

1 Proceedings

2 **Curcumin-in-cyclodextrins-in-liposomes: an alternative** 3 **for osteoarthritis treatment**

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17 **Abstract:** Osteoarthritis (OA) is one of the most frequent degenerative joint disease characterized by joint
18 pain and stiffness traditionally treated with symptomatic drugs like oral NSAIDs and, in extreme cases,
19 with intra-articular corticoids. However, both these drugs are not exempt from adverse effects. Curcumin
20 (Cur) has proven its anti-inflammatory properties and its potential as anti-osteoarthritic drug. However, its
21 low solubility hinders its usage and limits its therapeutic efficacy. To overcome this issue, drug-in-
22 cyclodextrin double-loaded liposomes (DCL-DL) were developed. These liposomes contained free drug in
23 the lipid bilayer and drug-cyclodextrin complex in the aqueous compartment. The aim of this work was to
24 evaluate the actual effectiveness of Cur-DCL-DL formulations in the OA treatment by intra-articular
25 treatment. For this purpose, the monoiodoacetate (MIA) model of OA pain in rats was used. A single dose
26 of samples containing Cur as DCL-DL, conventional liposomes (SL) and empty liposomes (EL, as control)
27 were injected once intra-articularly. Paw-pressure, beam-balance and incapacitation tests were performed to
28 evaluate OA progression at 7 and 14 days. After ending the assay, animals were sacrificed, and histological
29 evaluation of the ankle-joint tissue was performed. Results showed that DCL-DL significantly reduced pain
30 and ameliorated balance and gait of rats over the 14 days compared to SL. Histological tests showed that
31 DCL-DL had protective properties in some aspects of OA.

32 **Keywords:** drug-in-cyclodextrin-in-liposomes; double-loaded liposomes; Curcumin; Osteoarthritis

33

34 **1. Introduction**

35 Osteoarthritis (OA) is an inflammatory joint disease that affects primarily the elderly. This disease is
36 characterized by progressive degradation of articular cartilage, synovial hyperplasia, osteophyte formation
37 and subchondral bone injury [1].

38 Currently, OA pharmacotherapy is oriented on pain relief and improving function by means of NSAIDs,
39 COX-inhibitors, weak opioids administrated orally, or hyaluronic acid and glucocorticoids administrated via
40 intra-articular injection [2]. However, except for hyaluronic acid, these drugs have numerous adverse effects.
41 To solve this issue, many researchers propose the use of natural products, such as curcumin.

42 Curcumin (Cur) is a polyphenol with numerous properties, like anti-inflammatory action. Regarding
43 OA, Cur is able to inhibit IL-1 β and TNF- α activation of NF- κ B (inactivating multiple pathways of NF- κ B
44 activation) and antagonize COX-2 up regulation by IL-1 β and TNF- α in chondrocytes [3], inhibit



45 inflammatory cell proliferation, decrease the expression of IL-1 β and TNF α in macrophages [4], etc.
46 However, low water solubility of Cur represents a problem for its pharmacological use. To overcome this
47 issue, a double encapsulation in liposomes was proposed in which Cur is entrapped both as free drug in the
48 lipid compartment and as cyclodextrin complex in the aqueous compartment. This approach (Drug-in
49 cyclodextrin-in liposome double-loaded liposomes, DCL-DL) allows improving Cur release and stability [5].

50 The aim of the present work was to test the anti-inflammatory properties of Cur when administered
51 directly to the joint (intra-articular injection) in the form of DCL-DL in comparison with single loaded
52 liposomes (SL). For this purpose, DCL-DL, SL and empty liposomes were formulated and characterized. The
53 formulations were tested in vivo by a rat OA model in terms of paw-pressure, incapacitance and beam-
54 balance test. Histological examination was also performed.

55 2. Experiments

56 2.1. Materials

57 Curcumin (Cur), cholesteryl hemisuccinate (Chems) and 1,2-Dipalmitoyl-sn-glycero-3-phosphocholine
58 (DPPC) were supplied from Sigma-Aldrich Co (Barcelone, Spain). Hydroxypropyl- β -cyclodextrin (HP β CD)
59 was a gift of Roquette (Lestrem, France). Monoiodoacetate (MIA) was provided by Sigma-Aldrich (Milan,
60 Italy). Other chemicals were high-quality analytical products. Solvents were HPLC quality.

61 2.2. Quantification of curcumin

62 Cur concentration was measured by UV/VIS spectrophotometry, as previously reported in Fernández-
63 Romero et al., 2018 [5]. Cur content was measured by using a Shimadzu 1900 UV-visible spectrophotometer
64 (Shimadzu Italia S.r.l., Milan; Italy). 200- μ L of sample was diluted up to 5 mL with a 2% acetonitrile-acetic
65 acid 1:1 v/v mixture, and absorbance was measured at 425 nm. Any interference was observed from other
66 components.

67 2.3. Liposome preparation

68 Liposomes were prepared according to Fernández-Romero et al. (2018) protocol [5]. In brief, 0.0125
69 mmol of CHEMS, 0.064 mmol of DPPC and 0.0027 mmol of Cur were added to a round bottom flask and
70 dissolved in 3.2 mL of methanol and 4.8 mL of chloroform. The mixture was evaporated in a rotary
71 evaporator with a thermostatic bath fixed at 58 °C. The evaporation process was finished when a
72 homogenous film was formed.

73 For DCL-DL formation, 14.5 mM of HP β CD and 0.4 mM of Cur were dissolved in citric acid-disodium
74 phosphate buffer at pH 5.4. After 72 h of constant stirring, the sample was centrifuged at 1000 rpm 10 min to
75 separate the unbounded Cur. Afterwards, 3 mL of this solution was added to the previously formed film and
76 the system was heated at 58 °C in a thermostatic bath for 5 min and stirred with vortex for 1 min. The
77 process was repeated 5 times.

78 As the main objective of this work was to test the ability of Cur to ameliorate OA, two control
79 formulations were prepared: one without Cur (EL), and the other single-loaded liposomes (SL). For SL
80 formation, the procedure was similar to that of DCL-DL, but only citric acid-phosphate buffer was added as
81 aqueous solution (Cur concentration 0.4027 mM). Therefore, these liposomes only contained Cur on to the
82 bilayer. For EL formation, Cur was avoided both in the bilayer and the aqueous space without altering the
83 other components. In all cases, formulations were stored at 4 °C.

84 2.4. Liposome characterization

85 2.4.1. Liposome size, polydispersity index and zeta potential

86 Liposome size and polydispersity index (PDI) were measured by dynamic light scattering technique by
87 using Zetasizer Nano-S equipment (Malvern Instrument, Malvern, UK). Size results were expressed as



88 average liposomal hydrodynamic diameter (nm). PDI values were dimensionless and values below 0.5 were
89 considered indicative of homogenous dispersions.

90 The Z potential or surface charge of vesicles was measured by correlation spectroscopy from
91 electrophoretic mobility (μ), using the same apparatus mentioned before. Results were expressed as zeta
92 potential (Z, mV) after conversion of μ to Z by Smoluchowsky equation: $Z = \mu\eta/\epsilon$, where η represents viscosity
93 and ϵ is the permittivity of the solution. In both cases, samples were prepared equally. 200 μ L of samples
94 were diluted with 3.8 mL of citric acid-phosphate buffer.

95 2.4.2. Encapsulation efficacy

96 To evaluate the amount of Cur entrapped, the encapsulation efficacy (EE) was measured. An aliquot of
97 each sample was centrifuged at 10000 rpm for 1 h at 4 °C. Afterwards, the supernatant was collected and the
98 pellet was treated with sodium lauryl sulfate followed by three cycles of sonication (10 min) and vortex (1
99 min). Subsequently, the suspended pellet was analyzed following section 2.2.

100 2.5. *In vivo* test

101 2.5.1. Animals

102 Male rats (Sprague Dawley, 220-250 g, Envigo, Italy) were housed in cages under controlled conditions
103 (20–24 °C, 50–60% relative humidity, artificial 12-h light–dark cycle). Animals were housed in CeSAL (Centro
104 Stabulazione Animali da Laboratorio, University of Florence) and used at least one week after their arrival.
105 The accommodation was in the Department of Neuroscience, Psychology, Drug Research and Child Health
106 (Florence, Italy), according to European standards as for experimental animals' welfare (European ID-EL 09
107 BIO 03). The rats were kept at 23 ± 1 °C with a 12-h light–dark cycle, light at 7 a.m., and were allowed *ad*
108 *libitum* access to tap water and food. All animal manipulations were carried out according to the Directive
109 2010/63/EU of the European Parliament and of the European Union Council (22 September 2010) on the
110 protection of animals used for scientific purposes. The ethical policy of the University of Florence complies
111 with the guide for the care and use of laboratory animals of the US National Institutes of Health (NIH
112 Publication No. 85–23, revised 1996; University of Florence assurance number: A5278-01). Formal approval
113 to conduct the experiments described was obtained from the animal subjects review board of the University
114 of Florence. Experiments involving animals have been reported according to ARRIVE guidelines [6]. All
115 efforts were made to minimize animal suffering and to reduce the number of animals used.

116 2.5.2. Osteoarthritis animal model

117 Unilateral osteoarthritis was induced by injection of MIA into the tibiotarsal joint [7]. Briefly, rats were
118 lightly anesthetized by 2% isoflurane, the left leg skin was sterilized with 75% ethyl alcohol and the lateral
119 malleolus located by palpation; then, a 28-gauge needle was inserted vertically to penetrate the skin and
120 turned distally for insertion into the articular cavity at the gap between the tibiofibular and tarsal bone until
121 a distinct loss of resistance was felt. 2 mg MIA in 25 μ L saline was delivered into the left articular cavity.
122 Control rats were treated with an equal volume of saline.

123 2.5.3. Treatments

124 Seven days after MIA or vehicle injection, rats were treated, as was explained in section 2.5.2., with
125 40 μ L of solutions (time 0). Groups were as follow:

126 Sham group: 25 μ L of saline vehicle was injected at time -7 and 40 μ L of saline vehicle at time 0.

127 MIA group: 25 μ L of saline vehicle + 2 mg of MIA was injected at time -7 and 40 μ L of saline vehicle at
128 time 0.

129 MIA+ EL group: 25 μ L of saline vehicle + 2 mg of MIA was injected at time -7 and 40 μ L of empty
130 liposomes (EL) at time 0.



131 MIA+ SL group: 25 μ L of saline vehicle containing 2 mg of MIA was injected at time -7 and 40 μ L of
132 single-loaded liposomes (SL) at time 0.

133 MIA+ DCL-DL group: 25 μ L of saline vehicle containing 2 mg of MIA was injected at time -7 and 40 μ L
134 of double loaded liposomes (DCL-DL) at time 0.

135 Paw pressures, beam balance and incapitance tests were performed 7 and 14 days after Cur intra-
136 articular injection. In addition, histological test was performed.

137 2.5.4. Paw pressure test

138 The nociceptive threshold of rats was determined by an analgesimeter (Ugo Basile, Varese, Italy),
139 according to the method described by Leighton et al. [1988]. Briefly, a constantly increasing weight was
140 applied to a small area of the dorsal surface of the hind paw using a blunt conical probe by a mechanical
141 device. Mechanical weight (expressed in g) was increased until vocalization or withdrawal reflex occurred
142 while rats were lightly restrained. An arbitrary cut-off value of 100 g was adopted [7].

143 2.5.5. Beam balance test

144 A rectangular beam (3.2 cm wide, 122 cm long and 63.5 cm tall) was suspended between two tables (105
145 cm tall for the top of the beam). A black box is placed at the end of the beam as the finish point. Animals
146 were placed perpendicularly on the midpoint of the beam and allowed to traverse the beam for 120 s. A score
147 to the motor abilities of the animal was given: 0, correct gait; 1, clings with the 4 paws; 2, slips with one paw;
148 3, slips with two paws; falls in a time less than 60 sec [8].

149 2.5.6. Incapitance test

150 Weight bearing changes were measured using an incapitance apparatus (Linton Instrumentation, UK)
151 detecting changes in postural equilibrium after a hind limb injury [9]. The methodology was described in Di
152 Cesare Mannelli et al (2016) [10]. Data were expressed as the difference between the weight applied on the
153 limb contralateral to the injury and the weight applied on the ipsilateral one (Δ Weight).

154 2.5.7. Histological examination

155 Animals were killed by cervical dislocation on day 14 after the behavioral measurements. Legs were cut
156 under the knee, flayed and fixed in 4% formaldehyde in phosphate-buffered saline (PBS) for 48 h at room
157 temperature. Subsequently, samples were decalcified by 0.76 M sodium formate and 1.6 M formic acid
158 solution in H₂O for 4 weeks with a change of solution every 7 days. At the end of decalcification, these
159 samples were routinely dehydrated in alcohol and embedded in paraffin. Sections (6 μ M) thick were
160 observed and an histological score (0: absent; 1: mild; 2: moderate; 3: severe) was attributed to the following
161 morphological parameters: (a) inflammatory infiltrate; (b) synovial hyperplasia; (c) fibrin deposition; (d)
162 synovial vascularity; (e) cartilage erosion; (f) bone erosion; (g) joint space [11,12].

163 2.6. Statistical analysis

164 All experimental results are given as mean \pm S.E.M. Analysis of variance was followed by Fisher's post
165 hoc comparison to verify the significance between two means. Data were analyzed with the StatView
166 software for the Macintosh (1992). P values of less than 0.05 were considered statistically significant.

167 3. Results

168 3.1. DCL-DL characterization

169 Liposomes were characterized in terms of vesicle size, zeta potential and EE. As results showed, SL and
170 DCL-DL were bigger than EL; however, only DCL-DL liposomes showed a significant difference ($p=0.0142$)
171 with the EL system. On the other hand, no statistical significance was found between Z potential of the



172 different samples. Regarding EE, although SL had a higher percentage of Cur entrapped, there was no
 173 statistical significance between DCL-DL and SL.

174 This section may be divided by subheadings. It should provide a concise and precise description of the
 175 experimental results, their interpretation as well as the experimental conclusions that can be drawn.

176

177 **Table 1.** Characterization results * p<0.05, **p<0.01, ***p<0.001 compared with empty liposomes (EL).

Liposomes	Particle size ± s.d. (nm)	Z potential ± s.d. (mV)	EE ± s.d. (%)
SL	3259 ± 568	27,2 ± 3,5	94,8 ± 2,8
DCL-DL	4325 ± 462*	31,7 ± 2,8	92,2 ± 3.8
EL	3057 ± 257	24,5 ± 8,3	-

178

179 3.2. *In vivo* test

180 Table 2 shows the results of the different *in vivo* tests performed. Paw pressure test measured the force
 181 that the animal could stand before vocalizing or moved away from its leg. The more weight the paw was
 182 able to handle, less inflamed was the joint [13]. It was clear that MIA+DCL-DL group had a significantly
 183 higher pain tolerance when compared with the MIA group, which indicates that DCL-DL were able to
 184 reduce joint inflammation. After 7 days of treatment, joint pain reduction was extremely significant in the
 185 MIA+DCL-DL group when compared with sham group (p=0.0001), however, on day 14 this difference was
 186 reduced, although it was still very significantly different (p=0.0074). Considering MIA+ SL and MIA+ EL
 187 groups, results were unexpected. MIA+ SL group showed more pressure tolerance when compared with
 188 MIA group at day 7, but this tendency did not continue at day 14. MIA+ EL group, at day 7 showed a lower
 189 pressure tolerance when compared with MIA group and highly ameliorated at day 14, resulting in a
 190 pressure tolerance very significantly higher than that of OA group at day 14.

191 Beam balance test studies the ability of the animal to properly walk. As was mentioned before, it is
 192 scored 0-4 depending on animal's gait. The higher the score, the worst the gait is MIA+ DCL-DL group
 193 showed an extremely significant improvement on walking skills in days 7 and 14 when compared with MIA
 194 group. On day 14, animals treated with DCL-DL had the same ability to walk than sham group (p=0.1607).
 195 MIA+ EL and MIA+ SL groups did not show significant differences with MIA group except EL group at day
 196 14, which showed significantly better walking abilities than OA group at day 14.

197 Incapacitance test measured the differences in weight that animals place on their hind limbs. Animals
 198 with no limb issues should have minimal differences between both legs. As table 2 depicts, MIA+ EL and
 199 MIA+ DCL-DL groups had similar weight differences compared with MIA group. However, MIA+ DCL-DL
 200 group showed an extremely significant improvement on it, although, not even at day 14 results were close to
 201 those of sham group. Nonetheless, animals treated with DCL-DL ameliorated with time (p=0.0096).

202

203 **Table 2.** *In vivo* test results. Table shows paw pressure test on treated limb (weight the paw resists before
 204 vocalization or movement), beam balance test and incapacitance test (differential weight between non-treated
 205 and treated limb measures in grams) *p<0.05, **p<0.01, ***p<0.001 when data were compared with sham
 206 group. ^p<0.05, ^^p<0.01, ^^<p<0.001 when data were compared with MIA group.

Treatment	Paw pressure test (g)		Beam balance test (0-4)		Incapacitance test (g)
	Day 7	Day 14	Day 7	Day 14	Day 7
Sham	65.4 ± 0.8	67.6 ± 2.1	0.2 ± 0.2	0.3 ± 0.3	5.4 ± 0.6
MIA	41.7 ± 1.7***	46.7 ± 1.7***	3.2 ± 0.4***	2.8 ± 0.3***	51.0 ± 2.6***
MIA+EL	39.2 ± 0.8^	51.7 ± 1.7^^	3.5 ± 0.4	1.9 ± 0.4^	46.8 ± 4.8
MIA+SL	48.3 ± 4.4^	46.7 ± 4.2	2.9 ± 0.3	2.7 ± 1.7	52.3 ± 6.4
MIA+DCL-DL	55.3 ± 1.5^^^	62.6 ± 1.4^^^	1.3 ± 0.3^^^	0.7 ± 0.4^^^	32.5 ± 4.0^^^

207



218 After 14 days, animals were sacrificed and histological tests were performed. Results (data not shown)
219 highlighted, as expected, the ability of MIA to trigger OA. All parameters studied were significantly higher
220 when compared with those of the sham group. Regarding MIA+ EL group, no significant differences were
221 found when compared with OA group. MIA+DCL-DL group showed protective properties at least regarding
222 some aspects of OA.

223 4. Discussion

224 In the present work, an attempt has been made to elucidate if Cur has anti-inflammatory properties
225 applicable to OA. For this purpose, Cur-in HP β CD-in liposome in a double-loaded system was employed.
226 This formulation was previously developed by our research group [5]; however, major modifications had to
227 be made as the original formula contained didodecyldimethylammonium bromide (DDAB), a cationic lipid
228 that provides the liposome stability over time. Preliminary studies revealed that liposomes with DDAB
229 triggered joint swelling that did not disappear during the experiment (data not shown). On the other hand,
230 positively charged lipids are known for their cell toxicity [14,15] and then, for these reasons, DDAB was
231 removed from the formulations.

232 Analyzing the physicochemical characteristics, liposomes containing Cur are bigger than those empty.
233 However, no statistical differences were found between SL and DCL-DL, indicating that the presence of Cur
234 in both compartments do not affect the size of the liposomes. Regarding EE, it was higher than expected for
235 DCL-DL. Previous work on this field yielded 52.38% for DCL-DL, which is smaller than EE of DCL-DL in the
236 present study (92.25%), indicating that the absence of DDAB highly improves EE.

237 The main objective of this paper was to study if the developed innovative Cur liposomal formulation
238 ameliorated OA symptoms. For this, unilateral osteoarthritis was induced in rats by MIA injection on one of
239 their legs. 7 days after injection, OA was established in all cases. Afterwards, a single dose of the different
240 samples was applied, mimicking regular OA treatment with hyaluronic acid or glucocorticoids [2]. Three
241 tests were performed to evaluate animal's walking abilities and inflammation: Randall and Selitto's paw
242 pressure test, beam balance test and incapacitance test.

243 Randall and Selitto's paw pressure test is based on the premise that inflamed zones have more
244 sensitivity to pain [13]. Thus, when pressure is applied to an inflamed area, the weight that this area can
245 handle is lower than the weight that can handle a non-inflamed area. For comparison purposes, treated and
246 untreated limbs were tested. Non-treated limbs showed similar results in all cases (data not shown).
247 Regarding treated limbs, DCL-DL showed an extremely significant improvement in pressure tolerance,
248 which indicates a marked reduction in joint inflammation. This improvement increases over time, indicating
249 that DCL-DL have better control than SL over Cur release. However, it cannot be said that this system
250 completely eliminates inflammation since there are still significant differences between MIA+ DCL-DL group
251 and sham group at day 14 ($p < 0.01$).

252 To study motor abilities of rats after treatment, a beam balance test was performed. In this case, the
253 ability to walk and balance was measured using different scores. 7 days after treatment, animals treated with
254 DCL-DL exhibited an extremely significant improvement in their walking capacity, mostly grabbing the
255 beam with two paws. On day 14, these animals improved, even more, obtaining scores equal to those of
256 untreated animals. This indicates that DCL-DL reduces inflammation to a point that allows the animal to
257 normally walk.

258 Incapacitance test is an interesting essay that allows the researcher to evaluate pain. Normally, body
259 weight is equally distributed over both hind legs; however, when one of the paws inflamed, i.e. it is painful
260 to stand on it, the animal naturally compensates by overloading the non-inflamed paw. This compensation
261 was recorded as the difference in weight between them [9,10]. Data showed that only DCL-DL group
262 ameliorated with treatment at days 7 and 14, however, there was still a significant difference with the sham
263 group. This indicates that pain is significantly reduced but has not disappeared.

264 Surprisingly, SL showed a much lower capability to reduce inflammation compared with DCL-DL. As
265 SL do not contain HP β CD in the aqueous compartment, the amount of Cur able to be released from the



256 liposomes is inferior to that of DCL-DL [5]. As a result, SL showed only very limited anti-inflammatory
257 properties, evidencing the critical role of an effective drug formulation to fully exploit its therapeutic action.

258 5. Conclusions

259 DCL-DL is a novel approach that has been recently developed for allowing Cur intra-articular
260 administration. However, the presence of DDAB in the liposome bilayer of such formulation hinders its use
261 in vivo, due to its irritating action and potential cell toxicity. In the present work, DDAB was removed from
262 the liposomal formulation and the new DCL-DL system ability to ameliorate OA was tested in vivo in a MIA
263 rat model in comparison with a conventional single-loaded Cur liposomal formulation (SL).

264 From the results obtained, we can conclude that DCL-DL successfully reduced pain and inflammation
265 in OA joint with a prolonged action up to 14 days from administration, differently from SL that exhibited
266 only minimal effects in inflammation reduction and walking ability improvement of the animals.

267
268 **Author Contributions:** F.M., M.L.G-R, A.M.F-R, L.C.M. and L.M. conceived and designed the experiments; F.M., A.M.F-
269 R, L.M. and C.G. performed the experiments; L.M., L.C.M., C.G. and A.M.F-R analyzed the data; F.M., M.L.G-R, A.M.F-R,
270 L.C.M., A.M.R. and P.A.M. wrote the paper.

271 **Conflicts of Interest:** The authors declare no conflict of interest.

272 Abbreviations

273 The following abbreviations are used in this manuscript:

- 274 Cur: Curcumin
275 DCL-DL: Drug in Cyclodextrin-in-liposome double-loaded liposomes
276 SL: Single-loaded liposomes
277 EL: Empty liposomes
278 DDAB: Didodecyldimethylammonium bromide
279 HP β CD: Hydroxypropyl- β -cyclodextrin
280 OA: Osteoarthritis
281 MIA: Monoiodoacetate

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