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UCL Institute of  
Healthcare Engineering

**1st International Electronic Conference on Pharmaceuticals 2020**

# Advanced Manufacturing Research for Healthcare

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## Invited Special Issue

06<sup>th</sup> August 2019

24 key papers by key leading researchers covering every part of the world



## Microbubble size reduction

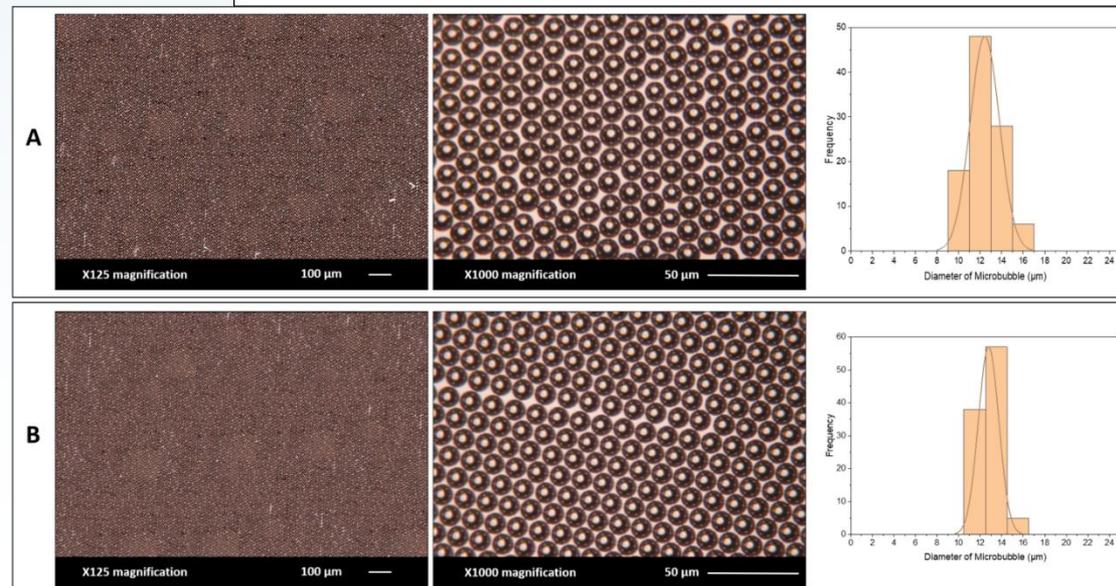
**(A): Triple (3) T-junction**  
 Capillary size: **65  $\mu\text{m}$**   
 Channel gap size: **65  $\mu\text{m}$**   
 Gas Pressure: **80 kPa**  
 BSA solution: **10 wt %**  
 Flow Rate: **650  $\mu\text{L} / \text{min}$**

The mean MB size is:  
**12.4  $\mu\text{m}$**

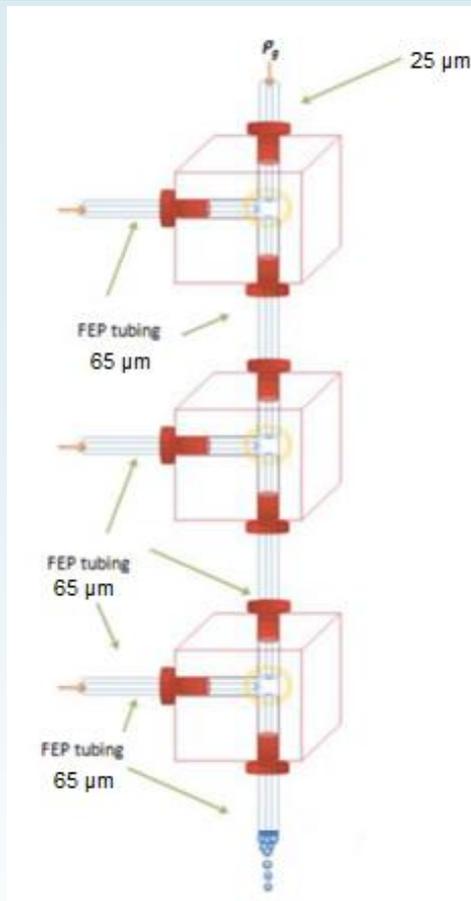


**(B): Triple (3) T-junction**  
 Capillary size: **25  $\mu\text{m}$**   
 Channel gap size: **25  $\mu\text{m}$**   
 Gas Pressure: **80 kPa**  
 BSA solution: **5 wt %**  
 Flow Rate: **400  $\mu\text{L} / \text{min}$**

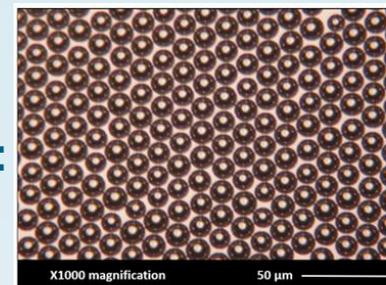
The mean MB size is:  
**12.8  $\mu\text{m}$**



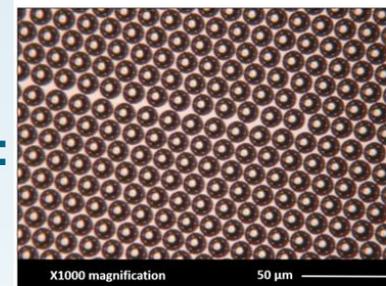
**(C): Triple (3) T-junction**  
 Capillary size: **25  $\mu\text{m}$**  (gas inlet) and **65  $\mu\text{m}$**  (rest)  
 Channel gap size: **65  $\mu\text{m}$**   
 Gas Pressure: **80 kPa**  
 BSA solution: **10 wt %**  
 Flow Rate: **700  $\mu\text{L} / \text{min}$**



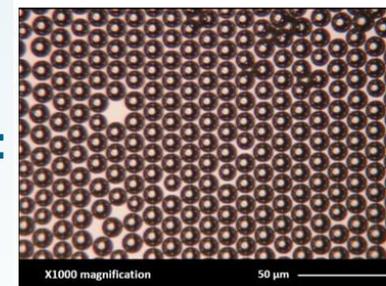
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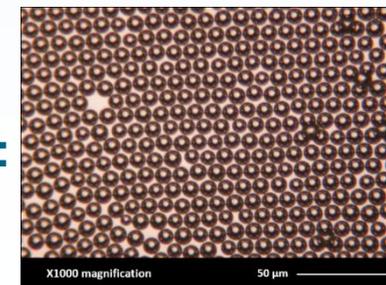
**(B):**



**(C):**



**(D):**



**(C):** The mean MB size is:  
**9.9  $\mu\text{m}$**

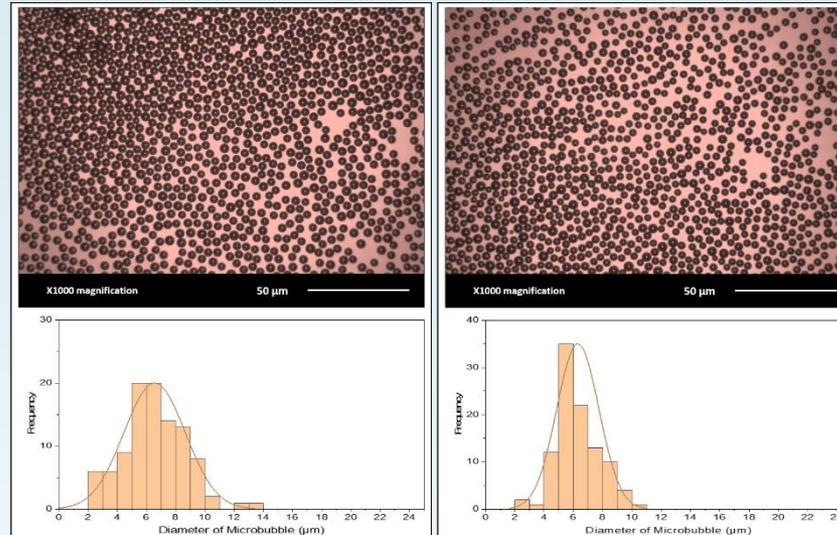
The mean MB size is:  
**8.7  $\mu\text{m}$**

**(D): Quadruple (4) T-junction**  
 Capillary size: **100  $\mu\text{m}$**   
 Channel gap size: **100  $\mu\text{m}$**   
 Gas Pressure: **80 kPa**  
 BSA solution: **10 wt %**  
 Flow Rate: **700  $\mu\text{L} / \text{min}$**

# Microbubble size reduction with fission processes:

**Quadruple (4)** T-junction  
 Capillary size: **65  $\mu\text{m}$**   
 Channel gap size: **100  $\mu\text{m}$**   
 Gas Pressure: **80 kPa**  
 BSA solution: **10 wt %**  
 Flow Rate: **600  $\mu\text{L} / \text{min}$**

The mean MB size is:  
**6.6  $\mu\text{m}$**

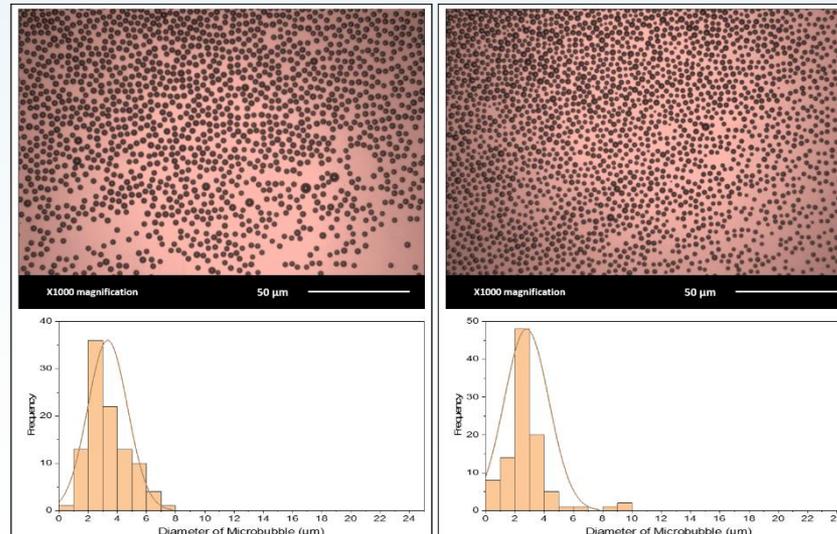


**Quadruple (4)** T-junction  
 Capillary size: **25  $\mu\text{m}$**   
 Channel gap size: **50  $\mu\text{m}$**   
 Gas Pressure: **80 kPa**  
 BSA solution: **15 wt %**  
 Flow Rate: **700  $\mu\text{L} / \text{min}$**

The mean MB size is:  
**6.30  $\mu\text{m}$**

**Quadruple (4)** T-junction  
 Capillary size: **25  $\mu\text{m}$  (gas inlet) and 100  $\mu\text{m}$  (rest)**  
 Channel gap size: **150  $\mu\text{m}$**   
 Gas Pressure: **80 kPa**  
 BSA solution: **15 wt %**  
 Flow Rate: **700  $\mu\text{L} / \text{min}$**

The mean MB size is:  
**3.3  $\mu\text{m}$**



**Quadruple (4)** T-junction  
 Capillary size: **25  $\mu\text{m}$  (gas inlet) and 65  $\mu\text{m}$  (rest)**  
 Channel gap size: **100  $\mu\text{m}$**   
 Gas Pressure: **80 kPa**  
 BSA solution: **10 wt %**  
 Flow Rate: **650  $\mu\text{L} / \text{min}$**

The mean MB size is:  
**2.80  $\mu\text{m}$**

## Fission processes:

- MB size can be further reduced by fission process. (But lose the monodispersity of MBs)
- When the channel gap size is greater than the diameter of capillary, fission process occurred

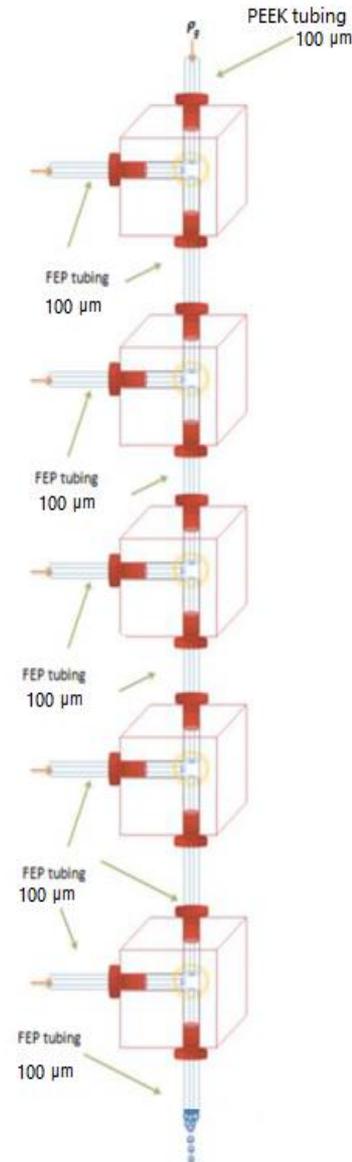
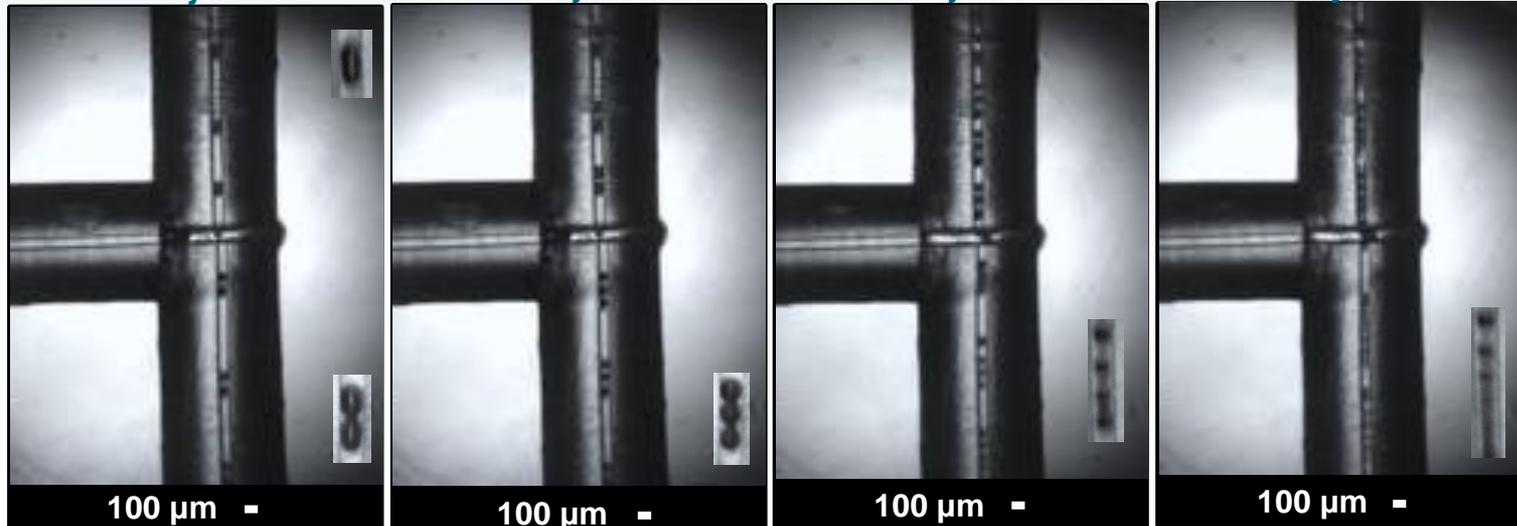
**Quintuple (5) T-junction**  
 Capillary size: **100  $\mu\text{m}$**   
 Channel gap size: **150  $\mu\text{m}$**   
 Gas Pressure: **80 kPa**  
 BSA solution: **15 wt %**  
 Flow Rate: **500  $\mu\text{L} / \text{min}$**

2<sup>nd</sup> T-junction

3<sup>rd</sup> T-junction

4<sup>th</sup> T-junction

5<sup>th</sup> T-junction

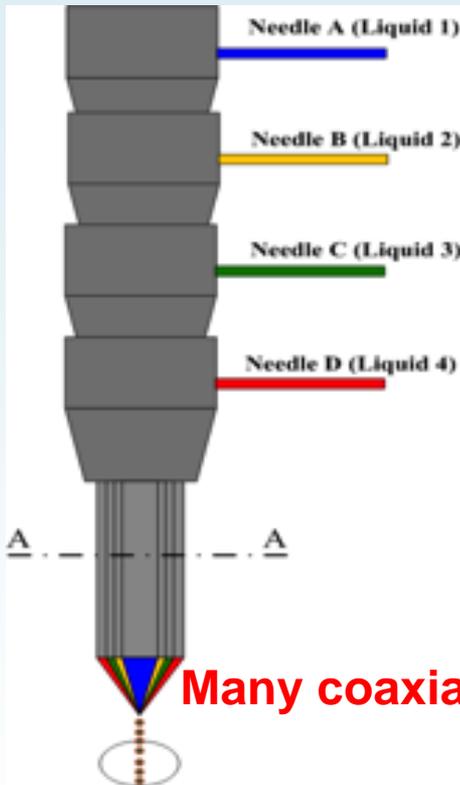


# Comparison study:

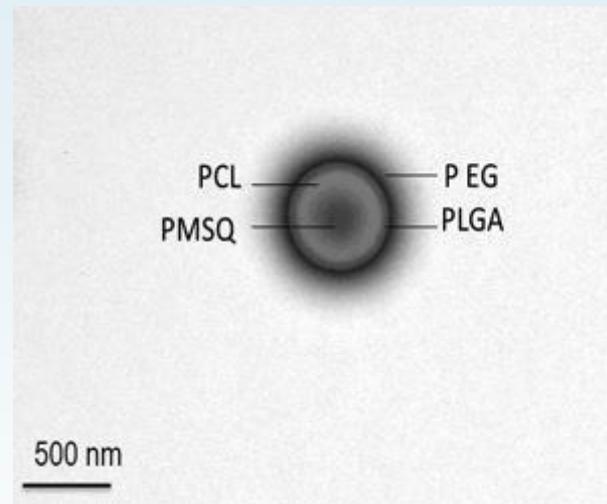
<b>Capillary size:</b>	100 $\mu\text{m}$	65 $\mu\text{m}$	25 $\mu\text{m}$	25 $\mu\text{m}$ for gas inlet 65 $\mu\text{m}$ for rest capillaries	
<b>Channel gap distance:</b>	100 $\mu\text{m}$	65 $\mu\text{m}$	25 $\mu\text{m}$	65 $\mu\text{m}$	
<b>Fission Process</b>	Not occurred	Not occurred	Not occurred	Not occurred	
<b>The smallest MB size:</b>	22.8 $\pm$ 1.4 $\mu\text{m}$	12.4 $\pm$ 1.2 $\mu\text{m}$	12.8 $\pm$ 1.7 $\mu\text{m}$	9.9 $\pm$ 1.3 $\mu\text{m}$	
<b>The optimal conditions:</b>	Triple (3) T-junction Gas Pressure: 80 kPa BSA solution: 15 wt % Flow Rate: 800 $\mu\text{L min}^{-1}$	Triple (3) T-junction Gas Pressure: 80 kPa BSA solution: 10 wt % Flow Rate: 650 $\mu\text{L min}^{-1}$	Triple (3) T-junction Gas Pressure: 80 kPa BSA solution: 5 wt % Flow Rate: 400 $\mu\text{L min}^{-1}$	Triple (3) T-junction Gas Pressure: 80 kPa BSA solution: 10 wt % Flow Rate: 700 $\mu\text{L min}^{-1}$	
<b>lifetime of MBs (without adding SiQD):</b>	40.0 $\pm$ 2.0 mins	25.0 $\pm$ 1.5 mins	5.0 $\pm$ 0.5 mins	25.0 $\pm$ 2.5 mins	
<b>Capillary size:</b>	100 $\mu\text{m}$	65 $\mu\text{m}$	25 $\mu\text{m}$	25 $\mu\text{m}$ for gas inlet 100 $\mu\text{m}$ for rest capillaries	25 $\mu\text{m}$ for gas inlet 65 $\mu\text{m}$ for rest capillaries
<b>Channel gap distance:</b>	100 $\mu\text{m}$	100 $\mu\text{m}$	50 $\mu\text{m}$	150 $\mu\text{m}$	100 $\mu\text{m}$
<b>Fission Process</b>	Not occurred	Occurred	Occurred	Occurred	Occurred
<b>The smallest MB size:</b>	8.7 $\pm$ 0.8 $\mu\text{m}$	6.6 $\pm$ 2.1 $\mu\text{m}$	6.3 $\pm$ 1.4 $\mu\text{m}$	3.3 $\pm$ 1.4 $\mu\text{m}$	2.8 $\pm$ 1.5 $\mu\text{m}$
<b>The optimal conditions:</b>	Quadruple (4) T-junction Gas Pressure: 80 kPa BSA solution: 15 wt % Flow Rate: 800 $\mu\text{L min}^{-1}$	Quadruple (4) T-junction Gas Pressure: 80 kPa BSA solution: 10 wt % Flow Rate: 600 $\mu\text{L min}^{-1}$	Quadruple (4) T-junction Gas Pressure: 80 kPa BSA solution: 5 wt % Flow Rate: 300 $\mu\text{L min}^{-1}$	Quadruple (4) T-junction Gas Pressure: 80 kPa BSA solution: 15 wt % Flow Rate: 700 $\mu\text{L min}^{-1}$	Quadruple (4) T-junction Gas Pressure: 80 kPa BSA solution: 10 wt % Flow Rate: 650 $\mu\text{L min}^{-1}$
<b>lifetime of MBs (without adding SiQD):</b>	35.0 $\pm$ 4.0 mins	20.0 $\pm$ 2.0 mins	5.0 $\pm$ 1.0 mins	20.0 $\pm$ 1.0 mins	15.0 $\pm$ 4.5 mins

# Multi-Layer Capsules

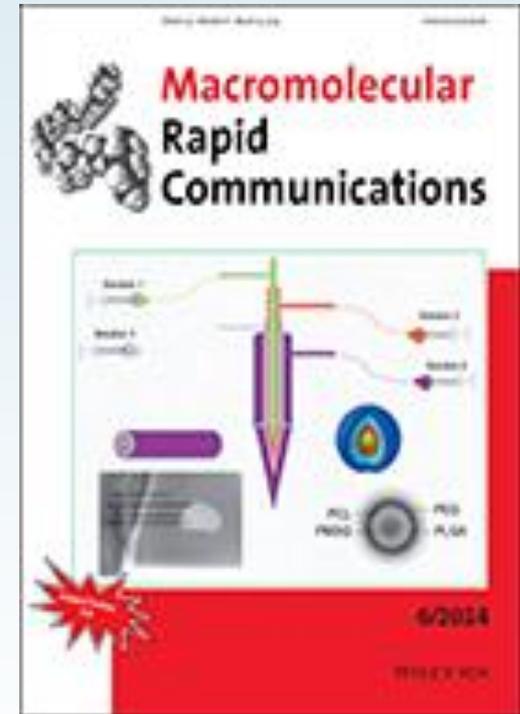
(prepared using electrohydrodynamics)



**Many coaxial Needles**



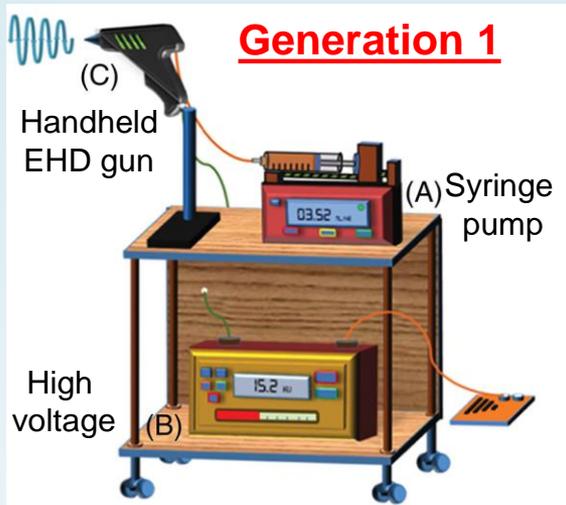
VENTURE PRIZE 2010



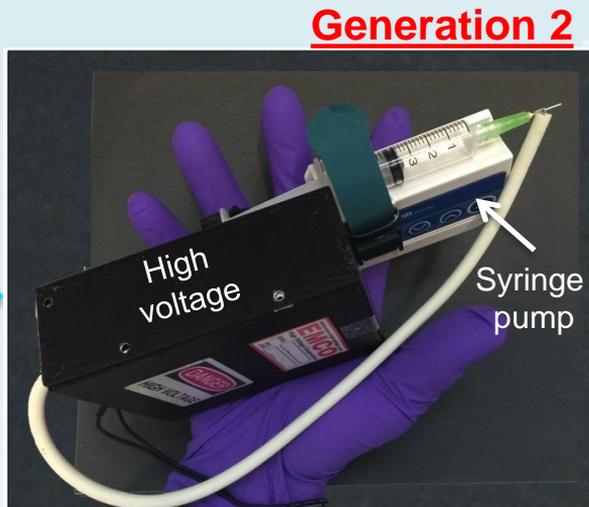
Labaf S, Ghanbar H, Stride E, Edirisinghe M. Preparation of multilayered structures using a novel four-needle coaxial device. *Macromolecular Rapid Communication* 2014 35: 618-623 (front cover article)

UK Patent Application No. 1102148.2 PCT filed: 08Feb 2012

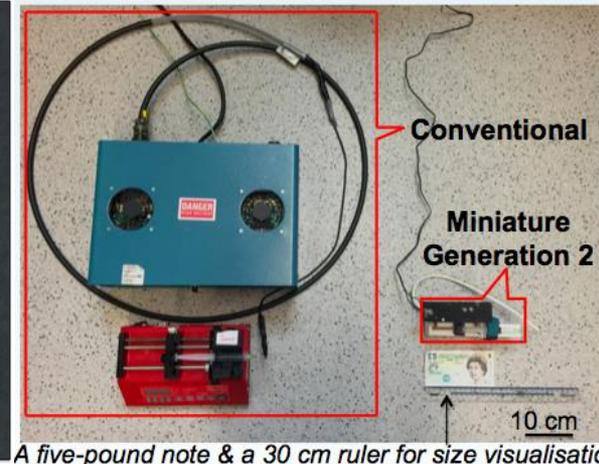
USA Patent Application 13/984345



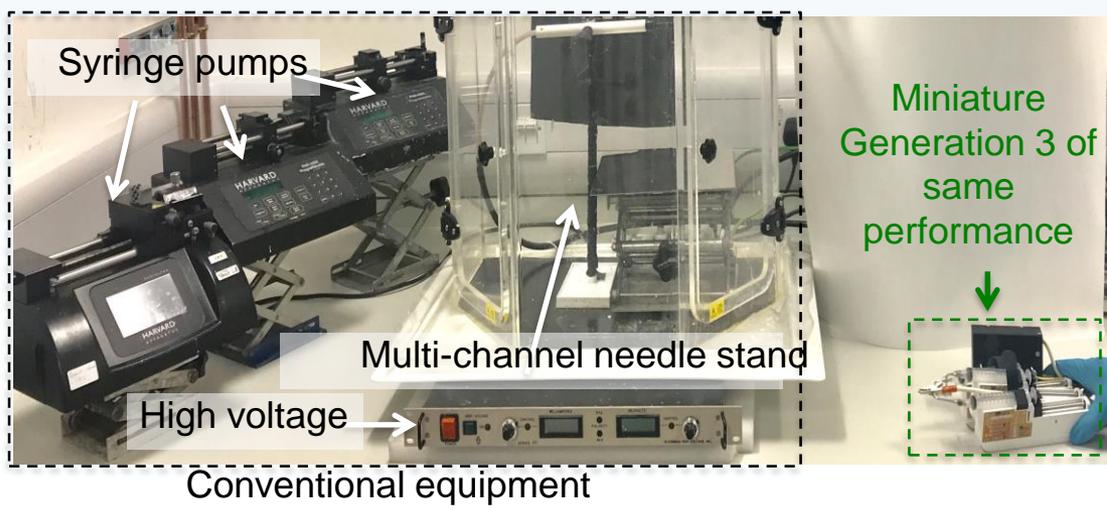
Sofokleous, P.; Stride, E.; Bonfield, W.; **Edirisinghe, M.** *Mater. Sci. Eng. C Mater. Biol. Appl.* **2013**, 33 (1), 213–223.



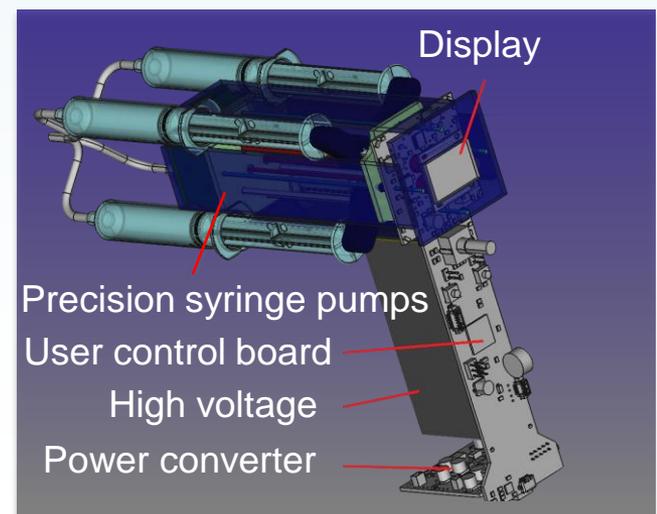
F. Brako, C.J. Luo, D.Q.M. Craig, **M. Edirisinghe.** *Macromolecular Materials and Engineering*, **2018**, 303 (5), 1700586.



## Generation 3

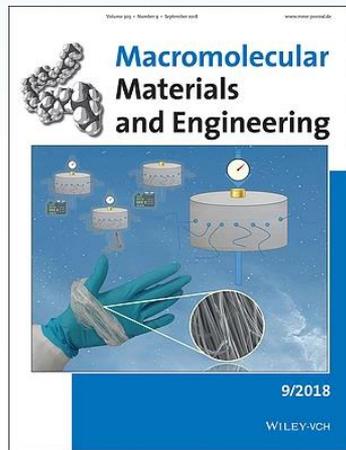
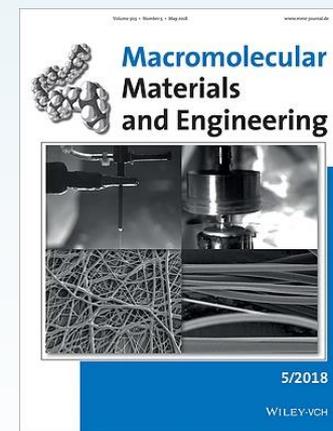
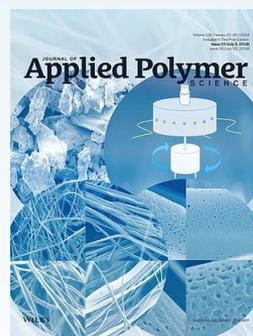
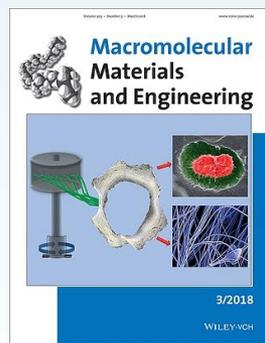
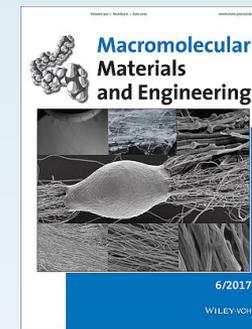
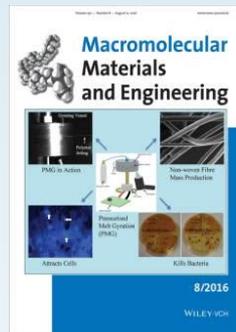
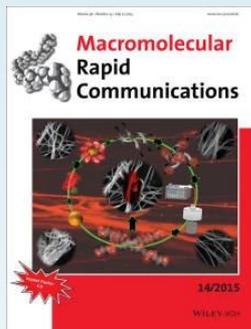
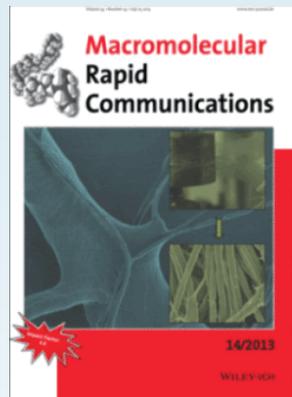


## Generation 4



# Pressurised Gyration & SISTER PROCESSES

**EPSRC grant EP/L023059/1:** Exploitation of Pressurised Gyration as an Innovative Manufacturing Route for Nanofibrous Structures. Industrial Support: BASF, Astra Zeneca  
(Collab: Professor Duncan Craig, UCL School of Pharmacy)



Invited Feature: 2018

BEST OF MACROS 2018

Featured in Advanced Science News July 23<sup>rd</sup> 2018

BEST OF MACROS 2019

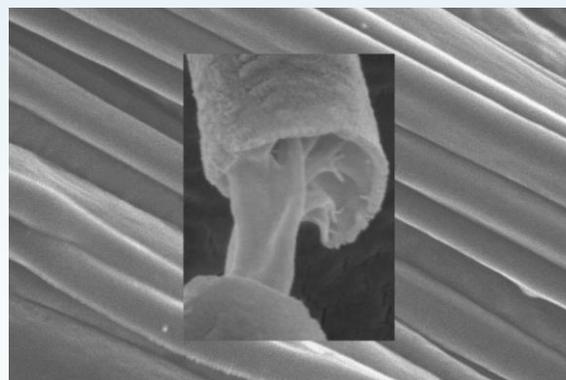
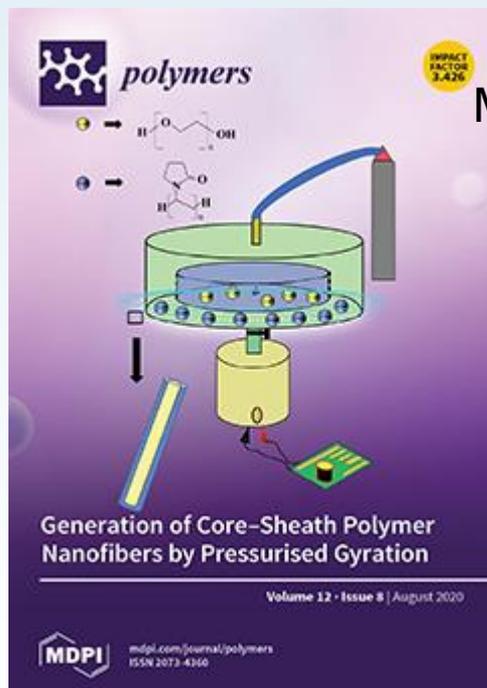
**EP/S016872/1:**

**Creation and Exploitation of Pressurised Gyration to Manufacture Core-Sheath Structures**

Mahalingam, S; Homer-Vanniasinkam, S; Edirisinghe, M; (2019) Novel pressurised gyration device for making core-sheath polymer fibres. **Materials & Design** , 178 , Article 107846. [10.1016/j.matdes.2019.107846](https://doi.org/10.1016/j.matdes.2019.107846).

**Polymers** 2020, 12, 1709; doi:10.3390/polym12081709

Mahalingam, S; Huo, S; Homer-Vanniasinkam, S; Edirisinghe, M



**UPI - SCIENCE NEWS**

OCT. 14, 2020 / 9:17 AM

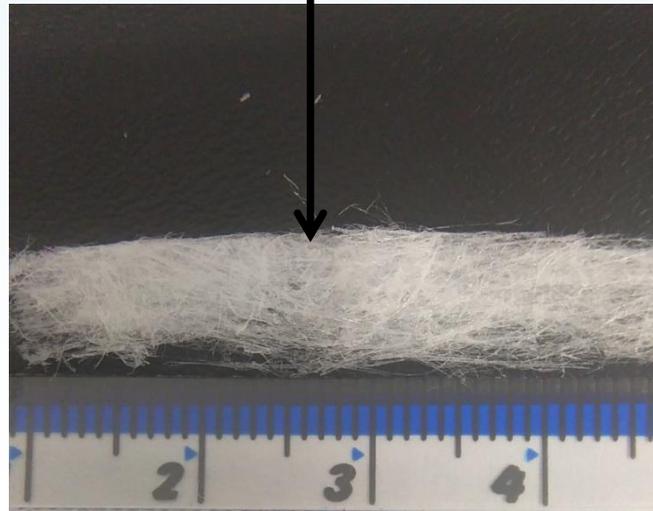
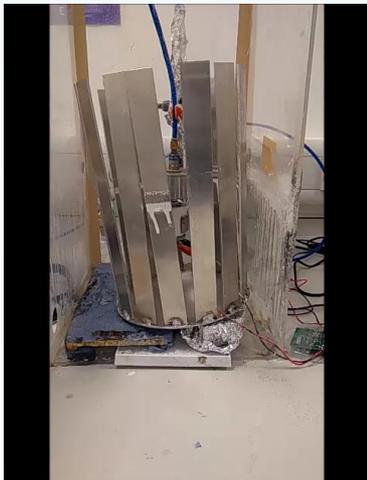
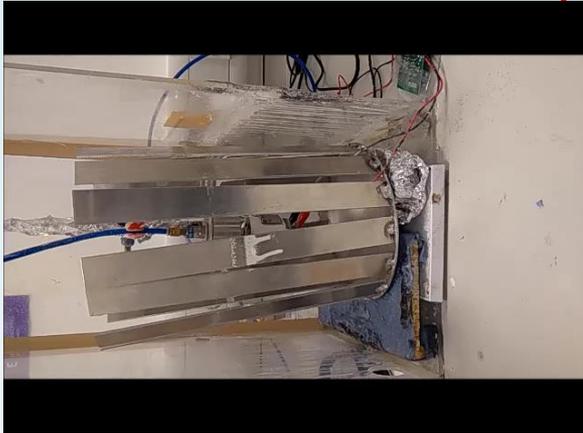
**Layered hybrid fibers could be used to build anti-viral masks, researchers say**

Current methodologies and approaches for the formation of core–sheath polymer fibers for biomedical applications

S. Mahalingam, R. Matharu, S. Homer-Vanniasinkam and M. Edirisinghe

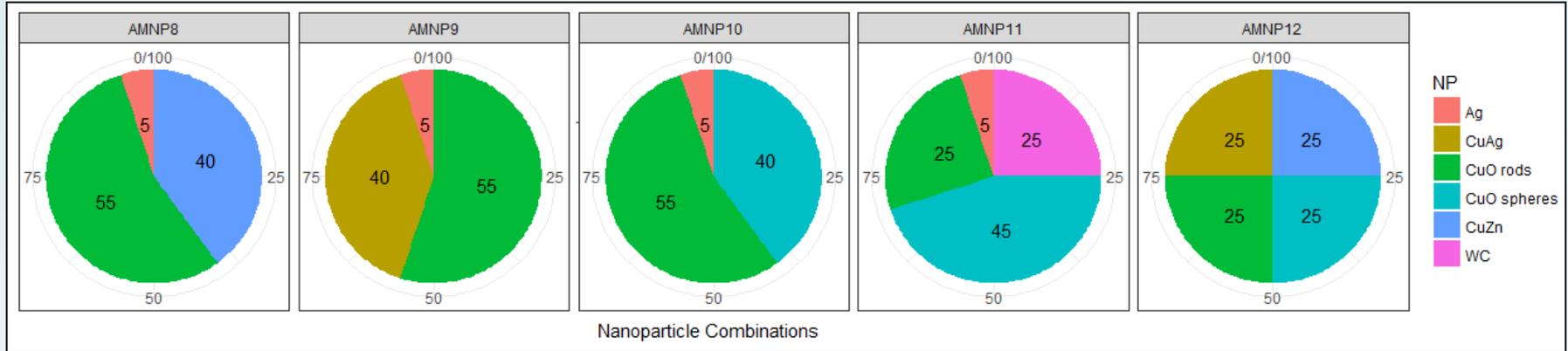
Applied Physics Reviews **7**, 041302 (2020); <https://doi.org/10.1063/5.0008310>

## Mark-II of device (2015-16)

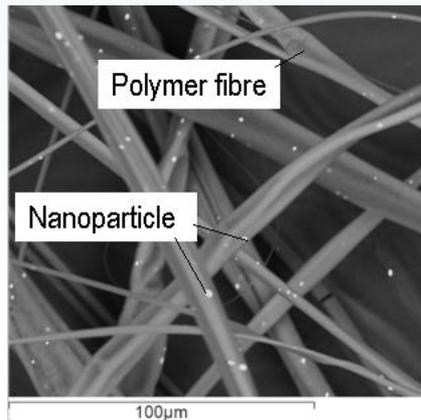
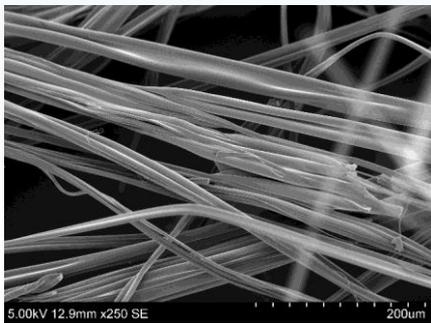


## Novel Antimicrobial Filters

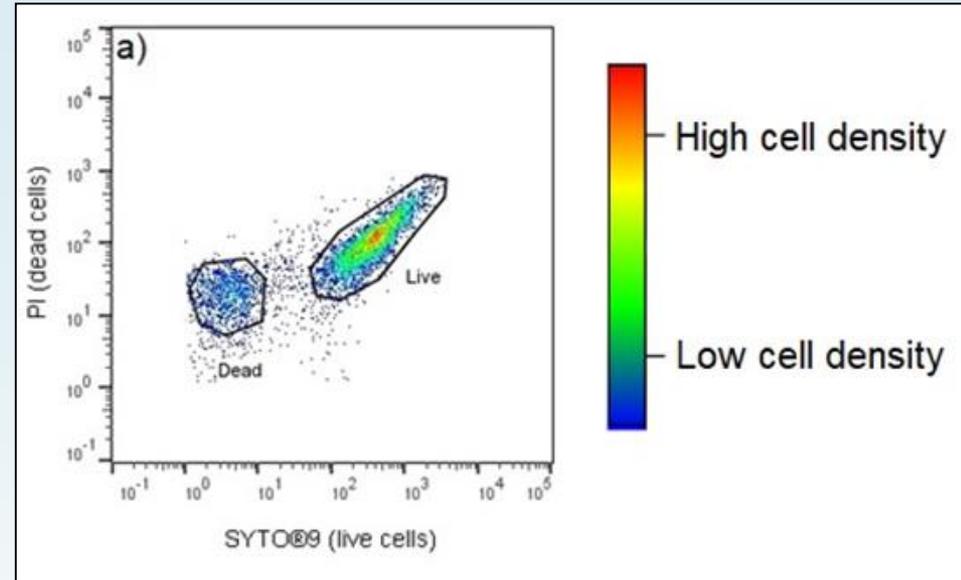
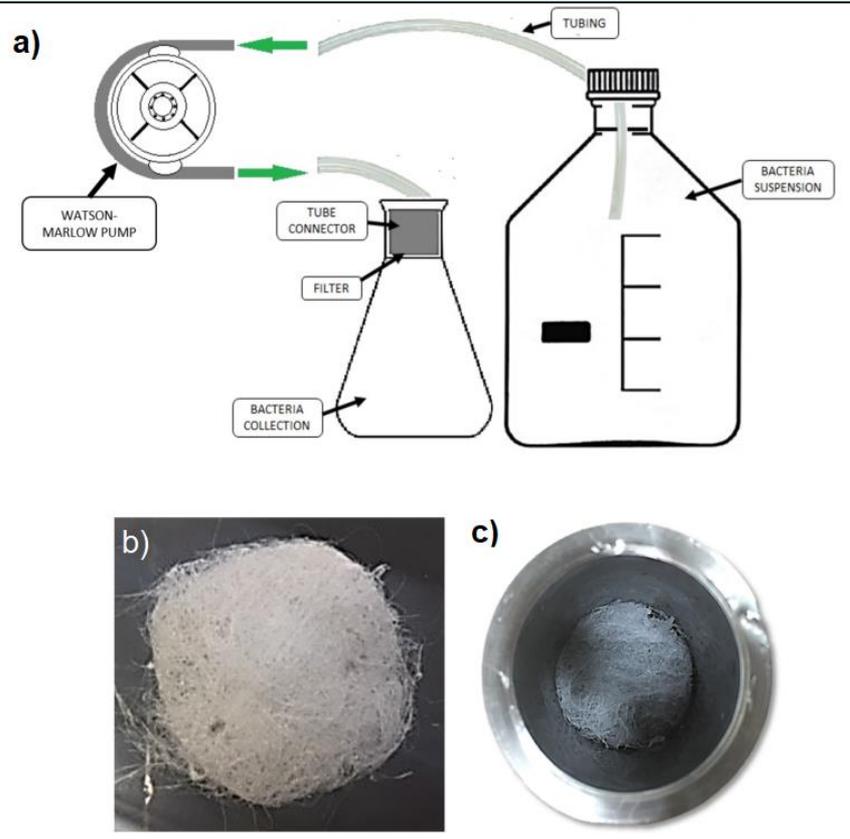
A number of nanoparticle preparations were assessed for their antimicrobial properties.



Nanocomposite filters were developed using pressurised gyration, during which poly(methylmethacrylate) (PMMA) solutions were loaded with **antimicrobial nanoparticles** and processed into nanofiber meshes.



## Novel Antimicrobial Filters



The viability of bacterial populations were quantified and analysed using **flow cytometry** to determine live and dead cells after filtration.

### Project Partners:

**GAMA Healthcare Ltd**

**Intrinsiq Materials Limited**

**Pall Corporation**

**Pathogen Solutions**

**Professor John Oxford**

Filters were tested using phosphate buffered saline solution spiked with high concentrations of *Pseudomonas aeruginosa* and *Staphylococcus aureus*. The bacterial suspensions were passed through a peristaltic pump at **500 mL min<sup>-1</sup>** to simulate standard UK tap pressure.

## Novel Antimicrobial Filters



AMNP8.1 filters showed reproducible results upon reuse every 30 minutes for 6 hours.

### Project Partners:

**GAMA Healthcare Ltd**

**Intrinsiq Materials Limited**

**Pall Corporation**

**Pathogen Solutions**

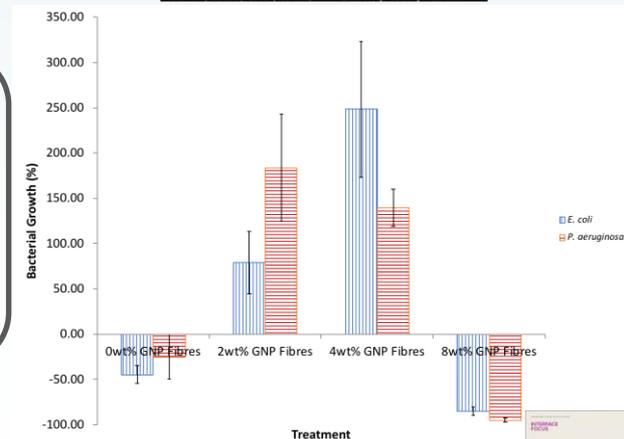
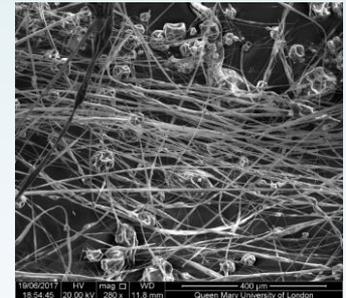
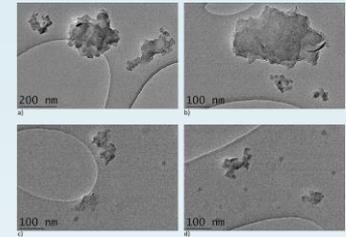
**Professor John Oxford**

# The Effect of Graphene-Poly(methyl methacrylate) Fibres on Microbial Growth.

Graphene nanoplatelets (GNPs) are the most **recently discovered** carbon-based nanomaterial. They are the two-dimensional counterpart of carbon nanotubes and are composed of a **single layer of sp<sup>2</sup> hybridized carbon atoms** arranged in a regular hexagonal lattice.

The microbial properties of a novel class of ultra-thin PMMA fibres containing **2, 4 and 8 wt% of (GNPs)** were studied. A series of polymer solutions loaded with GNPs were prepared and processed into continuous, tubular and smooth nanofiber meshes using pressurised gyration. As GNP concentration increased, the average fibre diameter increased from **0.75  $\mu\text{m}$  to 2.71  $\mu\text{m}$** .

Gram-negative bacteria *Escherichia coli* and *Pseudomonas aeruginosa* were used to investigate the bacterial properties of the composite fibres. Pure PMMA fibres were used as the control. The presence of **2 and 4 wt% GNP loaded fibres promoted microbial growth**, whilst **8 wt% GNP loaded fibres showed antimicrobial activity**.



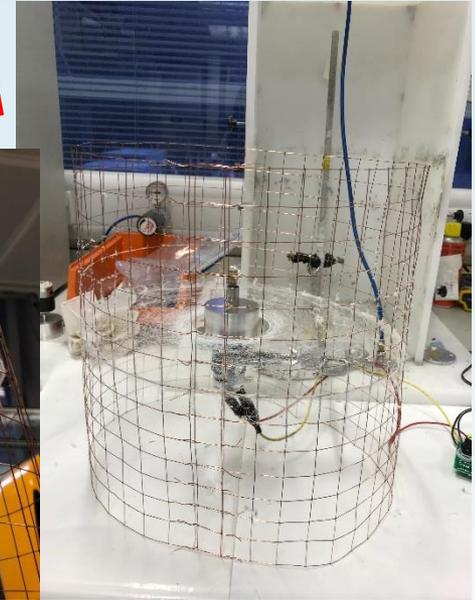
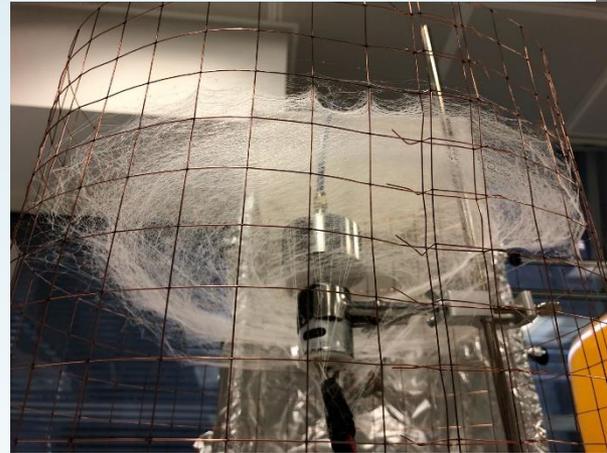
Matharu, R. K., Porwal, H., Ciric, L. and Edirisinghe, M. (2018). The Effect of Graphene-Poly(methyl methacrylate) Fibres on Microbial

Growth. *Royal Society Interface Focus*, 8, 20170058; doi:10.1098/rsfs.2017.0058. (Special issue on graphene in Biomed. Eng.)





- **Air Flow!!!**
- **Bigger Collector**  
(from 150 to 300 mm in diameter)

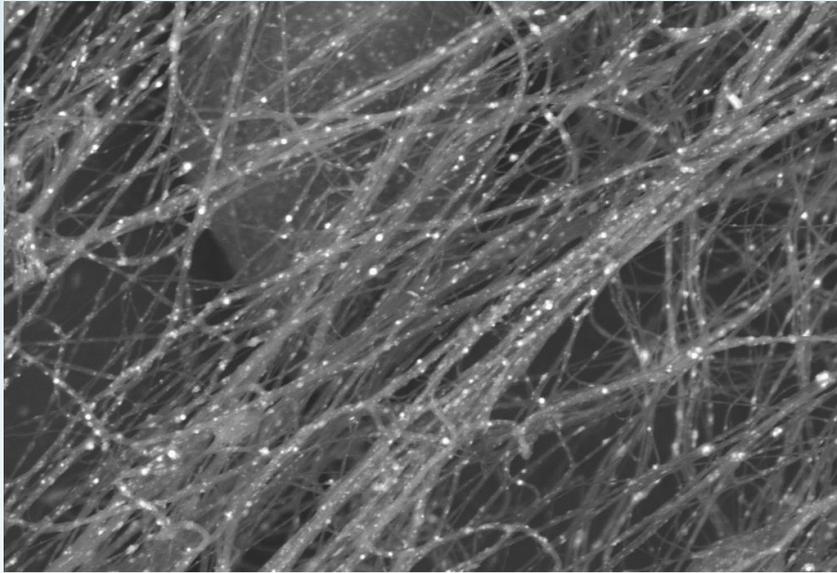


Sample Series	Relative Humidity (%)	Temperature (°C)
100 PLA to 100 BC	41.4 - 45.3	23.1 – 24.8
100 PCL to 100 BC	42.6 - 45.7	22.2 - 23.7
PLA:PCL	51.3 – 54.5	21.3 – 25.8
PLA+PCL : BC (70:30 wt ratio)	48.5 - 51.9	20.1 – 23.6



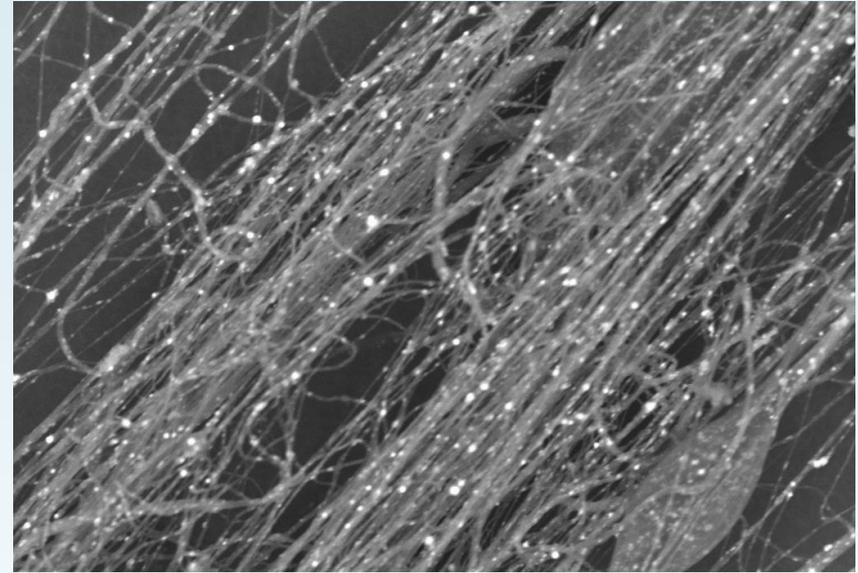
**PLA:PCL samples from collector to bandage form**

# PEO 15 wt.% + HA 30 wt.% at 36000 rpm and 0.3 MPa



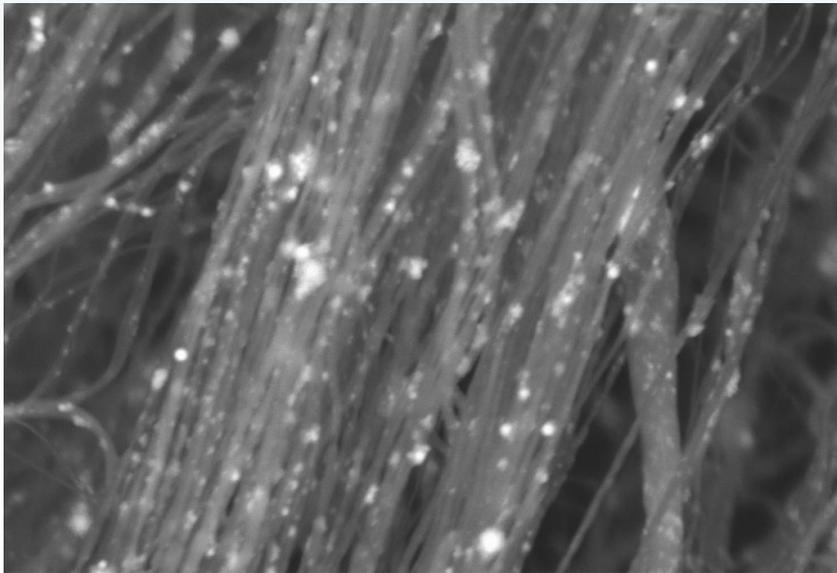
40µm

Electron Image 1



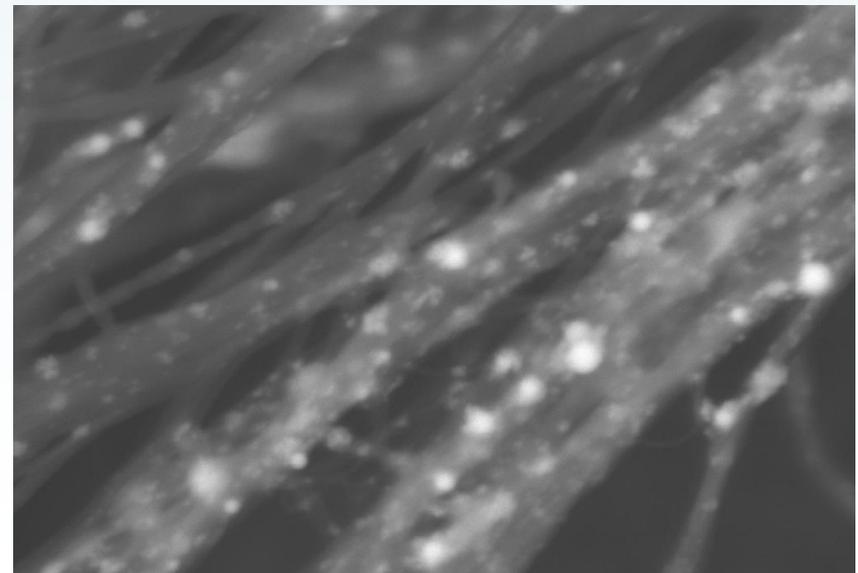
40µm

Electron Image 1



10µm

Electron Image 1

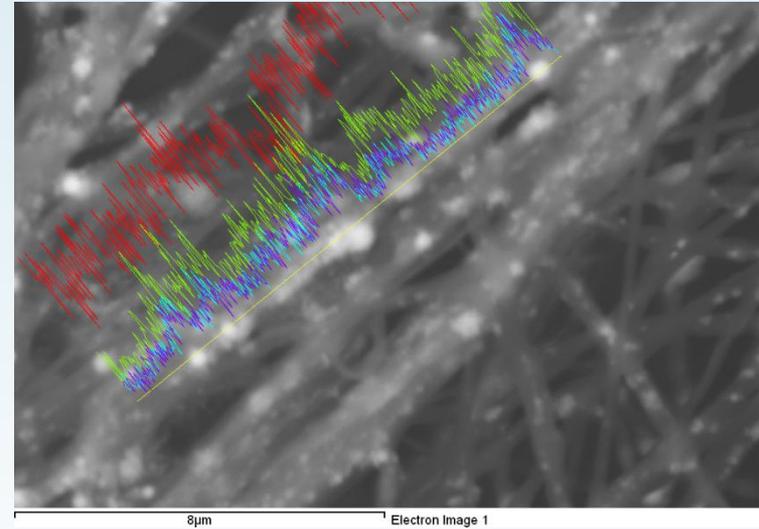
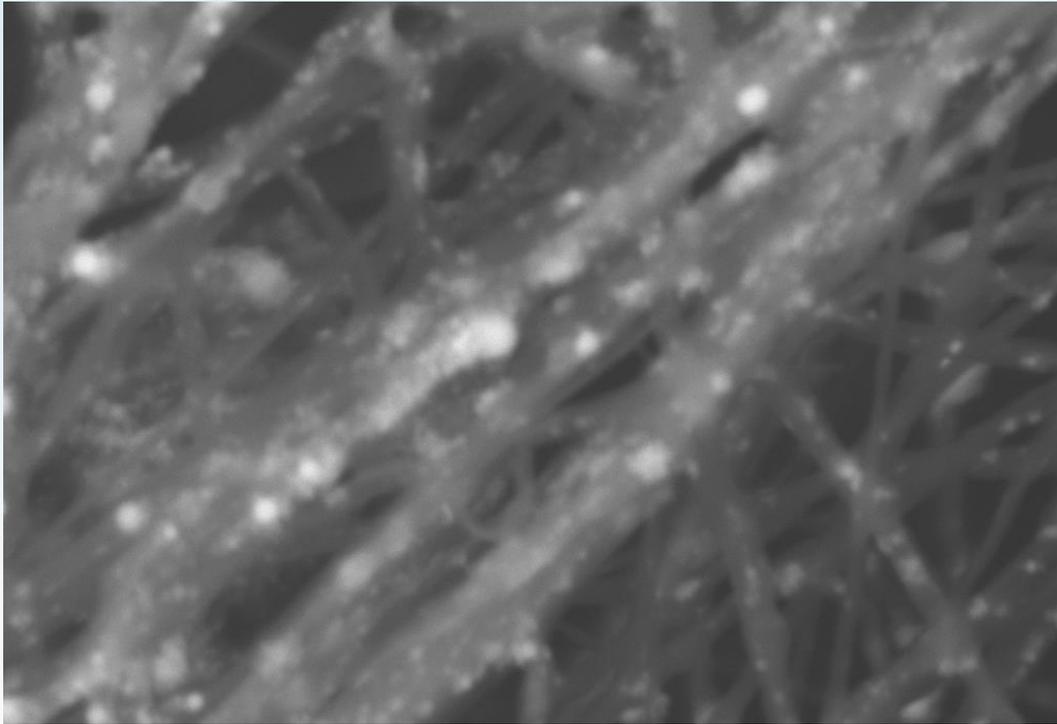


8µm

Electron Image 1

# PEO 15 wt.% + HA 30 wt.% at 36000 rpm and 0.3 MPa

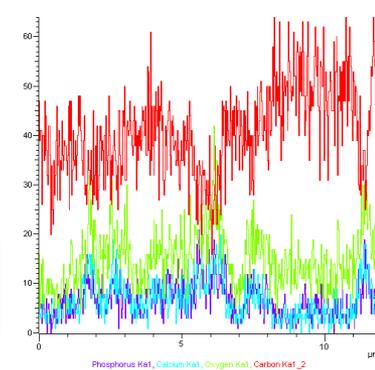
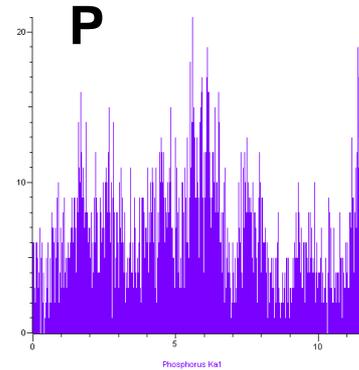
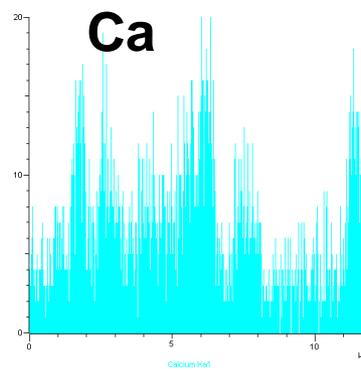
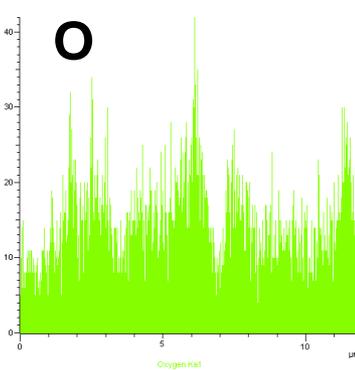
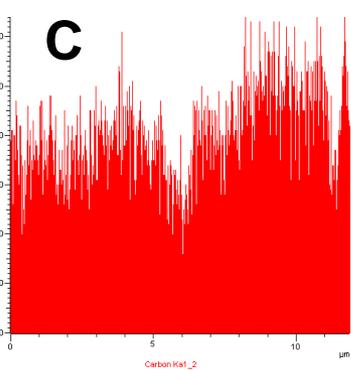
➤ EDS line scan along a fibre



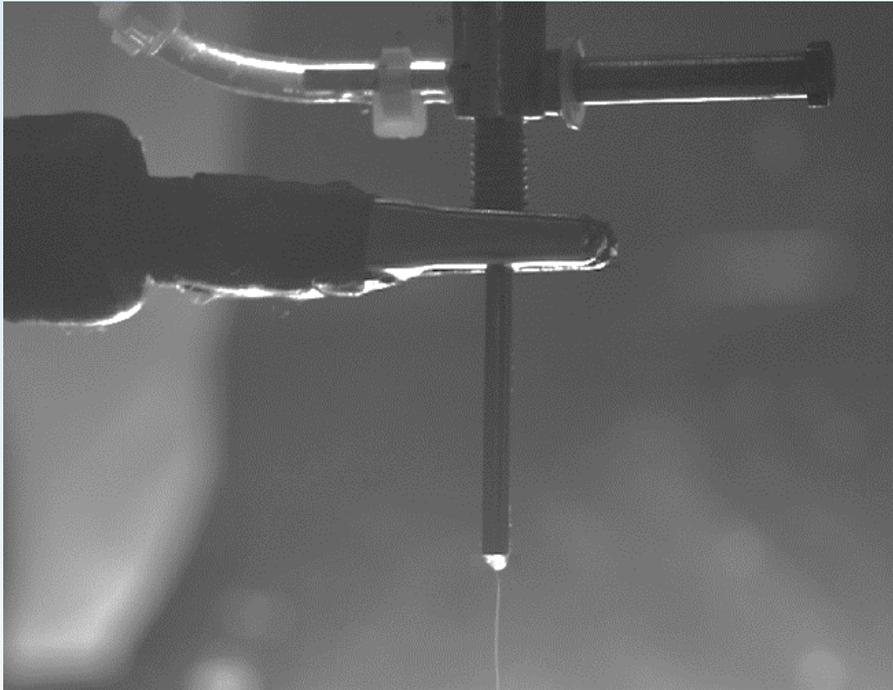
Electron Image 1

8µm

All



## Electrospinning



- Electric field overcomes surface tension of polymer solution which allows for a highly tuneable fibre production (voltage increase very incremental)
- Can create very fine fibres with wide range of biomedical applications
- Due to electric field, polymer solution must be conductive, which limits usage of various polymer systems

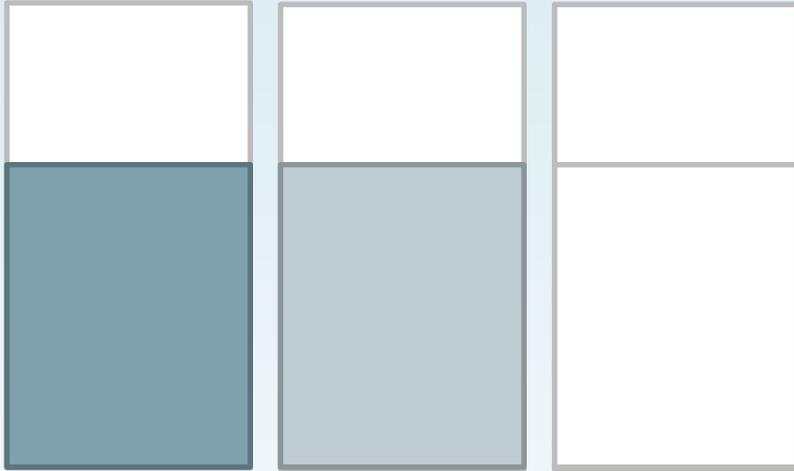
## Pressurised Gyration



- Centrifugal force and applied gas pressure overcome surface tension of solution which allow for a large range of materials to be used, in solution form or otherwise
- Very rapid fibre production rate with great potential for scale-up
- The full potential of this technology has not yet reached

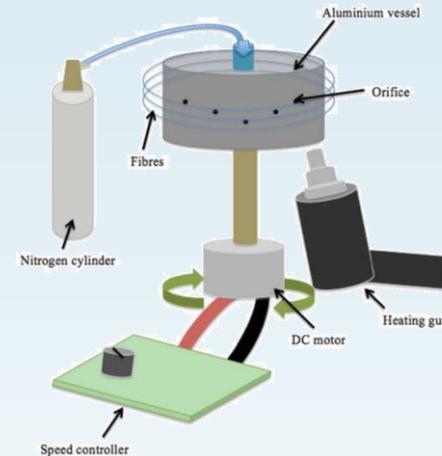
## Differences in Polymer Preparation

### Electrospinning



- Polymer solution must be prepared by dissolving a polymer within a solvent
- Polymer solution must be conductive in order to form fibres
- Solution must be homogenous otherwise it can effect the fibre formation
- Non-dissolved solutions can cause needle and pipe clogging and potentially damage setup

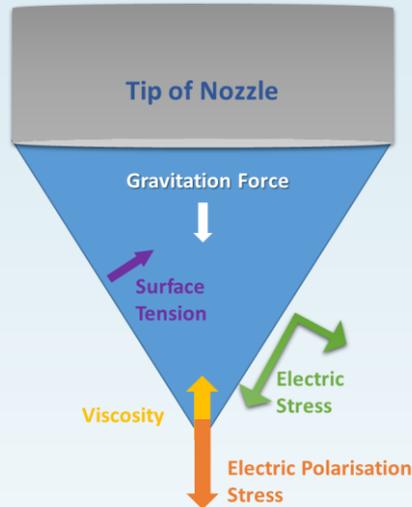
### Pressurised Gyration



- Polymer solution can be prepared just like with electrospinning
- Non-homogenous solution can be spun as vessel is large and creates mixing during spinning
- Polymer melts can also be spun using melt gyration
- Materials in vessel can be in different phases, but can still form fibres
- Thick and viscous solutions can be spun which can be a problem with EHD

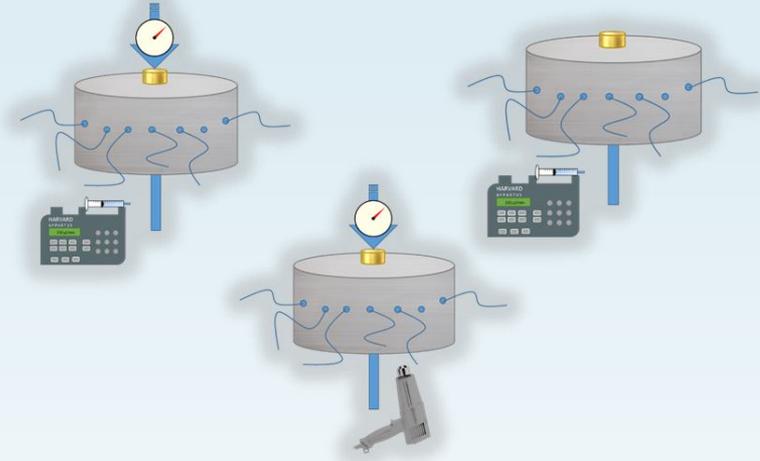
## Differences in Process Control

### Electrospinning



- Voltage can be adjusted very incrementally to give great control over final fibre morphology
- Flow rate is carefully governed to suit desired application and fibre size
- Collection distance can be altered to optimise fibre production by controlling fibre drying times
- Needle size can also have an effect on fibre production and morphology

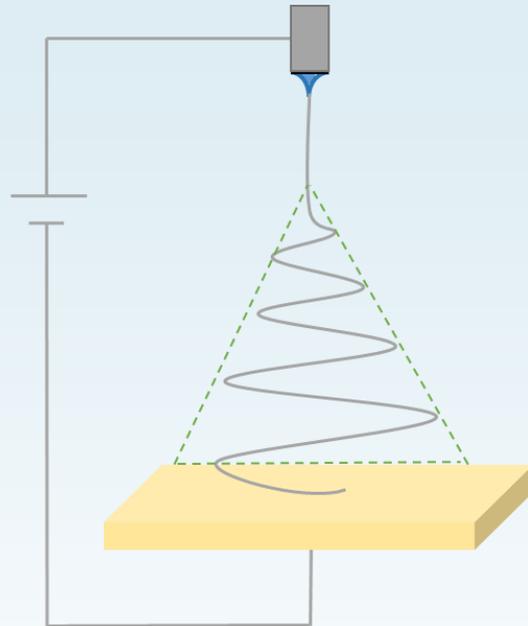
### Pressurised Gyration



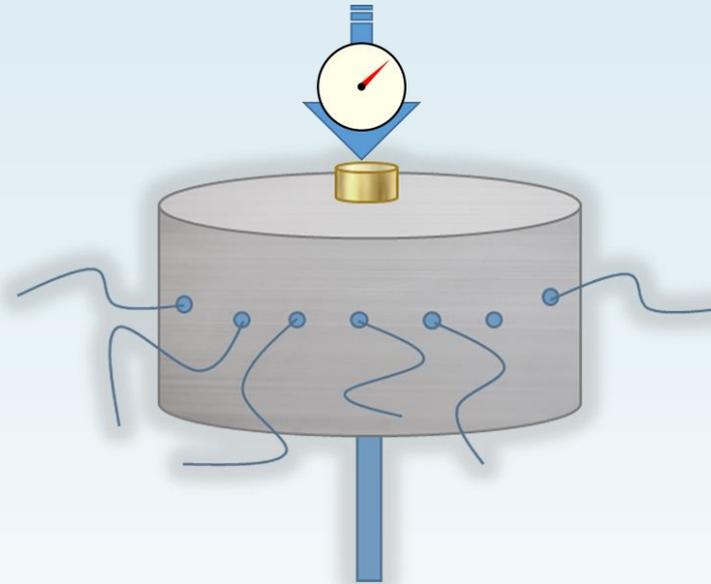
- Rotation speed influences centrifugal force and can be adjusted to control fibre morphology
- Applied gas pressure can also be increased to reduce fibre diameter, and control bead production
- Collection distances and setups do influence fibre structure
- Relative humidity can be used to regulate drying times and thus fibre morphology
- Melt temperature and infusion rate can further be used to regulate fibre production

## Differences in Fibre Production Rate

### Electrospinning



### Pressurised Gyration

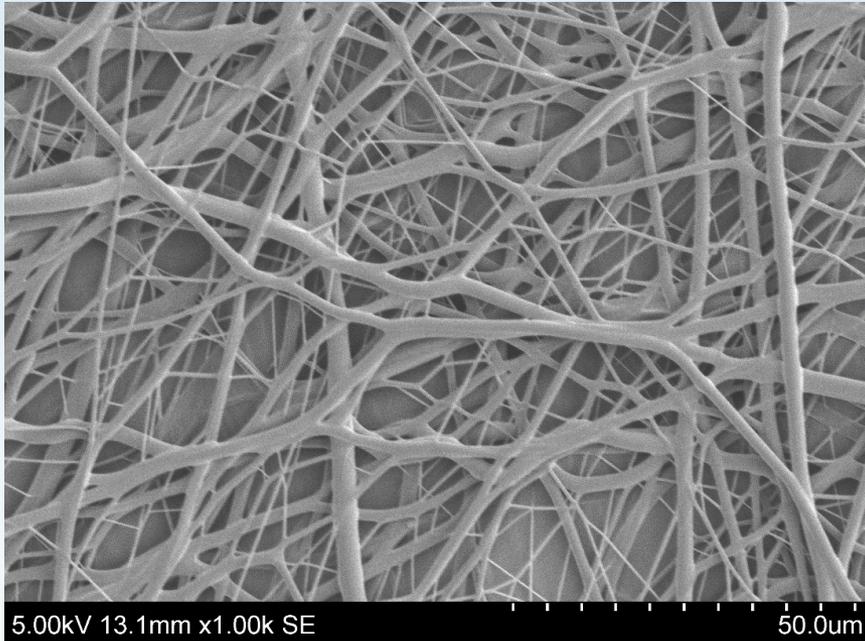


- Typical flow rates are 50-150  $\mu\text{L}/\text{min}$  in a single needle setup
- Typical fibre production in 1 hour is approximately **0.6 grams** of fibre for normal single-needle nanofibre setups
- Can be scaled up with multiple needle set-ups and advanced needle and collector devices which minimise solution-related problems

- 5mL of polymer solution can be produced into fibres in 15 seconds
- Typical fibre production rate in 1 hour is approximately **120 grams**
- Infusion gyration can further increase production rate to exceed 6 kg/hour
- Just like electrospinning, multiple gyration pot setups can be created to increase production rate

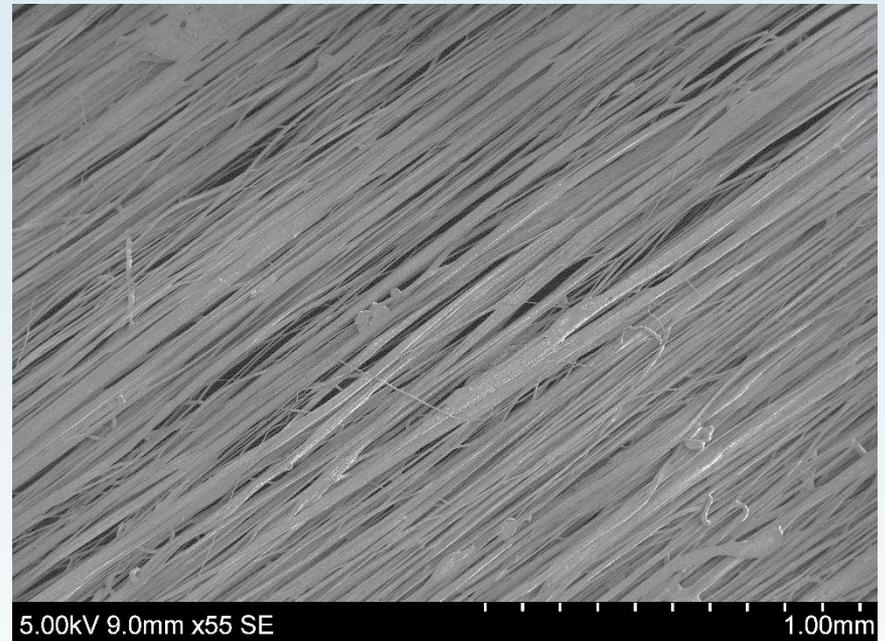
## Differences in Fibre Morphology

### Electrospinning



- Without a modified collector electrospun fibres form mesh-like fibres useful in mimicking the ECM
- Increased collection times allow for stacking up of layers to create 3D structures
- Very low fibre diameters can be achieved as low concentration polymers can be used

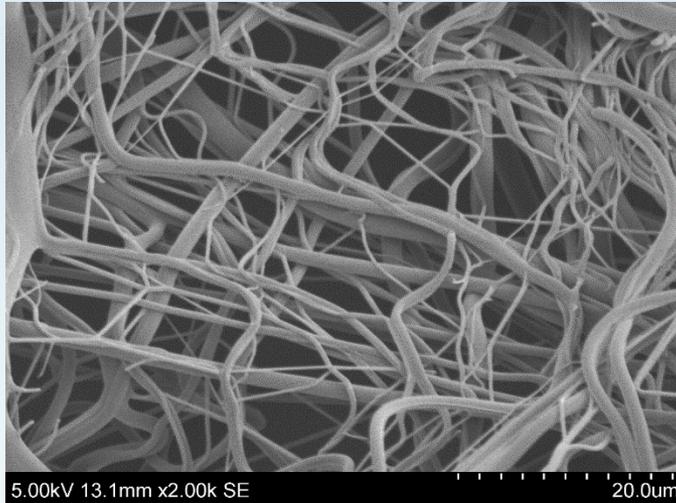
### Pressurised Gyration



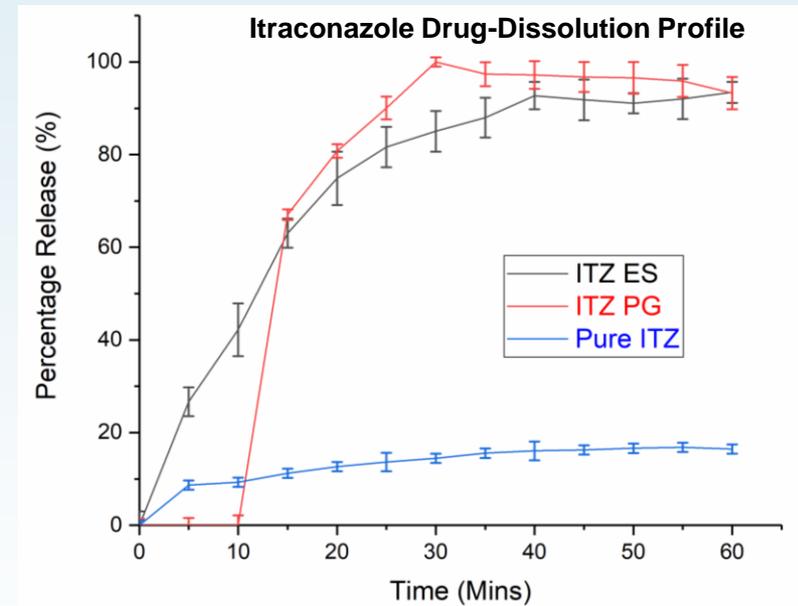
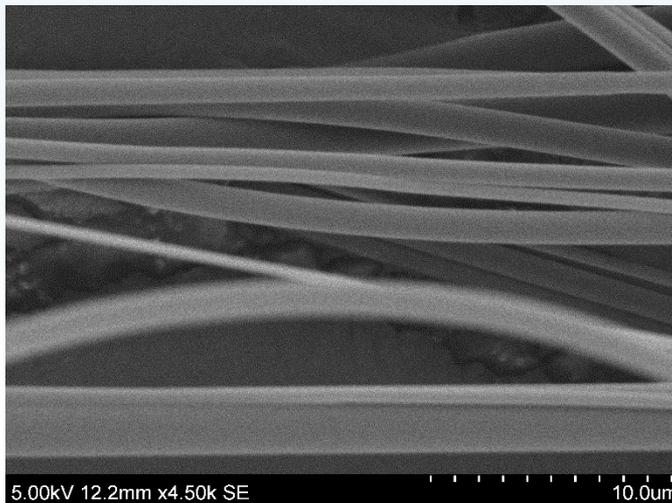
- PG fibres can easily form highly aligned structures suitable for use in neural tissue engineering
- The fibres can be collected into mats, bandages and other shapely materials and used directly as bandages without an additional step
- High production capable of fulfilling demand in medical scenarios

# Pressurised Gyration Vs. Electrospinning for Drug Delivery

**Electrospinning**  
 PVP-Itraconazole  
 Average Fibre  
 Diameter:  
 ( $0.94 \mu\text{m} \pm 0.34$ )



**Gyration**  
 PVP-Itraconazole  
 Average Fibre  
 Diameter:  
 ( $1.60 \mu\text{m} \pm 0.87$ )



## Differences in physical applications

### Electrospinning



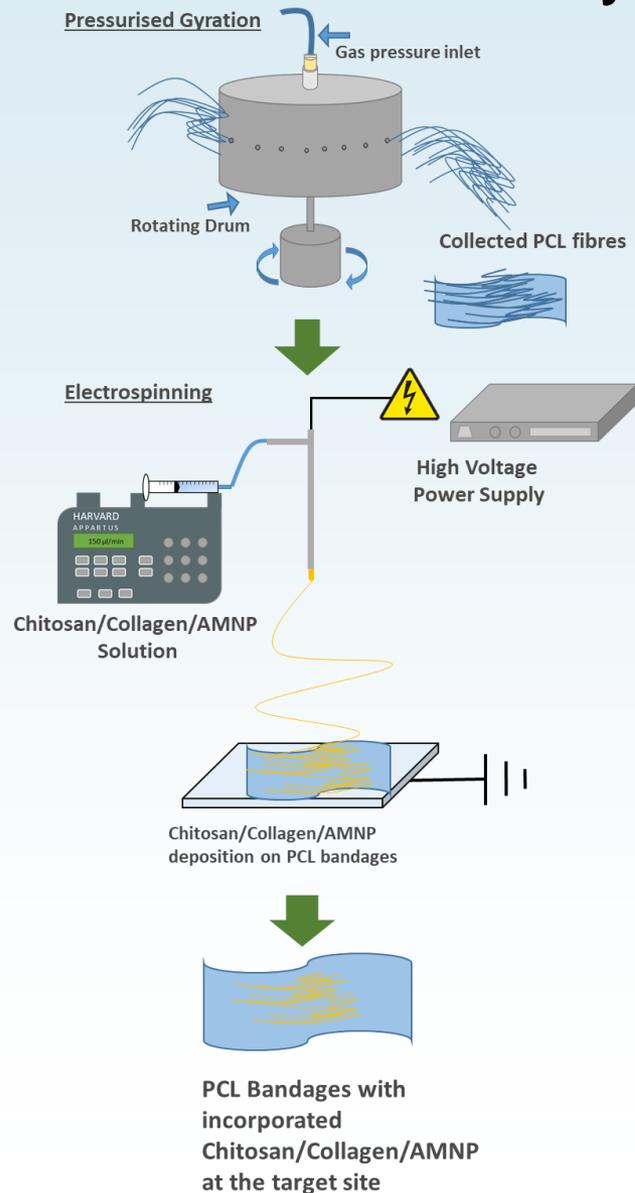
- Can be used to create filters with small mesh sizes that are capable of trapping microbes
- Portable EHD gun can be used to coat a layer of protective fibres directly onto the wound site in an emergency state

### Pressurised Gyration

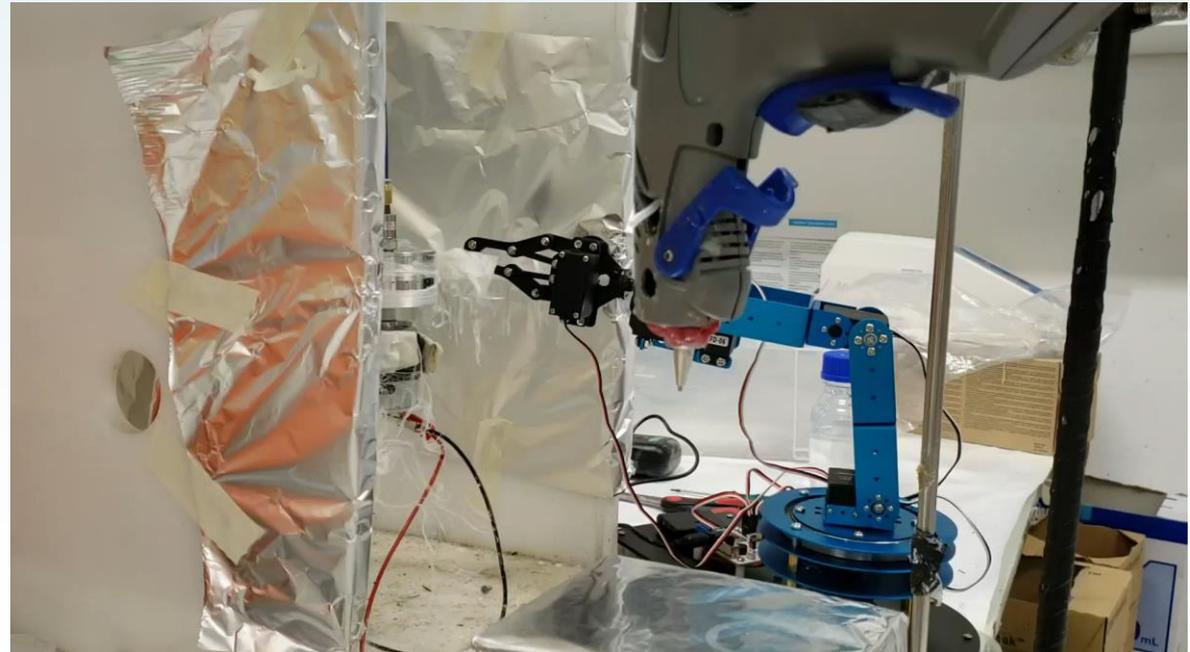


- Rapid spinning process allow for macro-scaled materials to be produced fast allowing for use in more physical applications such as wound dressings
- As the spinning material does not have to be in a solution, a wide range of antimicrobial and pro wound-healing additives can be incorporated into the bandages

# Combination of Gyration and Electro-spinning/spraying



- PCL high-yield fibres produced by gyration which can be collected as bandage-like sheets
- Fibrous ring is cut into pieces and robot transfers bandage to the electrospinning site
- Electrospinning of active nano-fibrous patches made of materials such as collagen and chitosan can be accurately printed onto the bandage



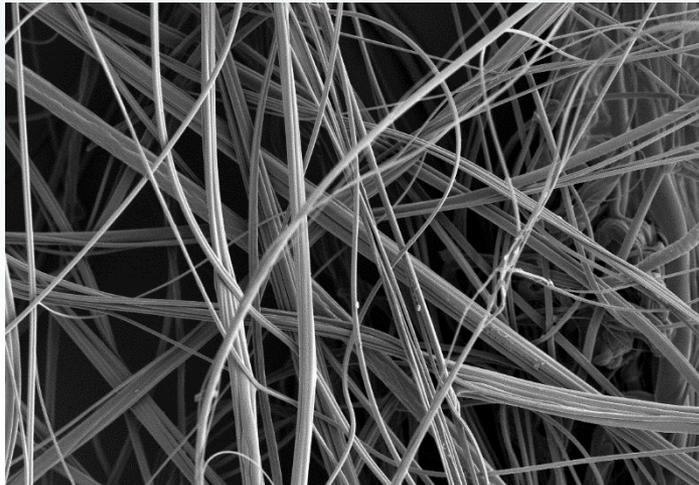
# Production of Bioactive Bandages & Patches



- PCL fibres produced via pressurised gyration which act as the bandage
- Collagen and Chitosan solutions are electrospun onto the bandages to provide a bioactive patch which both protects the wound site from bacterial infections and accelerates the wound healing process.

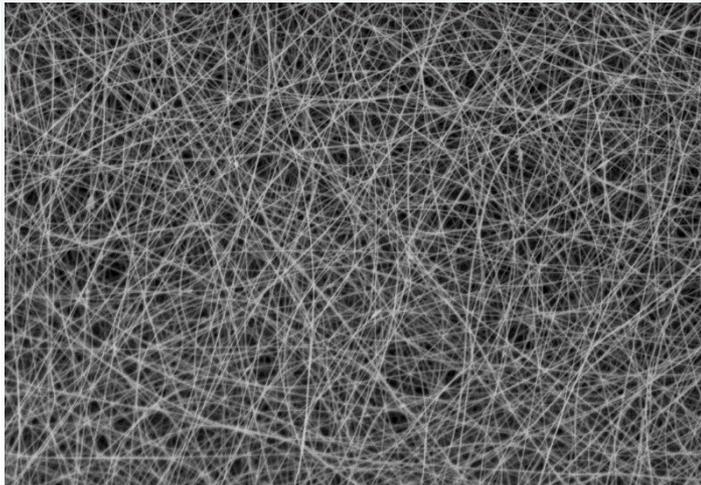


Bandage with patch  
on 90 mm Petri dish



5.00kV 10.0mm x320 SE

100um



5.00kV 9.2mm x1.30k SE

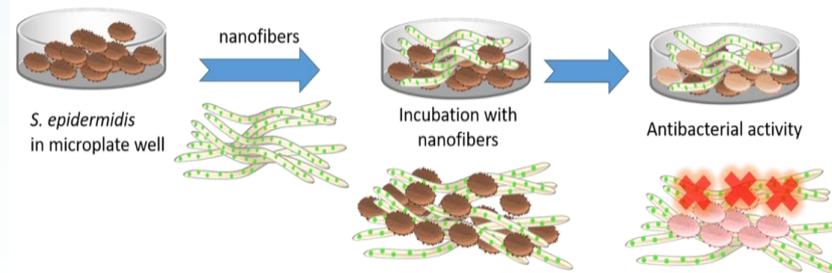
40.0um

**Gyration** ( $1.71 \pm 0.47 \mu\text{m}$ )

**Electrospinning** ( $142 \pm 31 \text{ nm}$ )

### Next generation wound dressing made of nanofibers incorporated with antimicrobial peptides (AMPs).

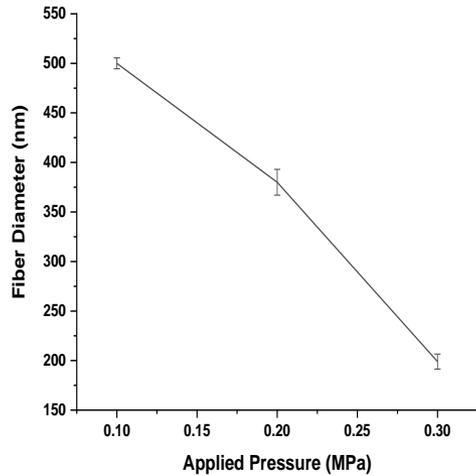
- Materials including hydrogels, decellularized dermal porcine dermal matrix and freezing dried or gas foaming scaffolds have been studied for wound healing applications, however, lack the ability to recapitulate the architecture of the skin's extracellular matrix (ECM)
- A new generation of wound dressing materials is anticipated to have a higher moisture level, and thereby provide sustained release results, which will enhance the healing of wounds due to the distinctive biological and non-sterile wound environment and to the difficult process of wound healing <sup>1-3</sup>.
- Pressurised gyration is selected in this research because of its ability to mass produce bandage like meshes with simple separation and enhanced process control <sup>4</sup>.
- A wound dressing material is created by loading peptides into water-soluble polymers that are proficient of efficient release. Thus, the growth of microorganisms is regulated by the antibacterial agents embedded in the structure of the fiber <sup>5</sup>.
- A series of PEO integrated AMP nanofibers are prepared using GH12-COOH-M2 and AMP2 antimicrobial peptides, which are synthesized by solid phase peptide synthesis (SPPS) using an AAPPTec Focus XC peptide synthesizer, supplied by Kansas University (USA).



**Schematic illustration of the antibacterial assessment of AMP integrated PEO nanofibers**

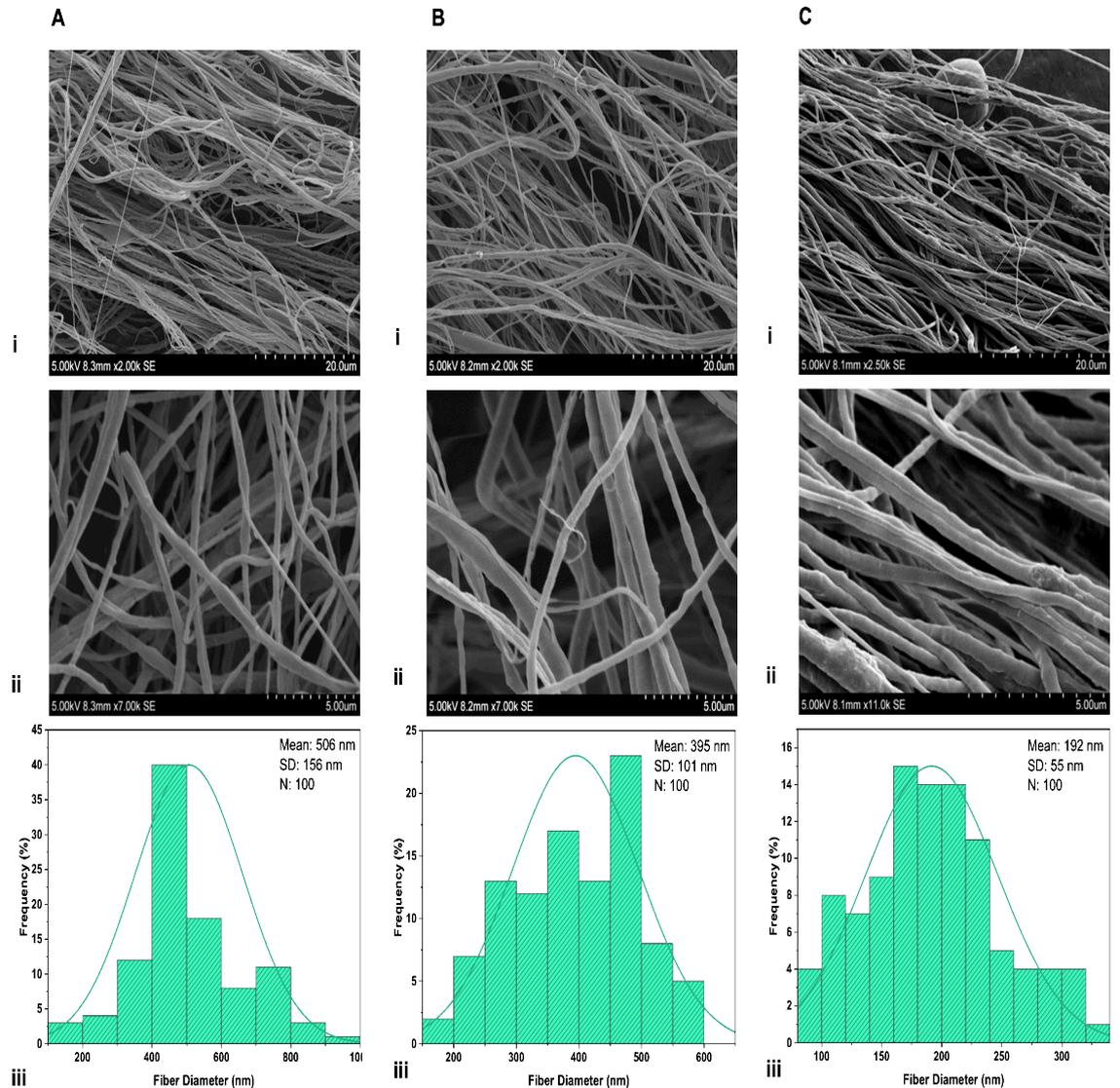
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3. Gizaw, M.; Thompson, J.; Faglie, A.; Lee, S.-Y.; Neunschwander, P.; Chou, S.-F., Electrospun Fibers as a Dressing Material for Drug and Biological Agent Delivery in Wound Healing Applications. 2018.
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5. Morais, D. S.; Guedes, R. M.; Lopes, M. A., Antimicrobial approaches for textiles: From research to market. 2016.

# Preliminary testing



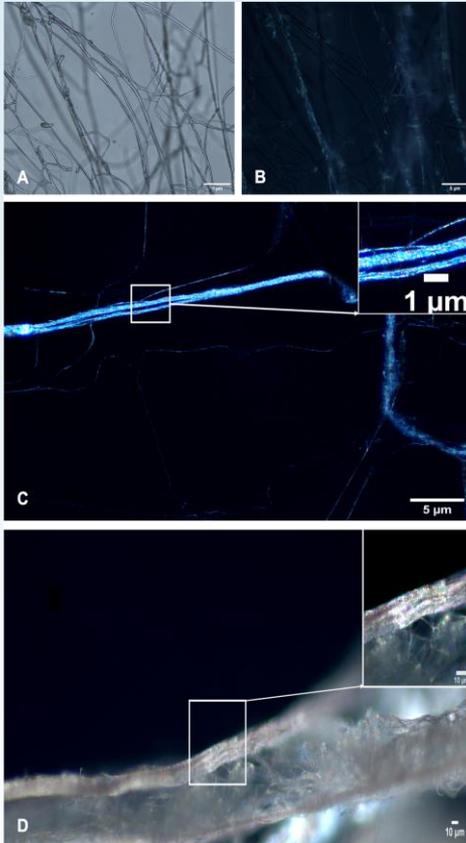
Average fiber diameters using M2 peptides at 35 µg/ mL at 0.1 – 0.3 MPa

- Average fiber diameter at 0.1 MPa – 0.3 MPa: 506 nm ± 155, 395 nm ± 101 nm and 191 nm ± 55 nm respectively.



Scanning electron microscopy (SEM) images and size distribution graphs at 15% PEO/water using M2 35 µg/ mL, a) 0.1 MPa, b) 0.2 MPa, c) 0.3 MPa

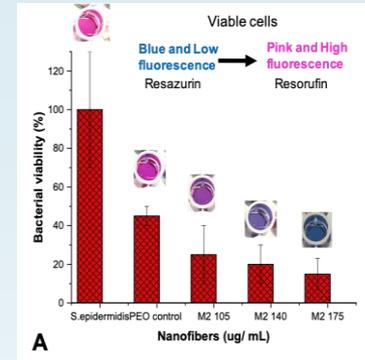
## AMP characterisation using fluorescence microscopy



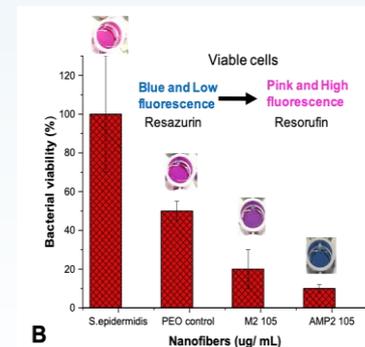
- Fluorescence microscopy was used to produce brightfield and polarisation contrast images
- Brightfield image A, and polarisation image B both serve as PEO control fibers. Images display fibers without attachment, this confirms samples are pure PEO fibers only
- Both polarisation images C and D display attachment of peptides (M2 and AMPs at 105 μg/mL) dispersed evenly inside and along fiber surface, illustrated as dots
- AMPs are embedded in the polymer matrix and fiber surface.

Polarisation contrast images captured on a Zeiss Axioplan2 microscope, with a 20xNA0.5 objective, a) PEO control without polarisation contrast, b) with polarisation contrast, c) PEO incorporated M2 Peptide, and d) PEO incorporated AMP2 polarization image

## Bacterial viability



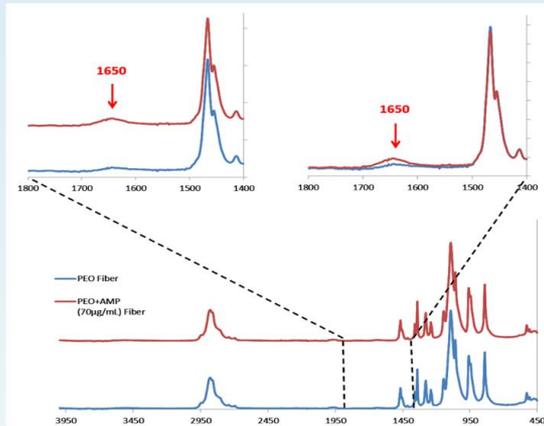
**A** Antimicrobial effect of PEO fibers loaded with M2 peptides (105 μg/mL – 175 μg/mL) against *S. epidermidis* evaluated with AlamarBlue cell viability assay



**B** Comparison of antimicrobial effect of PEO fibers loaded with M2 and AMP2 at 105 μg/mL against *S. epidermidis*

- Bacterial viability was assessed at various concentrations using only M2 integrated PEO nanofibers
- An increase in the M2 content has produced greater antimicrobial activity.
- Comparison of bacterial viability at 105 ug/mL using M2 and AMP2 suggest that antimicrobial activity was significantly higher using AMP2, indicating a greater release rate when tested against *S. epidermidis*.

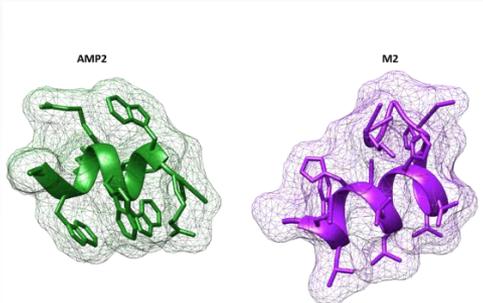
## Fourier-transform infrared spectroscopy



FTIR spectra of the nanofiber samples

- FTIR depicts that characteristic peaks of PEO observed at  $2900\text{ cm}^{-1}$  (methylene group CH<sub>2</sub> molecular stretching), and at  $1100\text{ cm}^{-1}$  and  $960\text{ cm}^{-1}$  (C O C group stretching)
- FTIR results showed that there was no change in chemical structure as a result of M2 integration to fibers
- FTIR of M2 samples integrated with PEO in two different concentrations did not reveal new chemical bonds
- Therefore, FTIR peaks of PEO fiber and fiber samples containing M2 are similar.

## Antimicrobial peptide structures

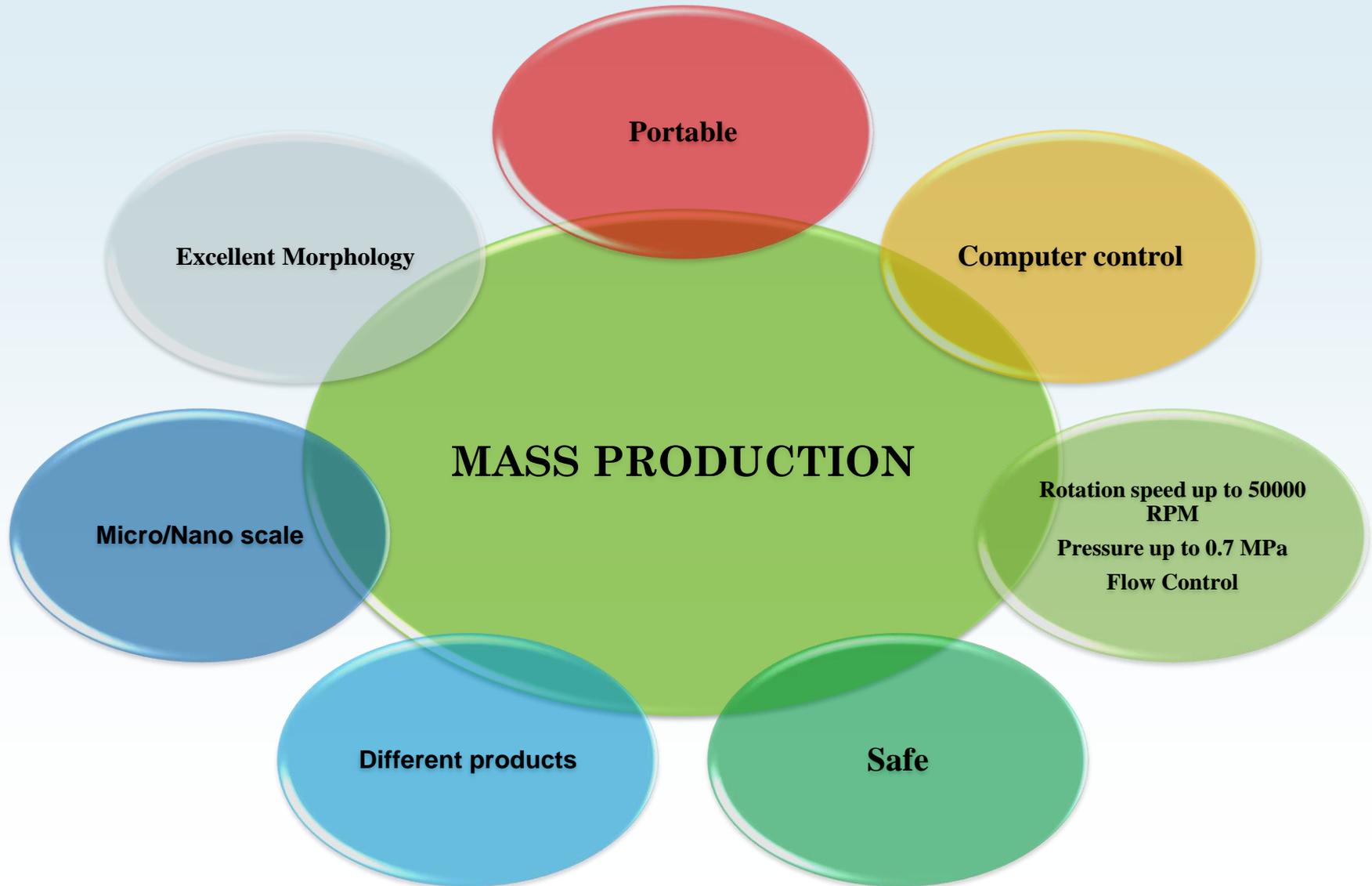


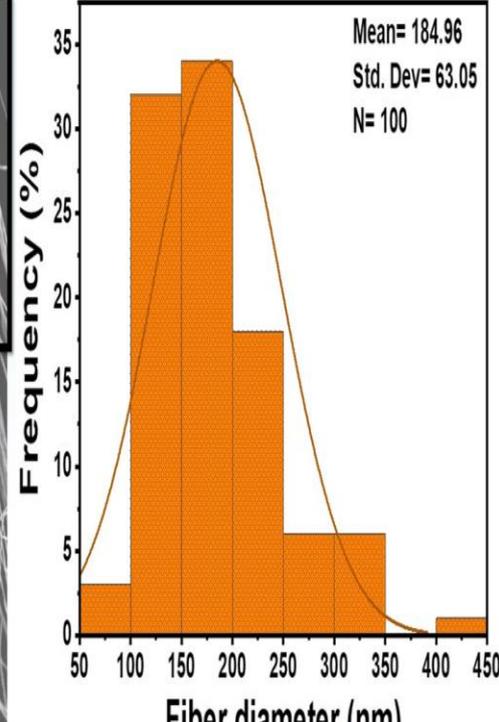
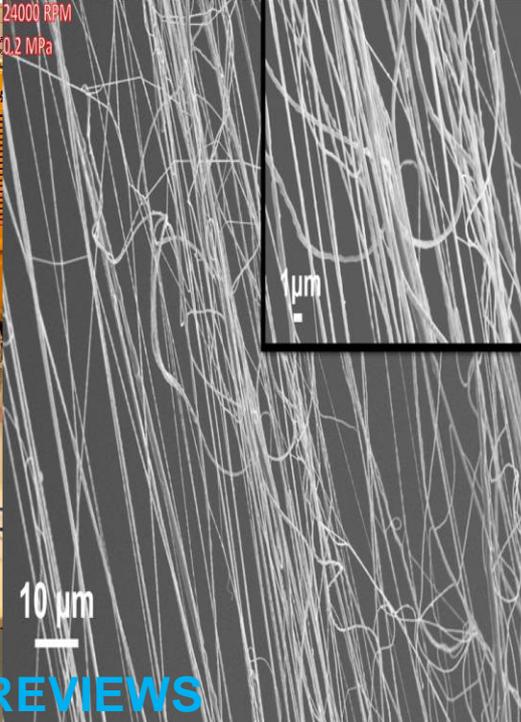
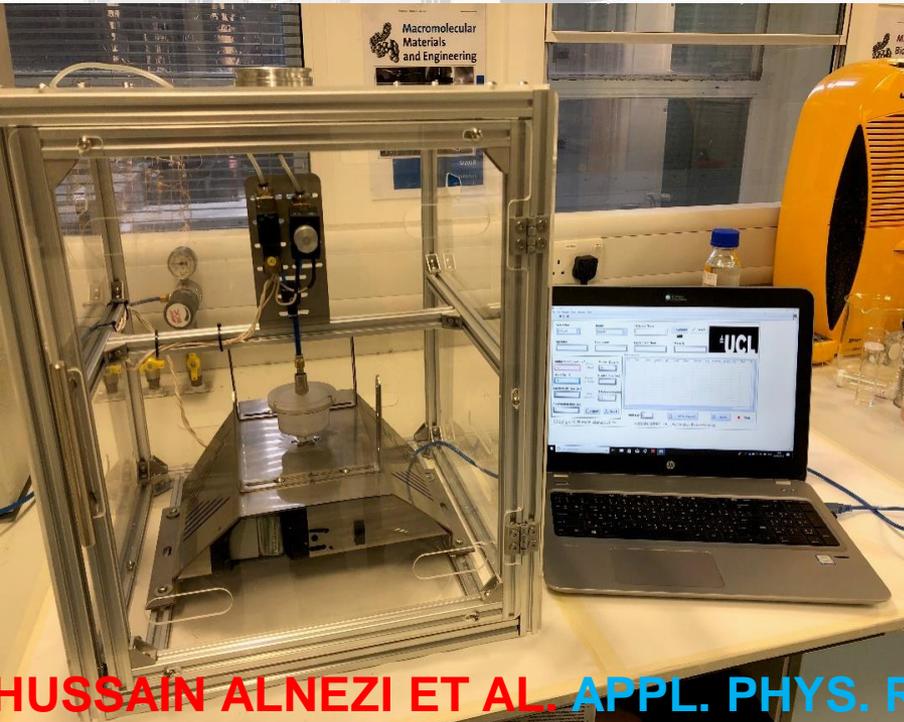
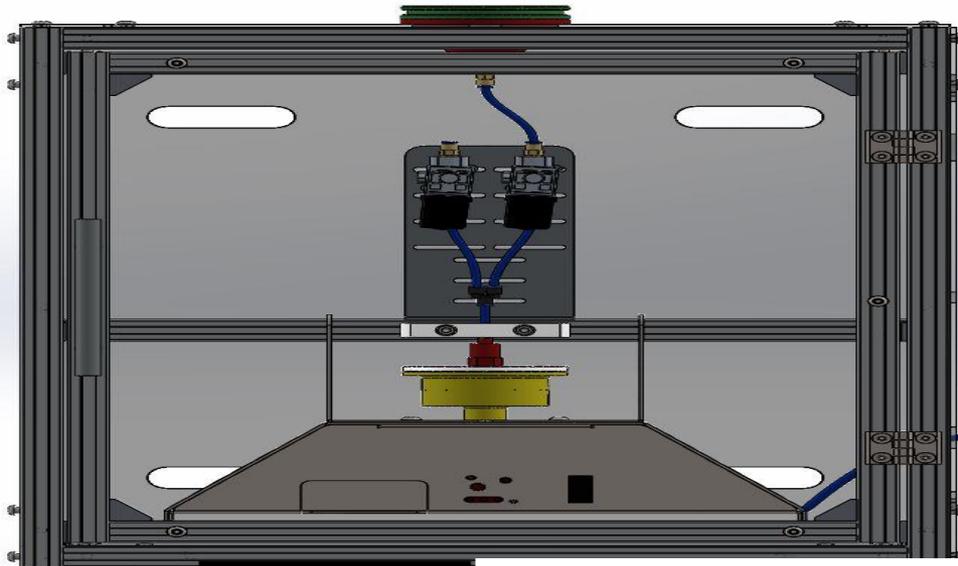
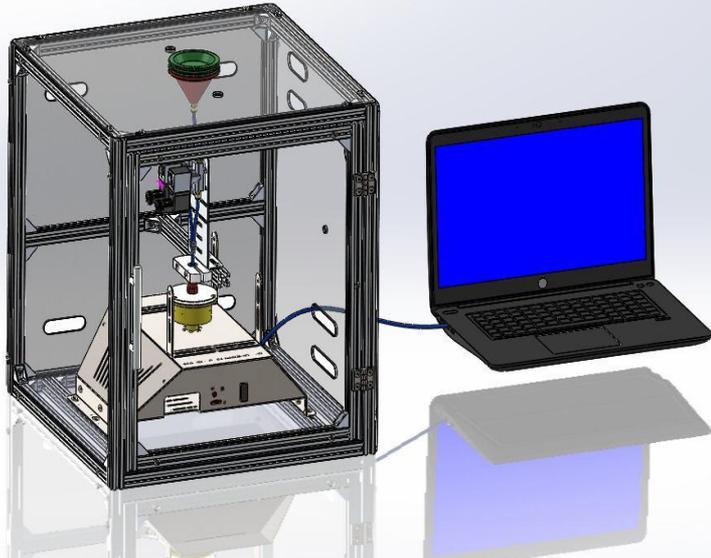
Secondary structure models for AMP2 and M2 peptides

- Secondary structure models of the peptides were obtained from the amino acid sequences using PEP-FOLD 3.5.8
- Chimeric peptides with AMP2 incorporated demonstrated effective antibacterial function against *S. epidermidis* bacteria.

## Conclusions

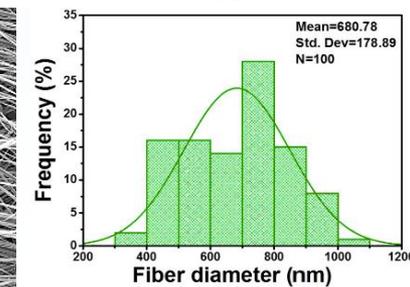
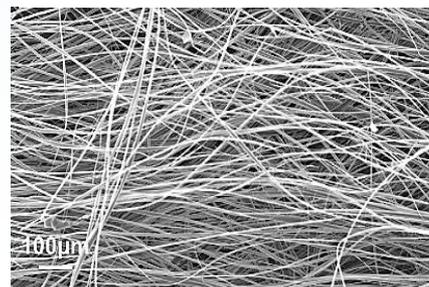
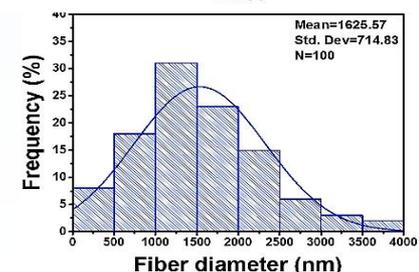
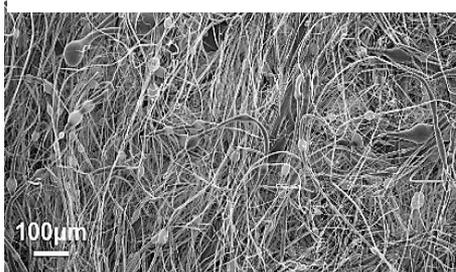
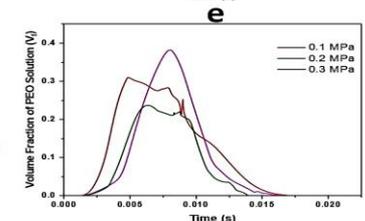
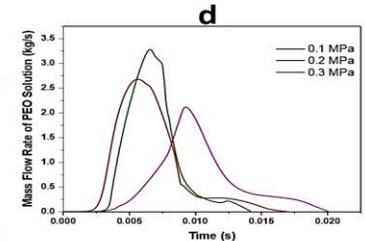
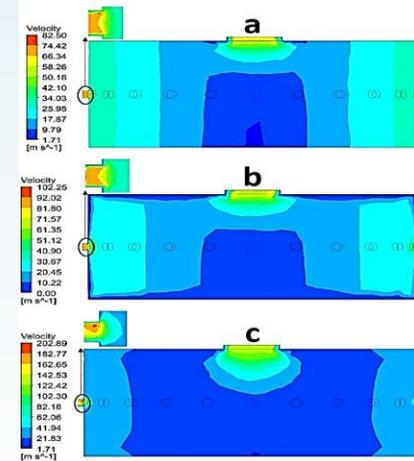
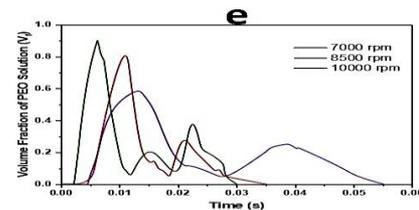
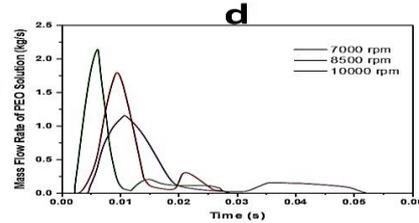
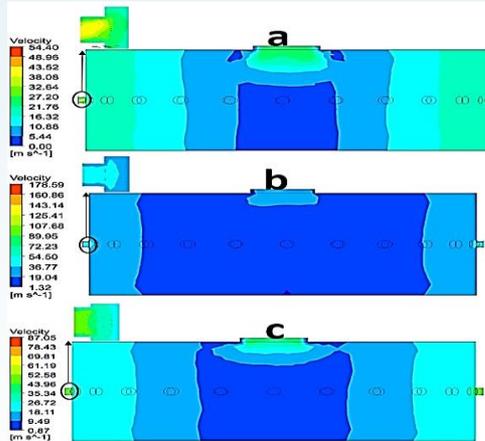
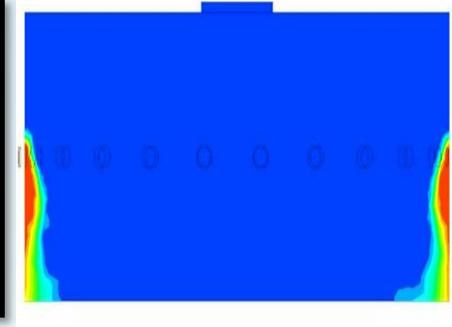
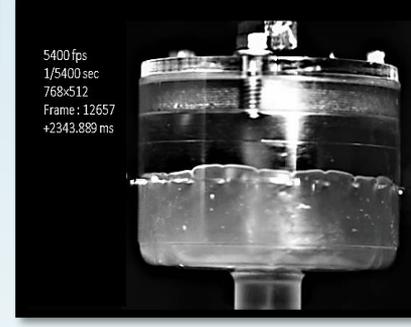
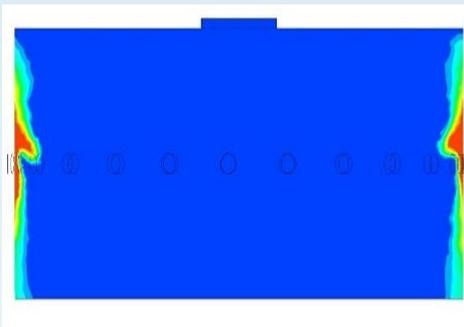
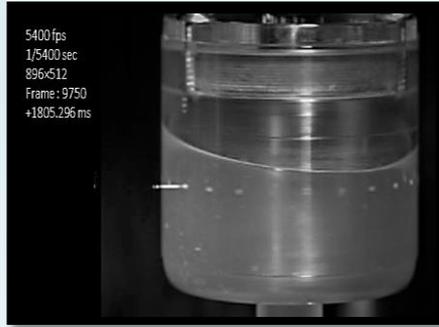
- The novel PEO nanofibers incorporated with M2 and AMP2 peptides generated an effective antibacterial activity against *S. epidermidis* to enhance wound healing applications
- The nanofibers created a high efficiency against *S. epidermidis* which is a common bacteria associated with wounds
- AlamarBlue assay confirms that AMP2 ( $105\text{ }\mu\text{g/ mL}$ ) performed the greatest antibacterial activity
- Therefore, loading antibacterial peptides with water soluble polymers such as PEO is an effective delivery method for the easy release of antimicrobial agents into a wound site
- Using water-based systems also allows to be more environmentally friendly whilst also ensuring maximum biocompatibility in an open wound scenario.





# Experimental and theoretical investigation of the fluid behavior during polymeric fiber formation with and without pressure

***C.S at 7000, 8500 and 10000 rpm no pressure*** **VS** ***P.G at 10000 rpm at 0.1, 0.2 and 0.3 MPa***





# Groundbreaking Research on Polymeric Fibers

## Creating Miracles with Polymeric Fibers

Amer Inst Phys News 15<sup>th</sup> October 2019

<https://publishing.aip.org/publications/latest-content/creating-miracles-with-polymeric-fibers/>

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<https://phys.org/news/2019-10-fabrication-polymeric-fibers-advanced-health.html>

<https://www.sciencedaily.com/releases/2019/10/191015131616.htm>

<https://www.parallelstate.com/news/researchers-studied-the-fabrication-of-polymeric-fibers-for-use-in-advanced-health-care/175626>

<https://www.eurekalert.org/multimedia/pub/213920.php>

**Experimental and theoretical investigation of the fluid behavior during polymeric fiber formation with and without pressure**  
**Applied Physics Reviews 6, 041401 (2019); <https://doi.org/10.1063/1.5110965>**

**[Hussain Alenezi](#)<sup>1,2</sup>, [Muhammet Emin Cam](#)<sup>1,3,4</sup>, and [Mohan Edirisinghe](#)<sup>1,a)</sup>**



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