

Proceedings

# Performance of a Specialist and a Generalist Herbivorous Moth on Different *Quercus robur* Genotypes <sup>†</sup>

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**Abstract:** Besides beech, oak is one of the ecologically and economically most important deciduous tree species in Central European forests. Due to progressive global warming, oaks are increasingly exposed to biotic and abiotic stress factors. In addition to drought and heat, thermophilic herbivorous insects also represent a challenge for the oak trees. During earlier investigations on *Tortrix viridana* in North Rhine-Westphalia, differences in the infestation intensity of individual pedunculate oaks (*Quercus robur*) by herbivorous insects, leads to a definition of tolerant (T-oaks, less infested) and susceptible (S-oaks, heavily infested) oaks. Performance studies with *T. viridana* and *Lymantria dispar* were conducted to validate the observations. Both herbivorous insects needed more time for their juvenile development when reared on T-oaks. While larvae of *L. dispar* tended to eat less leaf material from T-oaks than from S-oaks, *T. viridana* needed to consume more T-oak leaf material to achieve approximately the same pupal weight as larvae fed with leaf material from S-oaks. The mortality rate of *T. viridana* was significantly higher on T-oaks than on S-oaks, whereas no significant difference was found for *L. dispar*. Basically, T-oaks seem to offer a poorer nutritional basis for both kinds of leaf-eating insects.

**Keywords:** *Tortrix viridana*; *Lymantria dispar*; caterpillar; *Quercus robur*; performance; plant-herbivore-interaction; plant resistance

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## 1. Introduction

Covering 32% of the national territory, forests are a major part of Germany's landscape. They play a decisive role in climate and culture not only because they are an important carbon sink [1], but also as part of the water cycle, as an oxygen and wood producer, a habitat for many animal and plant species, a recreational area for humans and much more [2,3]. Poor silvicultural practices of the past together with the adverse anthropogenic effects have posed many challenges for forests, an effect, which is likely to increase. Among these challenges are temperature changes due to global warming, increasingly frequent drought and precipitation events [4] as well as evolving threats from new pests and diseases [5]. After beech, oak is the second most common deciduous tree genus in European forests. The most common oak species in central Europe are the pedunculate oak (*Q. robur*) and the sessile oak (*Quercus petraea*). They account for up to 10% of all trees in German forests [6]. Together with beech wood, oak has a high value as construction timber. In addition to soft coniferous woods, straight deciduous woods are particularly valued for their robustness and hardness. For example, railroad sleepers, constructions for half-timbered houses and pile dwellings are made of oak wood.

Interaction between trees and herbivorous insects are among the driving factors of co-evolutionary processes in the forest ecosystems [7,8]. *Q. robur* hosts a large number of herbivorous insects. Beside the gypsy moth (*L. dispar*) and the green oak leaf roller (*T. viridana*), also the small (*Operophtera brumata*) and the large (*Erannis defoliaria*) winter moth as well as the oak processionary

(*Thaumetopoea processionea*) belong to the oak feeding society, just to name a few. In years with mass occurrence of larvae of various herbivorous caterpillars, the oak is exposed to additional stressors. During a calamity of the pest, this can cause a growth loss of up to 30% (resources are needed for the budding of new leaves), a premature St. John's shoot and even the failure to fruit. In the worst case, oak dieback occurs [9,10] but this usually only occurs in connection with further weakening of the oak (fungal attack or severe drought) [11–13].

*T. viridana* is an oligophagous specialist. It occurs only on oak trees (usually *Q. robur* and sometimes *Q. petraea*) and cannot develop on other deciduous trees. In contrast, *L. dispar* as a generalist can develop on many different types of trees and shrubs. Both pests are common in Central Europe and infest deciduous forests.

In the present study, we compared the performance of *T. vidiana* and *L. dispar* on two different oak types defined as tolerant oaks (T-oaks: less defoliated during outbreak events) and susceptible oaks (S-oaks: heavily defoliated during outbreak events) in an earlier study [14]. In the same study, we showed that T- and S-oak leaves differ in their composition of polyphenolic constituents and their blend in VOCs (volatile organic compounds), leading us to the hypothesis that T- and S-oaks follow different strategies. Where T-oaks count on a constitutive high level of defence compounds, S-oaks react just in time emitting typical 'cry for help' [15] HIPVs (herbivore induced plant volatiles). We already could show that larvae of the green oak leaf roller needed to eat more T-oak leaf material to gain the same pupal weight as larvae fed with S-oak leaves [14,16]. Thus, here we present the comparison of performance studies with a specialist and a generalist herbivorous moth.

## 2. Experiments

### 2.1. Plant and Caterpillar Material

During an outbreak of the green oak leaf roller (2003–2005), oak trees in North Rhine-Westphalia were classified as susceptible or tolerant to the damage caused by the caterpillar of *T. viridana* [14,17]. In order to conduct further analysis, genetically comparable plant material was required. For this purpose, the summer grafting [17] was carried out. Young shoots of the classified oak trees were taken from the original stand in 2008, 2011 and 2015 and grafted onto 2-year-old seedlings of *Q. robur*. The oaks were wrapped in Crispac bags in spring to prevent foreign insects from settling on the trees. The bagged trees are brought to a shady greenhouse so that a slow acclimatization to the increasing sunlight could take place.

Breeding of larvae of *T. viridana* was established in 2007 with caterpillars from 15 stands in North Rhine-Westphalia and since then was continuously genetically refreshed with larvae from other oak stands.. Two clutches of *L. dispar* eggs were kindly provided by ARätin Andrea Stradner from University of Natural Resources and Life Sciences, Vienna.

### 2.2. Feeding Experiment with *Tortrix viridana*

307 freshly hatched larvae of *T. viridana* were placed on opening buds of either T-oaks (150) or S-oaks (157) with a fine brush and isolated with a Crispac bag. These bags also prevented the larvae from breaking out. The larvae were individualized by an assigned number. Under constant conditions (12–13 °C and 10 h/14 h light/darkness) in a climate chamber the larvae could develop on the trees until their pupation.

From larval stage 3 onwards, the larvae were weighed in a 4-day cycle and the leaf area eaten was determined with the help of a scalepaper. The larvae were checked daily in order to document the changes into next larval stages. The fresh pupae were transferred into Petri dishes and kept under the same conditions until the adult moth hatched. A statistical analysis was performed using the t-test for developmental time and amount of eaten leaves, and the Chi2-test for overall mortality. For details see Ghirardo et al. [14] and Schroeder et al. [16].

### 2.3. Feeding Experiment with *Lymantria dispar*

The newly hatched larvae of *L. dispar* were set upon a piece of wheat germ medium developed by Magnoler [18]. After reaching the 3rd instar the caterpillars were separately placed on S- or T-oak trees in the field to adapt them to leaf feeding. By reaching the 4th instar the larvae were taken into the experiment. Under constant conditions (20 °C ± 5 an 16 h/8 h light/darkness) the larvae were fed in plastic boxes with detached leaf material. The development was documented until pupation.

A total of 160 larvae were used for the experiment. They were divided into two groups, one fed with T-oak and the other with S-oak leaves. The two groups were further divided into two subgroups. One half of each group was fed with leaves from pre-infested trees and the other half with leaves of non-infested trees. For pre-infestation at least 25 larvae of *L. dispar* were put on single T- or S-oaks for a minimum of two days before starting the feeding experiment. The pre-infested trees hereafter referred to as ‘Damaged-trees’, while those that were left non-infested are referred to as ‘Control-trees’. The groups are abbreviated as follows: T\_C (T-oaks control) = fed with leaves from T-oaks without pre-infestation; S\_C (S-oaks control) = the same as before but fed with leaves from S-oaks; T\_D (T-oaks damaged) = fed with leaves from pre-infested T-oaks; S\_D (S-oaks damaged) = the same as before but fed with leaves from pre-infested S-oaks.

The larvae were monitored daily and fed with fresh leaves at least every fourth day, but always ensuring that they were adequately fed. Images of leaves were taken with a Canon Lide 210 scanner and Canon MP Navigator EX software. Leaves were scanned before and after feeding. Analysis of leaf images was performed using RStudio software [19,20] and the package ‘raster’ [21]. Comparison of images of old leaves (uneaten and lying in the plastic box for some days) and the respective image of the fresh leaf revealed a median difference in leaf-area of 1% (shrinking of leaves). For the *L. dispar* data statistical analysis was performed using RStudio Software [19,20] and the packages ‘PMCMRplus’ [22], ‘asbio’ [23]. Visualization of the results was performed using the package ‘ggplot2’ [24].

### 3. Results

#### 3.1. Developmental Time

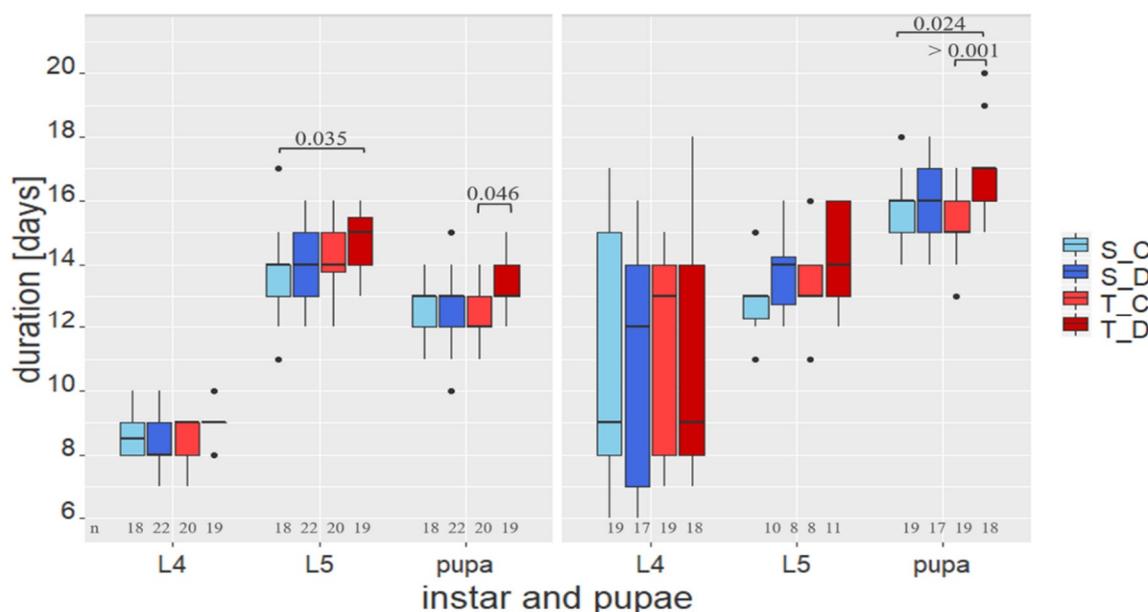
A calculation over all larval stages of *T. viridana* showed no significant differences in the developmental duration between larvae reared on T-or S-oaks. But, the duration time in the first two instars is significantly higher (t-test for L1  $p = 0.01$ ; for L2  $p = 0.02$ ) for larvae fed on T-oaks than on S-oaks. Interestingly, for the fourth instars it is the other way round. In this stage, larvae fed on S-oaks needed significantly more time (t-test  $p = 0.006$ ) to reach the next stage than larvae reared on T-oaks (Table 1), [16].

**Table 1.** Developmental time for *Tortrix viridana* on S- and T-oaks (time in days).

	L1	L2	L3	L4	L5	Pupa	Total
S	4.99	7.74	5.86	6.34	12.11	22.88	59.92
T	5.12	8.31	6.18	5.24	11.55	23.15	59.55

The sexes of *L. dispar* differ much more than those of *T. viridana*. Thus, for the gypsy moth all analyses have been done separately for males and females. The number of instars until pupation differed within and between the sexes. The female larvae pupated after the fifth instar, whereas the males nearly equally pupated after the fourth or the fifth instar, respectively.

The duration of the instars and pupal stage is shown in Figure 1. Both, females and males showed no significant difference in the duration of the fourth instar. The fifth instar of the females lasted significantly longer for the T\_D group than those of the S-oak control group (S\_C), while the males showed no significant differences. The pupal stage in males lasted significantly longer in the T\_D group than in both control groups (T\_C and S\_C). Whereas female pupae showed a significantly slower development only for T\_D compared to T\_C (Figure 1).



**Figure 1.** Duration of the instars and the pupal stage of *Lymantria dispar*. Boxplots colours indicate the different groups, S\_C: S-oaks control; S\_D: S-oaks damaged (pre-infested), T\_C: T-oaks control, and T\_D: T-oaks damaged. Data is grouped by sex: females (left), males (right), and by larval stage: L4 (fourth instar), L5 (fifth instar) and pupa. Bars with numbers above boxplots indicate significant differences between groups and p value from Nemenyi’s non-parametric all-pairs comparison. Numbers below show the numbers of observations for each boxplot.

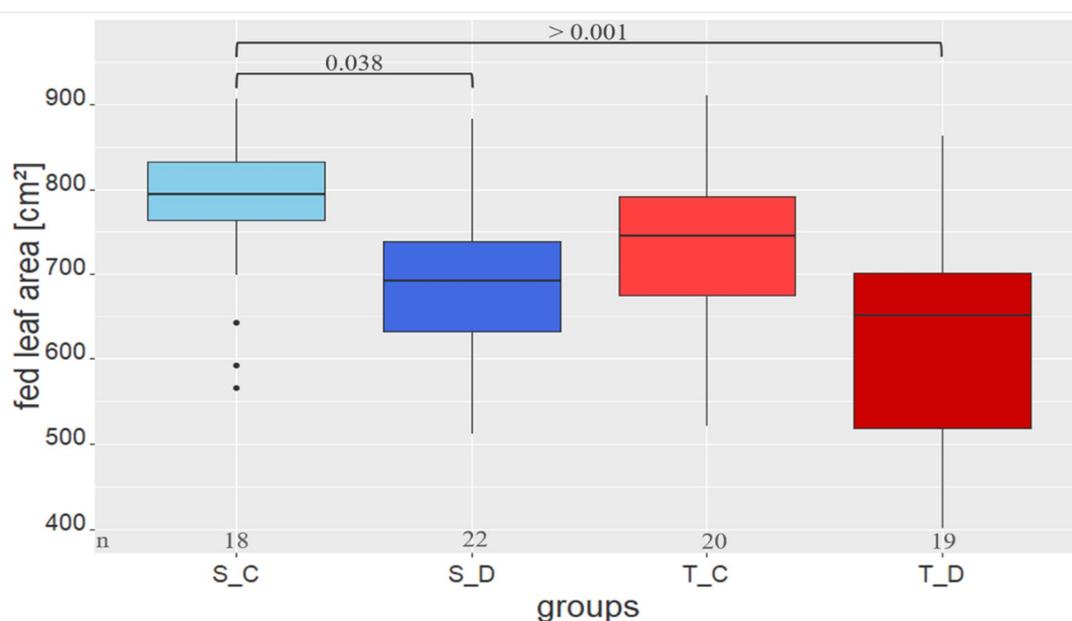
### 3.2. Leaf Area Fed

The amount of leaf area fed was observed from the third until the fifth instar for *T. viridana*. There are no significant differences in the leaf area fed between T- and S-oaks in the third and fourth instars. But, the leaf area fed differed significantly in the fifth instar (t-test  $p = 0.04$ ) and also when looking at all instars (t-test  $p = 0.02$ ). Larvae reared on T-oaks ate nearly 10% more leaves than larvae on S-oaks (Table 2) [16]. Even more important is the fact that larvae of *T. viridana* fed with T-oak leaves needed to consume significantly more leaf material to gain the same pupal weight than larvae fed with S-oak leaves [14].

**Table 2.** Amount/area of leaves fed (in mm<sup>2</sup>) on S- and T-oaks in total and within the stages.

	L3 to L4	L4 to L5	L5 to Pupa	Total
S	58.9	165.4	875.3	1099.6
T	66.9	175.0	968.8	1210.7

For *L. dispar*, the total leaf area fed was measured from the fourth instar until pupation. Female larvae of the S\_C group ate significantly more leaves than the larvae of the S\_D (posthoc.kruskal.nemenyi.test  $p = 0.038$ ) and T\_D (posthoc.kruskal.nemenyi.test  $p > 0.001$ ) groups, while the larvae of the T\_C group did not differ from the others (Figure 2).



**Figure 2.** Total leaf area fed by the female larvae from fourth instar until pupation. Abbreviations and explanations are as in Figure 1.

For the males none of the groups showed significant differences, but there was a tendency for the T\_D group larvae to eat slightly less than the others. Overall, the females ate much more than the males during the experiment.

### 3.3. Mortality Rate

From the originally 157 hatched larvae of *T. viridana* placed on S-oaks, 59 individuals completed their development to adult moths. On T-oaks only 40 individuals out of the original 150 reached the adult stage. That means a significantly different overall mortality rate of 62.4% for larvae reared on S-oaks and 73.3% for larvae reared on T-oaks ( $\chi^2 p < 0.05$ ) (Table 3). On both oak types, mortality rate was highest within the first two larval stages followed by the pupal stage with 10.8% in S-oaks and 14.7% in T-oaks (Table 3, [16]).

**Table 3.** Mortality of *Tortrix viridana* (in %) on S- and T-oaks.

	L1	L2	L3	L4	L5	Pupa	Total
S	26.1	16.6	3.2	3.2	2.5	10.8	62.4
T	24.7	29.3	2.7	1.3	0.7	14.7	73.3

For *L. dispar* the overall mortality rate was rather low. During the rearing of the larvae for the later experiment in the first three larval stages only five to seven individuals per instar died. During the experiment beginning with the fourth instar, five larvae had problems to pupate and died. Of these, three belonged to group S\_C and one each to S-D and T\_C. During the hatching to an adult moth two additional individuals died. One of the S\_C group and one of the T\_D group.

### 3.4. Comparison of the Performance of Both Moths

Both insects showed significant differences in their developmental times, mostly with an increased developmental time when fed with T-oaks. For *T. viridana* and the females of *L. dispar* also significant differences in the leaf area fed between S-oaks and T-oaks were observed but with different directions. While *L. dispar* larvae ate more leaves on S-oaks, larvae of *T. viridana* needed to eat more T-oak leaves. Most striking is the difference between the insects in the mortality rate. Where

more *T. viridana* larvae died during their development than reaching the adult stage, for *L. dispar* a very low overall mortality was witnessed (Table 4).

**Table 4.** Comparison of the performance parameters of both moth.

	Developmental Time		Amount of Fed Leaves	Mortality
<i>Tortrix viridana</i>	L1*, L2*, T > S	L4*** S > T	L5*, total* T > S	total* T > S
<i>Lymantria dispar</i>	L5*, pupa* both T > S		S_C-T_C*, S_C-S_D* T < S	hardly any mortality

#### 4. Discussion

For *T. viridana* the supposed worse physiological quality of leaves from T-oaks had no overall influence on the juvenile developmental duration. Whereas the more detailed experiment with *L. dispar* implicated a negative effect of pre-infestation especially in T-oaks. Thus, here pre-experimental induced defence compounds elongated the developmental time when fed with leaves from T-oaks.

For both insects the amount of the leaf area fed has to be considered together with the total weight of the pupae. *T. viridana* engaged in compensatory feeding on T-oaks to overcome the negative effects of plant defence related reduction in leaf quality resulting in nearly the same pupal weight for larvae reared on S-oaks or T-oaks, respectively [14]. This could not be observed for *L. dispar*. The differences in pupae weight were correlated with differences in food intake for *L. dispar* (data not shown), which instead slowed down in development. However, both insects overall seemed to prefer the leaves of S-oaks. This could be explained by the repellent herbivore induced plant volatiles (HIPVs) produced in T-oaks [14]. Rossiter et al. [25] found constitutive and elevated phenolic and nonphenolic constituents to be partly responsible for reduced performance in *L. dispar*. This supports the hypothesis of weaker development of larvae because of strong defence signalling in pre-infested T-oaks (T\_D) since T-oaks have elevated fractions of the herbivore-repellent herbivore induced plant volatiles (HIPVs) of the sesquiterpenes  $\alpha$ -farnesene and germacrene D [14]. An elevation in these volatile defence associated emissions was demonstrated by Copolovici et al. [26] in feeding experiments with *L. dispar* on *Q. robur* as well. This would mean that unlike the effects from T-oaks on *T. viridana*, the HIPVs rather than the secondary metabolites e.g., tannins would have had the negative effect on *L. dispar* performance. In *T. viridana*, on the other hand, the tannins may be the reason for the high mortality rate because it is proven that phenolic compounds in the guts of insects lead to destruction of essential nutrients and subsequently to damage of the midgut [27].

#### 5. Conclusions

In an earlier study, we hypothesized that T- and S-oaks follow different defence strategies. Where T-oaks prefer a constitutive way of defence shown by high amounts of sesquiterpenes and bioactive tannins, S-oaks react 'just-in-time' to insect's attacks by increasing defensive substances only after insects feeding [14,28]. The defence strategy of T-oaks may make them to a contributor to a stronger resistance of German forests against pests like *T. viridana* and *L. dispar* not at least in regards to climate change.

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## Abbreviations

T-oaks	Oaks, classified as tolerant; less infested during outbreak of herbivorous insects
S-Oaks	Oaks. Classified as susceptible, heavily infested during outbreak of herbivorous insects
VOCs	volatile organic compounds
HIPVs	herbivore induced plant volatiles
T_C	T-oaks control
S_C	S-oaks control
T_D	T-oaks damaged
S_D	S-oaks damaged
L (combined with a number, for example 3)	larval stage 3 or third instar
C	control-tree
D	damaged tree

## References

- Riedel, T.; Stürmer, W.; Hennig, P.; Dunger, K.; Bolte, A. Wälder in Deutschland sind eine wichtige Kohlenstoffsenke. *AFZ-Der Wald* **2019**, *14*, 14–18, Available online: [www.bundeswaldinventur.de/fileadmin/SITE\\_MASTER/content/Downloads/CI2017/AFZ\\_14\\_19\\_Kohlenstoff\\_Artikel\\_2\\_Riedel.pdf](http://www.bundeswaldinventur.de/fileadmin/SITE_MASTER/content/Downloads/CI2017/AFZ_14_19_Kohlenstoff_Artikel_2_Riedel.pdf) (accessed on 5 November 2020).
- Bartsch, N.; Röhrig, E. Prozesse der Walddynamik. In *Waldökologie*; Springer Spektrum: Berlin/Heidelberg, Germany, 2016; doi:10.1007/978-3-662-44268-5\_15.
- Ellenberg, H.; Dierschke, H. Vegetation Mitteleuropas mit den Alpen. In *Ökologischer, Dynamischer und Historischer Sicht; 203 Tabellen with Assistance of Christoph Leuschner*, 6th ed.; Verlag Eugen Ulmer: Stuttgart, Germany, 2010.
- Parry, M.L.; Canziani, O.; Palutikof, J.P.; van der Linden, P.; Hanson, C.E. *Climate Change 2007: Impacts, Adaptation and Vulnerability*; Contribution of Working Group II to the Fourth Assessment Report of the Intergovernmental Panel on Climate Change; 2007.
- Delb, H. *Modul Wald Teilbericht Waldschutz und Klimawandel*; Forstliche Versuchs- und Forschungsanstalt Baden-Württemberg: Abteilung Waldschutz, 2013. Available online: [www.kwis-rlp.de](http://www.kwis-rlp.de).
- BMEL Bundeswaldinventur. *Der Wald in Deutschland Ausgewählte Ergebnisse der dritten Bundeswaldinventur*; Bundesministerium für Ernährung und Landwirtschaft: Berlin, Germany, 2014.
- Ehrlich, P.R.; Raven, P.H. Butterflies and plants: A study in co-evolution. *Evolution* **1964**, *18*, 586–608, doi:10.2307/2406212.
- Becerra, J.X. Synchronous coadaptation in an ancient case of herbivory. *Proc. Natl. Acad. Sci USA* **2003**, *100*, 12804–12807, doi:10.1073/pnas.2133013100.
- Dengler, A.; Röhrig, E. *Waldbau. Der Wald als Vegetationstyp und seine Bedeutung für den Menschen*. Verlag Paul Parey: Berlin, Germany, 1980; 283p.
- Hunter, M.D. Differential susceptibility to variable plant phenology and its role in competition between two insect herbivores on oak. *Ecol Entomol* **1990**, *15*, 401–408, doi:10.1111/j.1365-2311.1990.tb00823.x.
- Hartmann, G.; Blank, R. Winterfrost, Kahlfraß und Prachtkäferbefall als Faktoren im Ursachenkomplex des Eichensterbens in Norddeutschland. *Forst und Holz* **1992**, *15*, 443–452.
- Führer, E. *Oak Decline in Central Europe: A Synopsis of Hypotheses Proceedings: Population Dynamics, Impacts and Integrated Management of Forest Defoliating Insects*; McManus, M.L., Liebhold, A.M., Eds., USDA Forest Service Technical Report NE-247; 1998; pp. 7–24.
- Schröder, H.; Ziegler, C. Die Situation der Eiche in NRW im Frühjahr 2005. *AFZ Der Wald* **2006**, *6*, 320–321.
- Ghirardo, A.; Heller, W.; Fladung, M.; Schnitzler, J.-P.; Schroeder, H. Function of defensive volatiles in pedunculate oak (*Quercus robur*) is tricked by the moth *Tortrix viridana*. *Plant Cell Environ.* **2012**, *35*, 2192–2207, doi:10.1111/j.1365-3040.2012.02545.x.
- Dicke, M.; Baldwin, I.T. The evolutionary context for herbivore-induced plant volatiles: Beyond the ‘cry for help’. *Trends Plant Sci.* **2010**, *15*, 167–175, doi:10.1016/j.tplants.2009.12.002.
- Schroeder, H.; Orgel, F.; Fladung, M. Performance of the green oak leaf roller (*Tortrix viridana* L.) on leaves from resistant and susceptible oak genotypes. In *Mitteilungen der Deutschen Gesellschaft für Allgemeine und Angewandte Entomologie*; 2016; pp. 265–269.

17. Schröder, H. Sommerveredelung bei Eichen-eine Erfolgsgeschichte, *AFZ Der Wald* **2010**, *5*, 16–17.
18. Magnoler, A. A wheat germ medium for rearing of the gypsy moth *Lymantria dispar* L. *Entomophaga* **1970**, *15*, 401–406, doi:10.1007/BF02370308.
19. R Core Team R: A language and Environment for Statistical. Version 3.6.1: R Foundation for Statistical Computing, Vienna, Austria, 2017. Available online: <https://www.R-project.org/>.
20. RStudio Team Integrated Development Environment for R; Version 1.2.5001; RStudio, Inc.: Boston, MA, USA, 2019; Available online: <http://www.rstudio.com/>.
21. Hijmans, R.J. Raster: Geographic Data Analysis and Modeling. R package version 3.0-7. 2019. Available online: <https://CRAN.R-project.org/package=raster>.
22. Pohlert, T. PMCMRplus: Calculate Pairwise Multiple Comparisons of Mean Rank Sums. Version 1.4.2. 2019. Available online: <https://CRAN.R-project.org/package=PMCMRplus>.
23. Aho, K. Asbio: A Collection of Statistical Tools for Biologists. Version 1.5-5. 2019. Available online: <https://CRAN.R-project.org/package=asbio>.
24. Wickham, H. *ggplot2: Elegant Graphics for Data Analysis*; Springer: New York, NY, USA, 2016. Available online: <https://ggplot2.tidyverse.org>.
25. Rossiter, M.C.; Schultz, J.C.; Baldwin, I.T. Relationships among Defoliation, Red Oak Phenolics, and Gypsy Moth Growth and Reproduction. *Ecology* **1988**, *69*, 267–277, doi:10.2307/1943182.
26. Copolovici, L.; Pag, A.; Kännaste, A.; Bodescu, A.; Tomescu, D.; Copolovici, D. Disproportionate photosynthetic decline and inverse relationship between constitutive and induced volatile emissions upon feeding of *Quercus robur* leaves by large larvae of gypsy moth (*Lymantria dispar*). *Environ. Exp. Bot.* **2017**, *138*, 184–192, doi:10.1016/j.envexpbot.2017.03.014.
27. Barbehenn, R.; Cheek, S.; Gasperut, A.; Lister, E.; Maben, R. Phenolic compounds in red oak and sugar maple leaves have prooxidant activities in the midgut fluids of *Malacosoma disstria* and *Orgyia leucostigma* caterpillars. *J. Chem. Ecol.* **2005**, *31*, 969–988, doi:10.1007/s10886-005-4242-4.
28. Kersten, B.; Ghirardo, A.; Schnitzler, J.P.; Kanawati, B.; Schmitt-Kopplin, P.; Fladung, M.; Schroeder, H.. Integrated transcriptomics and metabolomics decipher differences in the resistance of pedunculate oak to the herbivore *Tortrix viridana* L. *BMC Genomics* **2013**, *14*, 737, Available online: <http://www.biomedcentral.com/1471-2164/14/737>.

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