

Lab-on-chip platform for on-field analysis of Grapevine leafroll-associated virus 3

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Abstract:

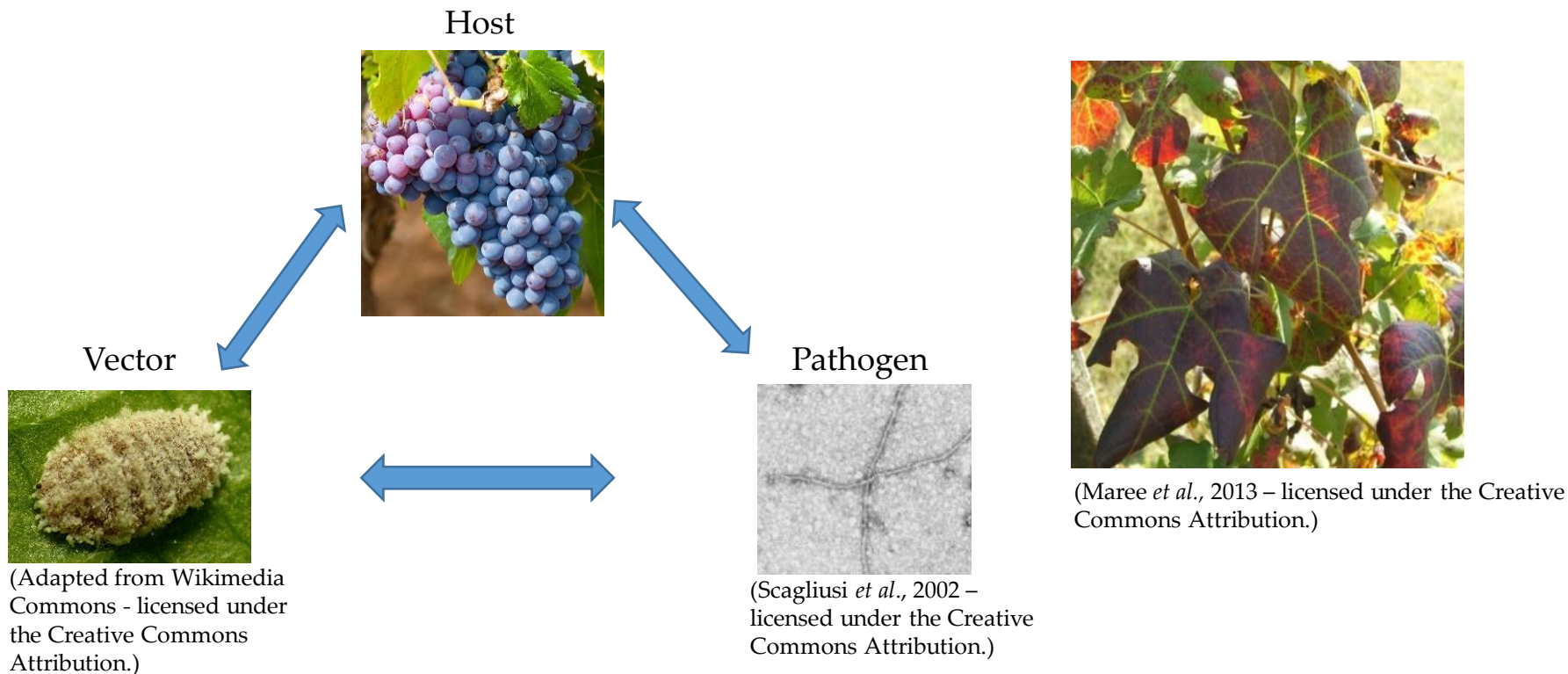
Human activities, especially the globalization of trade or tourism mass, had lead to he spreading of phytopathological adversities, all over the world.

These pathogens can have serious economic/environmental repercussions, due to the absence of therapeutic techniques and the need of rapid, in-field and low-cost detection methods.

Here we present a Lab-on-chip (LOC) platform, with electrochemical transduction method, recognizing serial dilutions of Grapevine leafroll-associated virus 3 (GLRaV-3).

LOC require small sample volumes, allowing a rapid detection of the target.

Keywords: Plant pathogens, environmental monitoring, sensors, lab-on-chip



Significant yield losses!

An estimated economic impact from \$ 25,000 to \$ 40,000 per hectare, in the absence of any control measure (Atallah *et al.*, 2012)



sensors



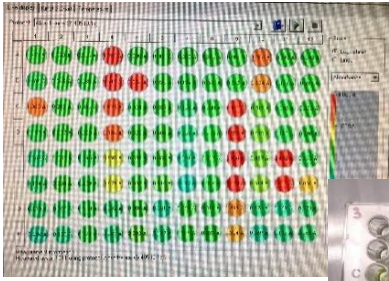
Review

Advances in Plant Disease Detection and Monitoring: From Traditional Assays to In-Field Diagnostics

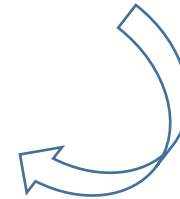
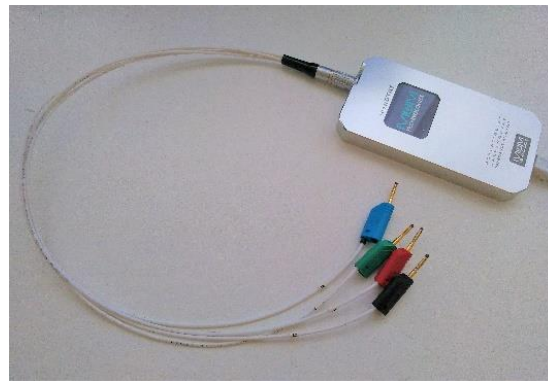
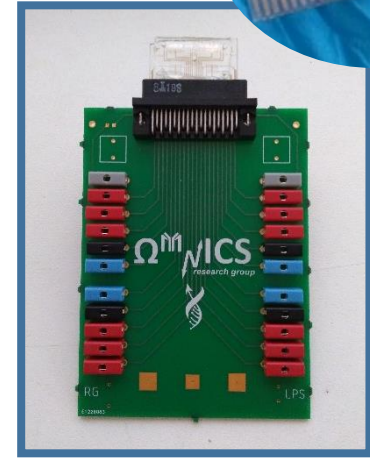
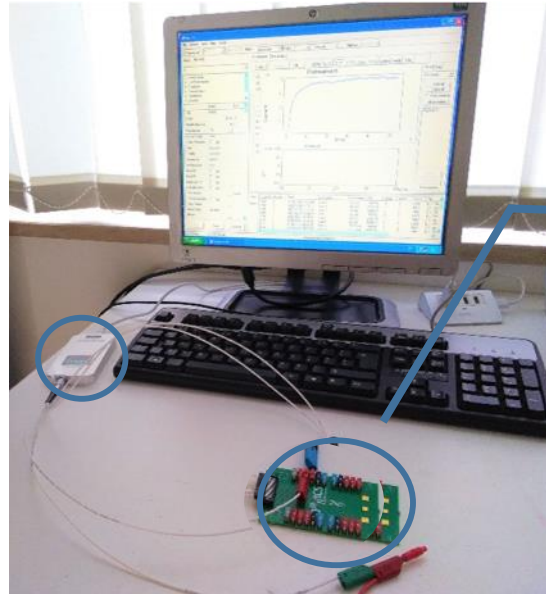
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ICMA
2021

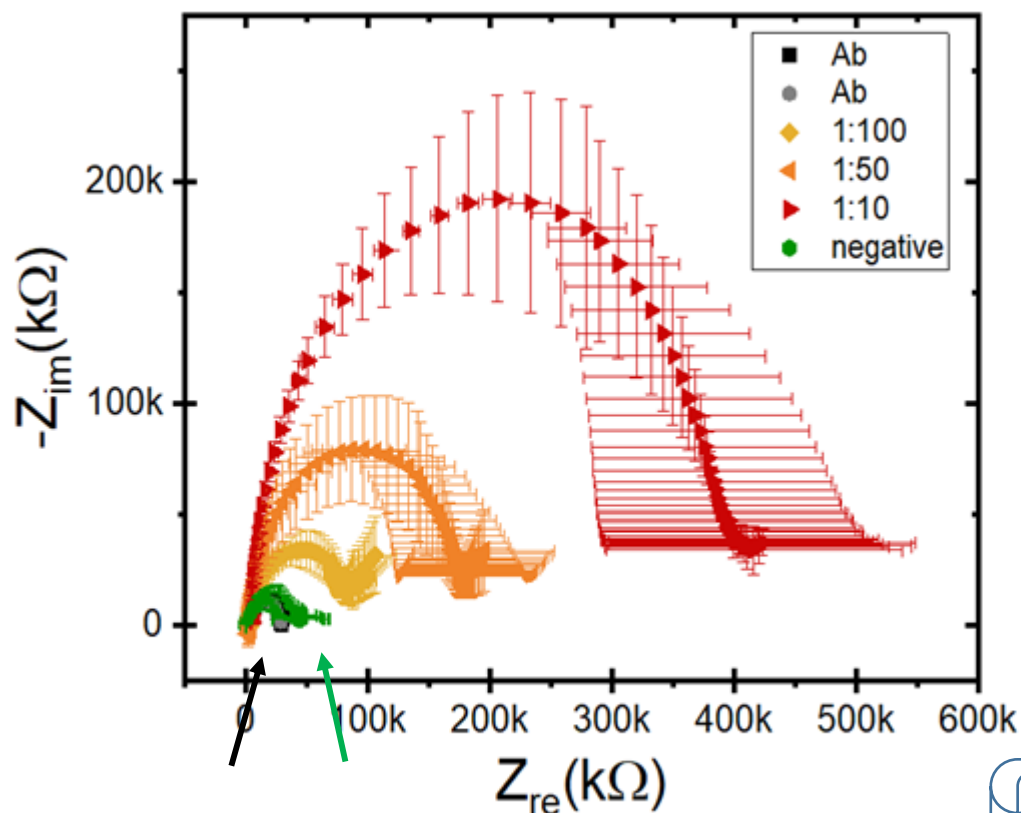
ELISA TEST



LOC



Results and discussion



The electron transfer resistance is about 40 k Ω after immobilization of the GLRaV-3 antibody and healthy sample.

ELISA test

Sample dilution	LOC	ELISA
1:3	+	+
1:5	+	+
1:10	+	-
1:20	+	-
1:50	+	-
1:100	+	-
1:1000	-	-

CURRENT DETECTION METHODS

Blouin, A. *et al.*, 2017;
Rowhani, A. *et al.*, 2017;
Bendel *et al.*, 2020.

Conclusions

- **LOC devices shows higher sensitivity compared to ELISA test.**
- **This device can be competitive with conventional diagnostic methods for costs and portability, making the difference in real time detection of the pathogens.**
- **Due to its ease of use, sensibility and specificity, is possible to extend its application, for the detection of other viruses.**



THANKS FOR
YOUR ATTENTION!

Supplementary Materials

Links:

- Maree, H. J.; Almeida, R. P. P.; Bester, R.; Chooi, K. M.; Cohen, D.; Dolja, V. V.; Fuchs, M. F.; Golino, D. A.; Jooste, A. E. C.; Martelli, G. P.; Naidu, R. A.; Rowhani, A.; Saldarelli, P.; Burger, J. Grapevine Leafroll-Associated Virus 3. *Front. Microbiol.* **2013**, *4*. <https://doi.org/10.3389/fmicb.2013.00082>.
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- Buja, I.; Sabella, E.; Monteduro, A. G.; Chiriaco, M. S.; De Bellis, L.; Luvisi, A.; Maruccio, G. Advances in Plant Disease Detection and Monitoring: From Traditional Assays to In-Field Diagnostics. *Sensors* **2021**, *21* (6), 2129. <https://doi.org/10.3390/s21062129>.

Supplementary Materials

Links:

- Blouin, A. G.; Chooi, K. M.; Cohen, D.; MacDiarmid, R. M. Serological Methods for the Detection of Major Grapevine Viruses. In *Grapevine Viruses: Molecular Biology, Diagnostics and Management*; Meng, B., Martelli, G. P., Golino, D. A., Fuchs, M., Eds.; Springer International Publishing: Cham, 2017; pp 409–429. https://doi.org/10.1007/978-3-319-57706-7_21.
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Acknowledgments

This work was supported by PON FSE – FESR 2014–2020 (CCI 2014IT16M2OP005) – Axis I “Investments in Human Capital” Action I.1 “Innovative PhDs with industrial characterization” – project DOT1712250 code 1 and by the Italian National FISR-CIPE Project “Inno-Sense”: Development of an innovative sensing platform for on-field analysis and monitoring (delibera CIPE n.78 del 07/08/2017).