditions were used for the subsequent stages. Eugenol and derivatives were prepared for docking using Datawarrior[30] and OpenBabel[31] and were docked into each structure with the optimized protocol with all the six scoring functions. A ranked list was prepared based on the average scores of each target.

3. Results and Discussion

Table 2 presents the average scores obtained for of all the eugenol derivatives for each potential target with each scoring function. The score for all the GOLD scoring functions is dimensionless and the higher the score, better the binding affinity. Vina and LeDock scoring functions, on the other

hand, use a metric that is a more precise approximation of binding free energy, so a more negative value means better affinity.

Table 2. Average eugenol derivate scores obtained for all PDB structures with the six different scoring functions.

Target	PDB	PLP	ASP	ChemScore	GoldScore	Vina	Ledock
Ecdysone receptor	**1R20**	57.3	27.5	28.1	52.5	−6.4	−4.7
	1R1K	59.3	26.4	28.3	54.5	−7.1	−5.2
Chitinase	**3WL1**	63.0	40.8	30.1	60.0	−6.9	−4.8
	3WQV	63.4	40.7	30.6	55.7	−6.5	−4.3
beta-N-acetyl-D-hexosaminidase OfHex1	**3NSN**	66.7	46.7	29.1	62.8	−6.1	−4.4
	3OZP	63.3	43.7	28.3	58.7	−7.1	−4.3
N-Acetylglucosamine-1-phosphate uridyltransferase (GlmU)	**2V0K**	55.0	24.1	23.3	54.3	−5.9	−4.6
	2VD4	46.9	22.2	21.6	43.8	−5.2	−3.7
Acetylcholinesterase	**1QON**	73.3	48.2	35.3	62.2	−7.6	−5.0
	4EY6	72.6	41.2	32.4	55.2	−7.1	−5.0
	1DX4	70.0	43.2	32.2	55.3	−7.2	−4.9
Polyphenol oxidase (PPO)	**1BUG**	56.7	27.2	25.9	56.2	−5.2	−4.1
p-hydroxyphenylpyruvate dioxygenase	**6ISD**	57.9	31.6	24.8	47.8	−6.3	−4.3
Oxidoreductase	**1YVE**	66.0	25.6	32.1	59.5	−6.3	−5.2
Voltage-gated sodium channel	**6A95**	53.1	23.6	22.3	56.5	−5.8	−4.5
Octopamine receptor	**4N7C**	68.1	37.9	35.1	65.2	−7.1	−4.5
Sterol carrier protein-2 (HaSCP-2)	**4UEI**	54.1	28.2	29.4	45.8	−6.4	−4.9
Peptide deformylase	**5CY8**	64.0	26.4	24.3	62.5	−6.8	−5.6
α-esterase-7	**5TYJ**	62.9	34.6	29.3	52.1	−6.4	−4.3
	5TYP	59.9	35.2	29.4	53.1	−6.4	−4.8
Odorant Binding Protein	**5V13**	72.1	43.2	35.9	59.4	−7.6	−5.1
	2GTE	63.1	33.8	34.3	56.9	−6.5	−3.1
	3N7H	64.8	34.5	28.9	56.6	−6.3	−4.6
	3K1E	73.4	39.6	35.8	62.4	−6.0	−5.5

Overall, the results show good consistency, with odorant binding proteins, acetylcholinesterases, octopamine receptors and chitinases yielding better scores. On the other hand, targets such as voltage-gated sodium channels, sterol carrier protein-2 (HaSCP-2) and N-Acetylglucosamine-1-phosphate uridyltransferase (GlmU), are consistently presenting lower scores for all scoring functions.

The structure with the best score was selected for each potential target and they were ranked from the best target to worst, according to the predictions of the different docking programs/scoring functions. The results are listed in Table 3. Globally, considering the results obtained with the several scoring functions, odorant binding proteins are the target with the highest affinity towards eugenol derivatives, followed closely by acetylcholinesterase, chitinases and octopamine receptors. Enan in 2001[5] suggested that the insecticidal activity of eugenol was mediated by octopamine receptors. Our study implies that there might be other targets involved as well, as the binding affinity of eugenol derivates was higher for OBPs and acetylcholinesterase.

Some variations between the predictions of different scoring functions exists. For example, for the PLP and ChemScore scoring function, odorant binding proteins and acetylcholinesterase come in first and second as preferable targets for eugenol derivates. However, for ASP and Vina, the preferable target is the acetylcholinesterase, and for both Vina and LeDock, odorant binding proteins are the second preferable target. The discrepancy is even higher for GoldScore, with odorant binding proteins coming in 3rd place and octopamine receptors presenting the highest binding affinity for eugenol derivates. This may be explained by the own nature of each scoring function, as they consider different aspects of protein-ligand binding.

Table 3. Ranking of Targets obtained with the different docking programs/scoring functions.

Ranking	PLP	ASP	ChemScore	GoldScore	Vina	LeDock	Overall Ranking
Odorant Binding Protein	1	4	1	3	2	2	1
Acetylcholinesterase	2	1	2	5	1	5	2
Chitinase	4	2	5	2	6	7	3
Octopamine receptor	3	5	3	1	5	10	4
Peptide deformylase	6	11	12	4	7	1	5
Oxidoreductase	5	12	4	6	11	4	6
β-N-acetyl-D-hexosaminidase OfHex1	7	3	9	7	3	13	7
Ecdysone receptor	9	9	8	10	4	3	8
α-esterase-7	8	6	7	12	9	8	9
Sterol carrier protein-2 (HaSCP-2)	13	8	6	14	8	6	10
p-hydroxyphenylpyruvate dioxygenase	10	7	11	13	10	12	11
Polyphenol oxidase (PPO)	11	10	10	9	14	14	12
N-Acetylglucosamine-1-phosphate uridyltransferase (GlmU)	12	13	13	11	12	9	13
Voltage-gated sodium channel	14	14	14	8	13	11	14

The consistency of the results was visually confirmed by the analysis of the corresponding poses. The hypothesis formed is that eugenol and eugenol derivatives can be used as repellents because they can bind to odorant binding proteins or used as pesticides, inhibiting insect acetylcholinesterase. As observed in Figure 2, they are very different targets, both in size and in function.

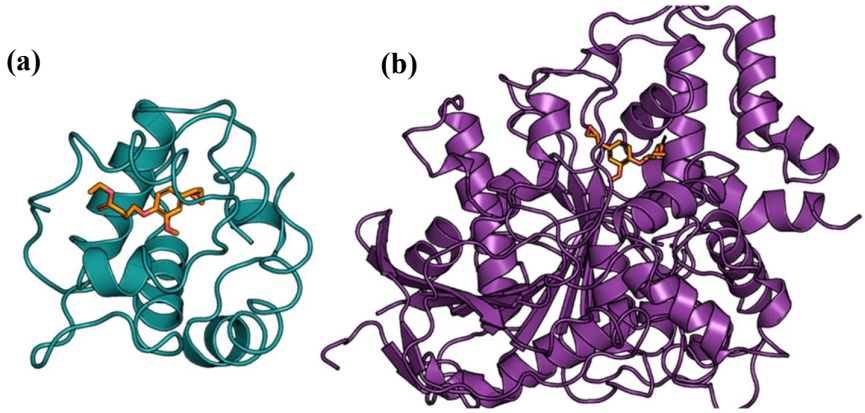

Figure 2. Docking-predicted binding mode of EU3e to OBPs (**a**) and Docking-predicted binding modes of EU3e to Acetylcholinesterase (**b**) with PLP scoring function.

Odorant binding proteins (OBPs) are a large family of insect proteins that are crucial for species survival and reproduction, as they use pheromones, plant volatiles and other odorant molecules to mate, find food and avoid predators[32]. OBPs are present in a variety of organisms, are highly expressed and highly divergent in sequence. They do however, present a few common features such as their small size and the presence of six conserved cysteines[33]. These features also make them good targets for rapid screenings. There is not enough consensus regarding the specificity of these proteins and further studies must be performed to better understand the sensitivity of OBPs[2].

Acetylcholinesterases (AChE) are one of the most common targets of the synthetic pesticides such as organophophates and carbamate [34] and has been a target of reference for over 50 years. This enzyme is a serine hydrolase and is responsible for regulating the levels of acetylcholine in a variety of organisms, from mammals to insects[35]. Due to its extensive "attack", some pests have become resistant to organophosphates and the search for new and effective alternatives is currently being promoted [36].

Insterestingly, during a search in the Protein Data Bank for eugenol, a structure of an odorant binding protein was found complexed with eugenol was found. It is an OBP of *Apis mellifera* (PDB: 3S0E) that exhibits high affinity for eugenol[37]. This reinforces the proposed theory that eugenol and derivatives can, in fact, bind to OBPs and could potentially work as repelents. Still, additional computational and experimental studies need to be performed to further optimize and develop this hypothesis.

4. Conclusions

In the present study, we report the application of an integrated molecular modelling—inverted virtual screening protocol of a collection of eugenol derivatives in order to find possible protein targets in which they present insecticidal activity.

First, we explored the literature for other virtual screening studies performed on known targets to minimize the candidate pool. Of 18 studies found, 14 targets were selected to continue the study. After careful optimization of the VS protocol, the eugenol derivatives were docked into each target with six different scoring functions (PLP, ASP, ChemScore, GoldScore, Vina and LeDock). The consistency of the scores was evaluated and a ranked list of most likely targets was created.

Eugenol derivates showed an increased binding affinity for odorant binding proteins and acetylcholinesterases. Since there is, already, in the PDB database a structure of an OBP bound to

eugenol not considered in the VS, it reinforces the proposal that eugenol derivatives can potentially be used as repellents.

This work presents a simple approach for the application of inverted virtual screening in identification of possible targets for new insecticides.

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