

Conference Proceedings Paper

# The Essential Oils of Thyme, Sage and Peppermint against Strawberry Anthracnose

Amina Morkeliūnė \*, Neringa Rasiukevičiūtė and Alma Valiuškaitė

Laboratory of Plant Protection, Institute of Horticulture, Lithuanian Research Centre for Agriculture and Forestry, LT-54333 Babtai, Kaunas District, Lithuania; neringa.rasiukeviciute@lammc.lt (N.R.); alma.valiuskaite@lammc.lt (A.V.)

\* Correspondence: armina.morkeliune@lammc.lt

Published: 30 November 2020

**Abstract:** Strawberry *Colletotrichum* spp. is a significant strawberry pathogen with yield losses of up to 50 %, due to agrometeorological conditions change its spread in a temperate climate is growing. The most convenient way for controlling diseases is the use of chemical fungicides. The demand for alternative measures for biological protection is growing because of the increasing pathogens resistance and pesticides harm for human and environment. Thyme, sage and peppermint are a source of natural antioxidants and biologically active compounds. The findings of antimicrobial activities of these plants, low toxicity and biodegradability of essential oils (EO) make them suitable for biological protection against pathogens. This study aims to evaluate the inhibition of *Colletotrichum* spp. by thyme, sage and peppermint EO in vitro and on detached strawberry leaves. The results revealed that thyme EO inhibited *Colletotrichum* spp. completely above 200  $\mu\text{L L}^{-1}$  concentration in vitro. Peppermint and sage EO reduced mycelial growth of *Colletotrichum* spp. However, in vitro, results are auspicious for biological control. The experiments on detached strawberry leaves showed that *Colletotrichum* spp. disease reduction 4 days after inoculation were 15.8% at 1000  $\mu\text{L L}^{-1}$  of peppermint EO and 5.3% at 800  $\mu\text{L L}^{-1}$  of thyme compared with control. Our findings could potentially help to manage *Colletotrichum* spp.; however, the detached strawberry leaves assay showed that EOs efficacy was relatively low on tested concentrations and need further investigations with higher concentrations.

**Keywords:** biocontrol; inhibition; *Mentha piperita*; *Salvia officinalis*; *Thymus vulgaris*

## 1. Introduction

The strawberry anthracnose is one of the important diseases, which caused by several species complex of *Colletotrichum* spp.: *C. acutatum* J. H. Simmonds, Brooks and *C. gloeosporioides* (Penz.) Penz. & Sacc. [1,2]. It also infects and causes diseases in many economically important crops. Strawberry anthracnose caused yield losses of up to 50% and 80% plant death and was considered to be warmer climate zone pathogen, where the optimal temperature from 15 to 30 °C, with optimal 25 °C temperature [3–5]. Strawberry diseases controlled by several fungicide applications [6,7]. The growing resistance to pesticides and their adverse environmental effects leads to a new environment safer disease control strategy [8–10].

Essential oils (EO) demonstrated a distinct level of antimicrobial activity to various ranges of strawberry fungal and bacterial pathogens [6,9,11,12]. The EO includes terpenes, terpenoids, aromatic and aliphatic constituents are a source of natural antioxidants and biologically active compounds. The EO mostly are secondary metabolites, which play an essential role in plant defence as they often possess antimicrobial properties and are non-toxic and biodegradable [13–17].

Plant EO are developed commercially on a large scale, most of which are the members of the *Lamiaceae* family, including *T. vulgaris*, *S. officinalis*, *M. piperita* [18,19]. Thyme (*Thymus vulgaris* L.), sage (*Salvia officinalis* L.) and peppermint (*Mentha piperita* L.) EO, as products from plants, have a wide application in pharmacy, fragrance, food industries, however recent studies of essential oils

revealed their potential antimicrobial activity [20–22]. *Salvia officinalis* EO affects *Fusarium* spp. growth [15]. *Mentha piperita* EO inhibits spread of *Alternaria* spp. and *Fusarium* spp. pathogens [23]. *Thymus vulgaris* EO has antifungal activity against plant pathogens such as *M. fructicola*, *B. cinerea*, *A. flavus* [16,24,25]. *T. vulgaris*, EO can be used as a natural preservative in food against casual agents of food-borne diseases like *E. coli*, *Pseudomonas* spp. and others [26].

Plant protection measures against plant diseases are necessary to avoid yield and crop losses. However, pesticides have an adverse effect on plants and humans, as they leave residues in it. The European Green Deal provides a plan to increase environmentally friendly technologies by supporting strategy to reduce the usage of pesticides and make agriculture more sustainable. Growing pathogens resistance is because of the extensive use of chemical pesticides for plant protection. The new sources of natural active ingredients for plant protection may solve pesticides resistance problem and reduce environment and food contamination [7,11,12,27].

This study aims to evaluate the inhibition of *Colletotrichum* spp. by thyme, sage and peppermint EO in vitro and on detached strawberry leaves.

## 2. Experiments

### 2.1. Essential Oil Extraction.

The common thyme (*Thymus vulgaris* L.), common sage (*Salvia officinalis* L.), peppermint (*Mentha piperita* L.) essential oils (EO) extracted from dry herbs by Clevenger-type hydro-distillation. Volatile compounds of essential oils were determined by gas chromatography/mass spectrometry (GC-MS).

### 2.2. Antifungal Activity In Vitro

The research was carried out at the Laboratory of Plant Protection, Lithuanian Research Centre for Agriculture and Forestry Institute of Horticulture (LAMMC) in 2017–2020. To evaluate antifungal activity against *Colletotrichum* spp. 50–1800  $\mu\text{L L}^{-1}$  concentrations of EO were used. Isolates were obtained from infected strawberry fruits and had previously been identified as *Colletotrichum* spp. The 5 mm mycelial plugs of 7-day old fungus cut and placed in the centre of Petri plates containing PDA. Different concentrations of pure essential oil added to cooled at 45 °C PDA.

The mycelium put upside down (mycelia side) in the centre of Petri. The Petri plates incubated at  $25 \pm 2$  °C in the dark. The control treatments were essential oil-free but inoculated with pathogen. The diameter (mm) of *Colletotrichum* spp. colony growth in two directions measured 4 days after inoculation (DAI). Four replications were carried out.

The mean of colony growth diameter used for mycelial growth inhibition (MGI) calculations. The MGI (%) was determined using the formula:

$$\text{MGI (\%)} = (C - T)/C \times 100,$$

where C is mycelium diameter of the pathogen colony in control Petri dish, mm; T—mycelium diameter of the pathogen colony in the essential oil-treated Petri dish, mm [11,28,29].

### 2.3. EO antifungal Activity on Detached Strawberry Leaves

The essential oils inhibitory effect evaluated on detached strawberry cultivar 'Deluxe' leaves. Healthy strawberry leaves without any visible disease symptoms were soaked in 70% ethanol solution for 3 min and rinsed 4–5 times with sterile distilled water (SDW). Each leaf placed in a Petri dish with 5 mL of SDW. Detached strawberry leaves were sprayed with essential oils, then wounded with a sterile needle and a 9-mm plug of 7-day-old *Colletotrichum* spp. was placed on the wound. Incubation was carried out at  $25 \pm 2$  °C in the dark for 7 days. The disease severity and reduction evaluated 4 DAI. The control treatments were not sprayed with EO but inoculated with pathogen. Four replications were carried out.

Disease severity (DS) of each inoculated plant leaf assessed at 4 DAI by calculating the percentage of leaf area affected: (1) 0%—no visible infection, (2) 5%, (3) 10%, (4) 20% and (5) 50% or more area of leaf infected [30,31].

$$DS(\%) = \frac{(0 \times P_0) + (1 \times P_1) + (2 \times P_2) + (3 \times P_3) + (4 \times P_4) + (5 \times P_5)}{N \times G} \times 100$$

where P0 to P5 is the total number of observed leaves in each corresponding scale, N—total number of leaves, G—number of maximum grades observed in scale [32].

$$DR(\%) = \frac{X_c - X_t}{X_c} \times 100,$$

where X<sub>t</sub> is the mean of DS per treatment, and X<sub>c</sub> is the mean of DS in the untreated inoculated control [33].

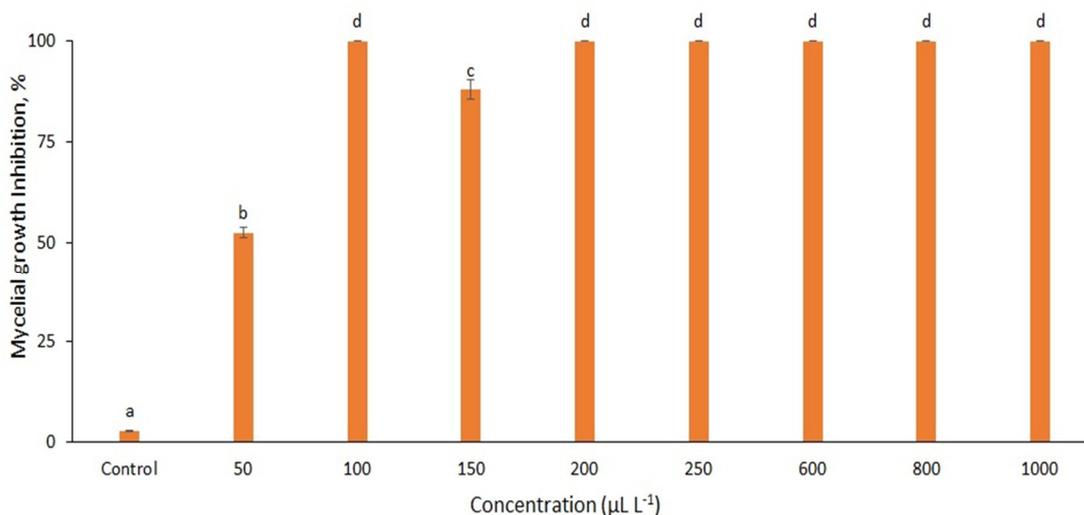
#### 2.4. Statistical Analysis

SAS Enterprise Guide 7.1 program (SAS Institute Inc., Cary, NC, USA) was applied for the analysis of experimental data. Analysis of variance (ANOVA) procedure was processed, and Duncan's multiple range test ( $p < 0.05$ ) was used for the comparison of obtained means. The standard error (SE) in the figures marked as an error bar estimated for growth rates of isolates.

### 3. Results

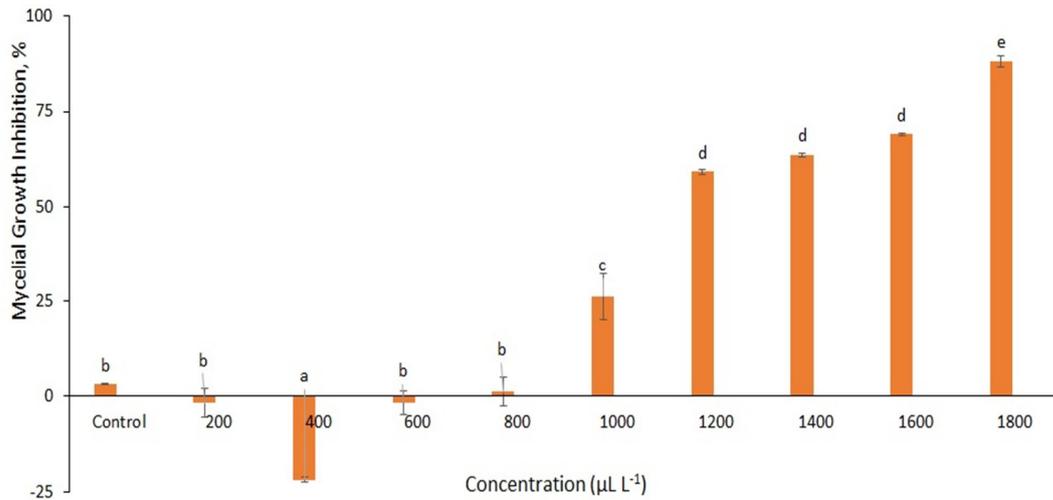
#### 3.1. Antifungal Activity In Vitro

The antifungal activity of EO was investigated on PDA (potato dextrose agar) under different concentrations. The inhibition of *Colletotrichum* spp. by thyme EO is presented in Figure 1. Thyme EO showed 100% MGI at 100  $\mu\text{L L}^{-1}$  at 4 days after inoculation (4 DAI) but did not demonstrate highest antifungal effect at 150  $\mu\text{L L}^{-1}$  at 4 (87.96%) DAI. However, mycelial pathogen growth was inhibited above 200  $\mu\text{L L}^{-1}$ .



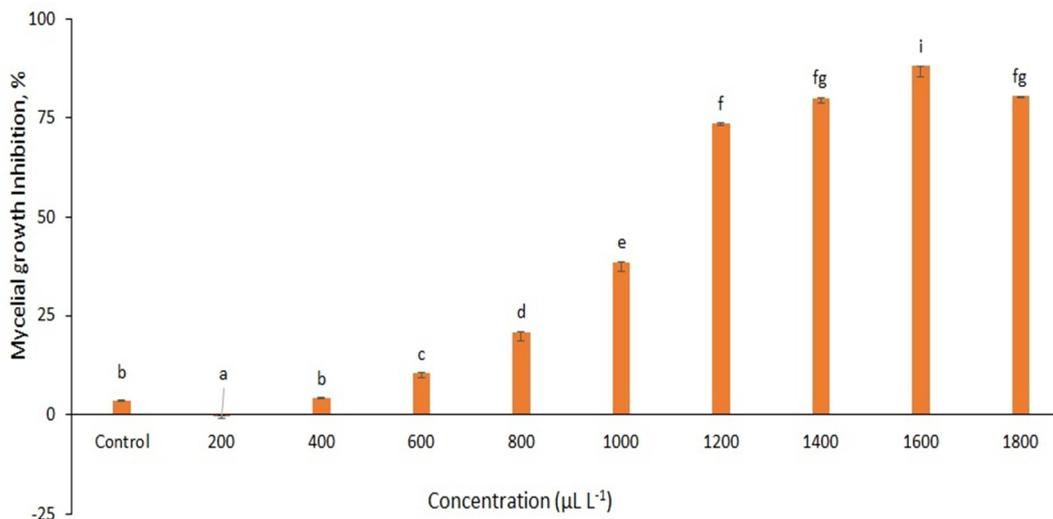
**Figure 1.** Inhibition (%) of *Colletotrichum* spp. mycelial growth by thyme (*T. vulgaris*) EO at 4 days after inoculation (4 DAI). Results are presented as means ( $n = 4$ ). The same letter indicates no significant differences between treatments ( $p < 0.05$ ).

The inhibition of *Colletotrichum* spp. by sage EO is presented in Figure 2. Data indicate that this EO was not as effective as thyme. Sage EO showed antifungal activity up to 1000  $\mu\text{L L}^{-1}$  at 4 DAI and achieved the highest effect of 88.14% at 1800  $\mu\text{L L}^{-1}$ .



**Figure 2.** Inhibition (%) of *Colletotrichum* spp. mycelial growth by sage (*S. officinalis*) EO at 4 days after inoculation (4 DAI). Results are presented as means (n = 4). The same letter indicates no significant differences between treatments ( $p < 0.05$ ).

The fungicidal activity of peppermint EO against *Colletotrichum* spp. is shown in Figure 3. This EO had a similar effect on *Colletotrichum* spp. comparing with sage. Meanwhile, peppermint EO reduced the mycelial growth at 600–1800 µL L<sup>-1</sup> from 20% to 88%. However, the highest antifungal activity was reached at 1600 µL L<sup>-1</sup> at 4 DAI. 1800 µL L<sup>-1</sup> EO efficiency decreased to 62.54%.



**Figure 3.** Inhibition (%) of *Colletotrichum* spp. mycelial growth by peppermint (*M. piperita*) EO at 4 days after inoculation (4 DAI). Results are presented as means (n = 4). The same letter indicates no significant differences between treatments ( $p < 0.05$ ).

*T. vulgaris* EO suppressed the mycelial growth at 200–1000 µL L<sup>-1</sup>. The determined MIC (minimal inhibitory concentration) was equal to 200 µL L<sup>-1</sup>. *S. officinalis* and *M. piperita* EO reduced mycelial growth compared to control. However, the antifungal effect was insufficient to inhibit the spread of anthracnose infection.

### 3.2. Antifungal Activity on Detached Strawberry Leaves

The detached strawberry leaf assay developed to determine the efficiency of the essential oil against *Colletotrichum* spp. on detached strawberry leaves. The results revealed that among all the investigation treatments, only 1000  $\mu\text{L L}^{-1}$  concentration of peppermint EO (5.3%) and 800  $\mu\text{L L}^{-1}$  concentration of thyme (15.8%) reduced the infection on strawberry leaves compared to inoculated control 4 DAI (Table 1). Sage EO had no positive influence on infection spread.

**Table 1.** The disease severity and reduction of anthracnose regarding strawberry cultivar 'Deluxe' by different essential oils concentrations at 4 days after inoculation. Means n = 18.

Treatments	Disease Severity (%)	Disease Reduction (%)
Inoculated control	79.2 $\pm$ 0.2	n.a. *
<i>Thymus vulgaris</i> 800 $\mu\text{L/L}$	75 $\pm$ 0.3	5.3
<i>Salvia officinalis</i> 1000 $\mu\text{L/L}$	80.6 $\pm$ 0.2	0
<i>Mentha piperita</i> 1000 $\mu\text{L/L}$	66.7 $\pm$ 0.2	15.8

#### 4. Discussion

The growing interest in essential oils and their components, due to their volatility, relatively safe condition and wide acceptance by consumers, ecologically and biodegradable properties. For our study, we selected *T. vulgaris*, *S. officinalis* and *M. piperita* EOs. We analyzed antifungal activities also their effect on detached strawberry leaves, to assess the feasibility of using EOs as biocontrol agents in disease control measure. EO from thyme, sage, peppermint presented noticeable antifungal activity against *Colletotrichum* spp. *in vitro*.

In an investigation conducted by Duduk et al. [34], thyme EO showed good antifungal efficacy against *C. acutatum* on strawberry fruit. In our study, thyme EO inhibited *Colletotrichum* spp. mycelial growth *in vitro* above 200  $\mu\text{L L}^{-1}$ . This suggests that the antifungal effect presence of the dominant components of EO, as main activity carriers. Palfi et al. [17] reported that thyme, sage and peppermint EOs completely inhibited the mycelial growth of *F. oxysporum* *in vitro*. However, sage EO had a low inhibitory effect against *B. cinerea*. Oliveira et al. [35] reported that 5  $\mu\text{L/mL}$  peppermint EO resulted in 100% MGI on all tested *Colletotrichum* stains. In our research peppermint, EO highest antifungal activity reached 1600  $\mu\text{L L}^{-1}$  (88%).

In our research, the antifungal activity of sage EO against *Colletotrichum* spp. achieved the highest effect of 88.14% at 1800  $\mu\text{L L}^{-1}$ . In comparison, Yilmaz et al. [36] studied, that the application of sage EO resulted in slight inhibition on mycelial growth of *C. gloeosporioides* in fumigation bioassay and contact bioassay *in vitro* (solid media) and *in vivo* (apple) conditions. These results supported our findings, shows antifungal effect against *Colletotrichum* spp., but did not suppress totally.

However, to the best of our knowledge, no investigations have been previously performed on the antifungal effect of thyme, sage and peppermint EOs on detached strawberry leaves against *Colletotrichum* spp. In the present research, investigated EO result on detached strawberry leaves assay shows a less positive effect of reducing the spread of anthracnose infection. 1000  $\mu\text{L L}^{-1}$  concentration of peppermint EO (15.8%) and 800  $\mu\text{L L}^{-1}$  concentration of thyme (5.3%) reduced the infection on strawberry leaves. In the future research, a higher concentration of this EO should be investigated for higher effectiveness. The effect of EOs from plants as volatile compounds on the surface of strawberry leaves may induce a stressful environment [11,33].

In summary, examination with various EO and their concentrations *in vitro* exhibited promising prospects against strawberry anthracnose. However, the antifungal effect on detached strawberry leaves was relatively low.

#### 5. Conclusions

Essential oil thyme showed total inhibition against *Colletotrichum* spp. *in vitro*. Peppermint and sage EOs showed significant antifungal activity at the highest concentrations. Infection of anthracnose on detached strawberry leaves was slightly reduced by the application of thyme EO and

more suppressed by peppermint EO at tested concentration. Sage EO did not influence the spread of *Colletotrichum* spp. on detached strawberry leaves. The detached strawberry leaves assay showed that the investigated essential oils were not equally effective and need further investigations with higher concentrations.

**Author Contributions:** Conceptualization: N.R.; methodology: A.M., N.R. and A.V.; software: A.M.; validation: N.R.; formal analysis: A.M.; investigation: A.M. and N.R.; resources: N.R.; data curation: A.M.; writing—original draft preparation: A.M.; writing—review and editing: A.M., N.R. and A.V.; visualization: A.M. and N.R.; supervision: A.V. All authors have read and agreed to the published version of the manuscript.

**Funding:** No funds received.

**Conflicts of Interest:** The authors declare no conflict of interest.

## Abbreviations

The following abbreviations are used in this manuscript:

EO	essential oil
DAI	days after inoculation
LAMMC	Lithuanian Research Centre for Agriculture and Forestry
PDA	potato dextrose agar
MGI	mycelial growth inhibition
DS	diseases severity
DR	disease reduction
SDW	sterile distilled water
GC-MS	gas chromatography/mass spectrometry

## References

1. Cannon, P.F.; Damm, U.; Johnston, P.R.; Weir, B.S. *Colletotrichum*—current status and future directions. *Stud. Mycol.* **2012**, *73*, 181–213.
2. Udayanga, D.; Manamgoda, D.S.; Liu, X.; Chukeatirote, E.; Hyde, K.D. What are the common anthracnose pathogens of tropical fruits? *Fungal Divers.* **2013**, *61*, 165–179.
3. Jayawardena, R.S.; Huang, J.K.; Jin, B.C.; Yan, J.Y.; Li, X.H.; Hyde, K.D.; Bahkali, A.H.; Yin, S.L.; Zhang, G.Z. An account of *Colletotrichum* species associated with strawberry anthracnose in china based on morphology and molecular data. *Mycosphere* **2016**, *7*, 1147–1163.
4. He, L.; Li, X.; Gao, Y.; Li, B.; Mu, W.; Liu, F. Characterisation and fungicide sensitivity of *Colletotrichum* spp. from different hosts in Shandong, China. *Plant Dis.* **2019**, *103*, 34–43.
5. Forcelini, B.B.; Seijo, T.E.; Amiri, A.; Peres, N.A. Resistance in strawberry isolates of *Colletotrichum acutatum* from Florida to quinone-outside inhibitor fungicides. *Plant Dis.* **2016**, *100*, 2050–2056.
6. Hong, J.K.; Yang, H.I.; Jung, H.; Yoon, D.J.; Sang, M.K.; Jeun, Y.C. Application of volatile antifungal plant essential oils for controlling pepper fruit anthracnose by *Colletotrichum gloeosporioides*. *Plant Pathol. J.* **2015**, *31*, 269–277.
7. Rasiukevičiūtė, N.; Uselis, N.; Valiuškaitė, A. The use of forecasting model IMETOS® for strawberry grey mould management. *Zemdirb. Agric.* **2019**, *106*, 143–150.
8. Zhang, L.; Song, L.; Xu, X.; Zou, X.; Duan, K.; Gao, Q. Characterisation and fungicide sensitivity of *Colletotrichum* species causing strawberry anthracnose in Eastern China. *Plant Dis.* **2020**, doi:10.1094/PDIS-10-19-2241-RE.
9. Aguado, A.; Pastrana, A.M.; Santos, B.; Romero, F.; Sánchez, M.C.; Capote, N. The efficiency of natural products for the control of *Colletotrichum acutatum* monitored by real-time PCR. *Acta Hort.* **2014**, *1049*, 329–334.
10. Mahilrajan, S.; Nandakumar, J.; Kailayalingam, R.; Manoharan, N.A.; Srivijeindran, S. Screening the antifungal activity of essential oils against decay fungi from palmyrah leaf handicrafts. *Biol. Res.* **2014**, *47*, 35.
11. Šernaitė, L.; Rasiukevičiūtė, N.; Valiuškaitė, A. The extracts of cinnamon and clove as potential biofungicides against strawberry grey mould. *Plants* **2020**, *9*, 613.

12. Šernaitė, L.; Rasiukevičiūtė, N.; Valiuškaitė, A. Application of plant extracts to control postharvest gray mold and susceptibility of apple fruits to *B. cinerea* from different plant hosts. *Foods* **2020**, *9*, 1430.
13. Lopez, N.L.; Gutiérrez-Grijalva, E.P.; Vazquez-Oliva, G.; Heredia, J.B. Essential oils of Oregano: Biological activity beyond their antimicrobial properties. *Molecules* **2017**, *22*, 989.
14. Naeini, A.; Ziglari, T.; Shokri, H.; Khosravi, A.R. Assessment of growth inhibiting effect of some plant essential oils on different *Fusarium* isolates. *J. Mycol. Med.* **2010**, *20*, 174–178.
15. Foltinová, D.; Tančinová, D.; Císarová, M. Influence of essential oils on the growth of *Aspergillus flavus*. *Potravinárstvo* **2017**, *11*, 322–331.
16. Oliveira, R.C.; Moreno, M.C.; Correa, B.; Callejas, F.J. Cellular, physiological and molecular approaches to investigate the antifungal and anti-aflatoxigenic effects of thyme essential oil on *Aspergillus flavus*. *Food Chem.* **2020**, *315*, 126096.
17. Palfi, M.; Konjevoda, P.; Vrandečić, K. Antifungal activity of essential oils on mycelial growth of *Fusarium oxysporum* and *Botrytis cinerea*. *Emir. J. Food Agric.* **2019**, *31*, 544–554.
18. Sil, A.; Pramanik, K.; Samantaray, P.; Mondal, M.F.; Yadav, V. Essential oils: A boon towards eco-friendly management of phytopathogenic fungi. *J. Entomol. Zool. Stud.* **2020**, *8*, 1884–1891.
19. Moumni, S.; Elaissi, A.; Trabels, A.; Merghni, A.; Chraief, I.; Jelassi, B.; Chemli, R.; Ferchichi, S. Correlation between chemical composition and antibacterial activity of some *Lamiaceae* species essential oils from Tunisia. *BMC Complement Med. Ther.* **2020**, *20*, 103.
20. Kumar, A.; Kudachikar, V.B. Antifungal properties of essential oils against anthracnose disease: A critical appraisal. *J. Plant Dis. Prot.* **2018**, *125*, 133–144.
21. Starović, M.; Ristic, D.; Pavlović, S.; Ristic, M.; Stevanović, M.; Aljuhaimi, F.; Svetlana, N.; Özcan, M.M. Antifungal activities of different essential oils against anise seeds mycopopulations. *J. Food Saf. Food Qual.* **2016**, *67*, 72–78.
22. Micucci, M.; Protti, M.; Aldini, R.; Frosini, M.; Corazza, I.; Marzetti, C.; Mattioli, L.B.; Tocci, G.; Chiarini, A.; Mercolini, L.; et al. *Thymus vulgaris* L. Essential Oil Solid Formulation: Chemical Profile and Spasmolytic and Antimicrobial Effects. *Biomolecules* **2020**, *10*, 860.
23. Desam, N.R.; Al-Rajab, A.J.; Sharma, M.; Mylabathula, M.M.; Gowkanapalli, R.R.; Albratty, M. Chemical constituents, in vitro antibacterial and antifungal activity of *Mentha × piperita* L. (peppermint) essential oils. *J. King Saud. Univ. Sci.* **2017**, *31*, 528–533.
24. Santoro, K.; Maghenzani, M.; Chiabrando, V.; Bosio, P.; Gullino, M.L.; Spadaro, D.; Giacalone, G. Thyme and savory essential oil vapor treatments control brown rot and improve the storage quality of peaches and nectarines, but could favor gray mold. *Foods* **2018**, *7*, 7.
25. Taghavi, T.; Kim, C.; Rahemi, A. Role of Natural Volatiles and Essential Oils in Extending Shelf Life and Controlling Postharvest Microorganisms of Small Fruits. *Microorganisms* **2018**, *6*, 104.
26. Çetin, B.; Çakmakçi, S.; Çakmakçi, R. The investigation of antimicrobial activity of thyme and oregano essential oils. *Turk. J. Agric. For.* **2011**, *35*, 145–154.
27. Park, J.Y.; Kim, S.H.; Kim, N.H.; Lee, S.W.; Jeun, Y.C.; Hong, J.K. Differential Inhibitory Activities of Four Plant Essential Oils on In Vitro Growth of *Fusarium oxysporum* f. sp. *fragariae* Causing Fusarium Wilt in Strawberry Plants. *Plant Pathol. J.* **2017**, *33*, 582–588.
28. Idris, F.M.; Ibrahim, A.M.; Forsido, S.V. Essential oils to control *Colletotrichum musae* in vitro and in vivo on banana fruits. *AEJAES* **2015**, *15*, 291–302.
29. Šernaitė, L.; Rasiukevičiūtė, N.; Dambrauskienė, E.; Viškelis, P.; Valiuškaitė, A. Efficacy of plant extracts and essential oils for biocontrol of strawberry pathogen *Botrytis cinerea*. *Zemdirb. Agric.* **2020**, *107*, 147–152.
30. EPPO. Efficacy evaluation of fungicides: Leafspots of vegetables. *EPPO PP 1/121* **1996**, *2*, 134–139.
31. Bajpai, S.; Shukla, P.S.; Asiedu, S.; Pruski, K.; Prithiviraj, B. A biostimulant preparation of brown seaweed *Ascophyllum nodosum* suppresses powdery mildew of strawberry. *Plant Pathol. J.* **2019**, *35*, 406–416.
32. Kone, N.; Asare-Bediako, E.; Silue, S.; Kone, D.; Koita, O.; Menzel, W.; Winter, S. Influence of planting date on incidence and severity of viral disease on cucurbits under field condition. *Ann. Agric. Sci.* **2017**, *62*, 99–104.
33. Hosseini, S.; Amini, J.; Saba, M.K.; Karimi, K.; Pertot, I. Preharvest and postharvest application of garlic and rosemary essential oils for controlling anthracnose and quality assessment of strawberry fruit during cold storage. *Front. Microbiol.* **2020**, *11*, 1855.

34. Duduk, N.; Markovic, T.; Vasic, M.; Duduk, B.; Vico, I.; Obradovic, A. Antifungal activity of three essential oils against *Colletotrichum acutatum*, the casual agent of strawberry anthracnose. *Essent. Oil-Bear. Plants*. **2015**, *18*, 529–537.
35. Oliveira, K.Á.R.; Berger, L.R.R.; Araújo, S.A.; Câmara, M.P.S.; Souza, E.L. Synergistic mixtures of chitosan and *Mentha piperita* L. essential oil to inhibit *Colletotrichum* species and anthracnose development in mango cultivar Tommy Atkins. *Food Microbiol.* **2017**, *66*, 96–103.
36. Yilmaz, A.; Ermis, E.; Boyraz, N. Investigation of in vitro and in vivo antifungal activities of different plant essential oils against postharvest apple rot diseases—*Colletotrichum gloeosporioides*, *Botrytis cinerea* and *Penicillium expansum*. *J. Food Saf. Food Qual.* **2016**, *67*, 113–148.

**Publisher's Note:** MDPI stays neutral with regard to jurisdictional claims in published maps and institutional affiliations.



© 2020 by the authors. Licensee MDPI, Basel, Switzerland. This article is an open access article distributed under the terms and conditions of the Creative Commons Attribution (CC BY) license (<http://creativecommons.org/licenses/by/4.0/>).