

# Investigating the *structure-dynamics-function* relationship in antibodies

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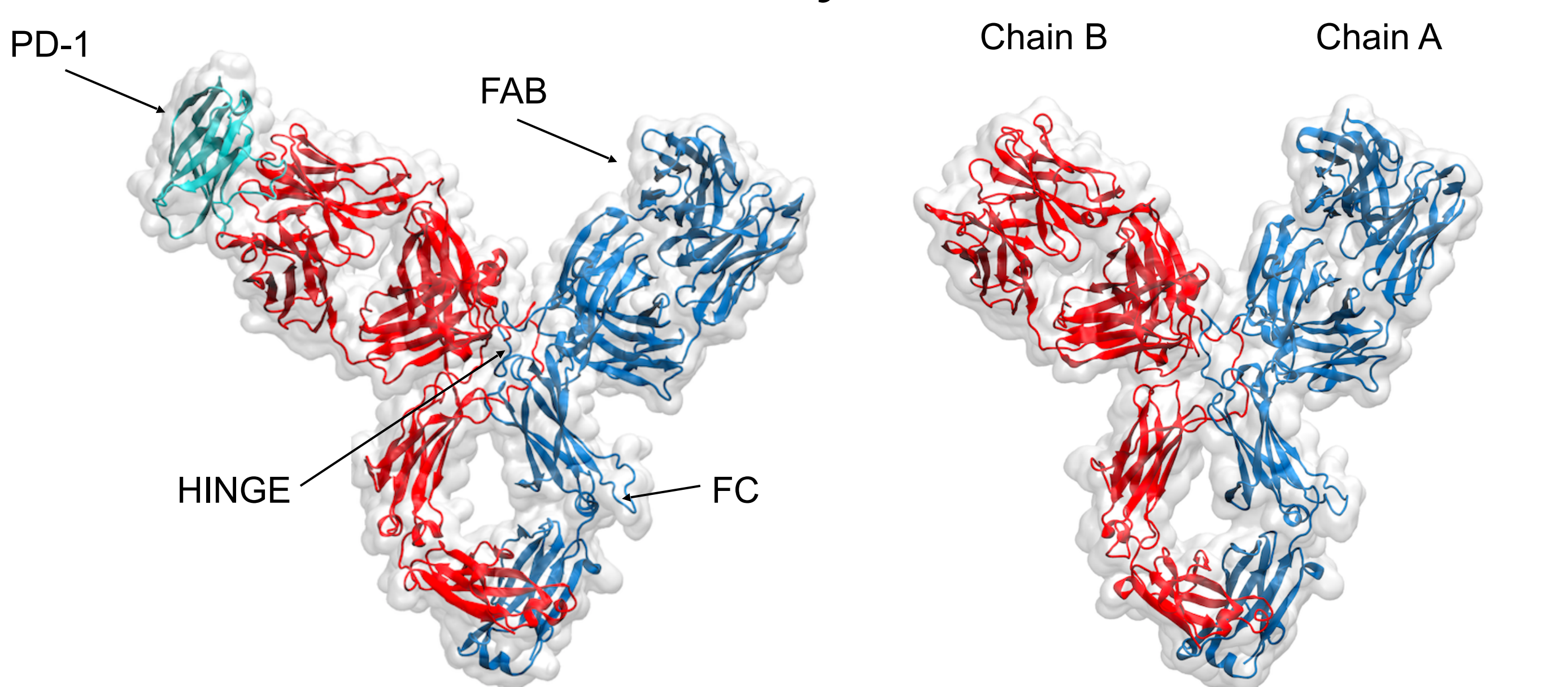
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## INTRODUCTION

Molecular Dynamics (MD) simulations can help to understand the **complex behaviours** observed in proteins. The **characterization of the large-scale dynamics** through MD and the **correlation between dynamical properties and structural features** is of fundamental importance for the **elucidation of biological mechanisms**. The all-atom dynamics also serves as a guide in the **identification of those structural patterns** whose preservation is necessary in the construction of simplified, that is **coarse-grained**, models. This study is meant as a starting point for the **application of multi-scale methods** to biologically relevant macromolecules.

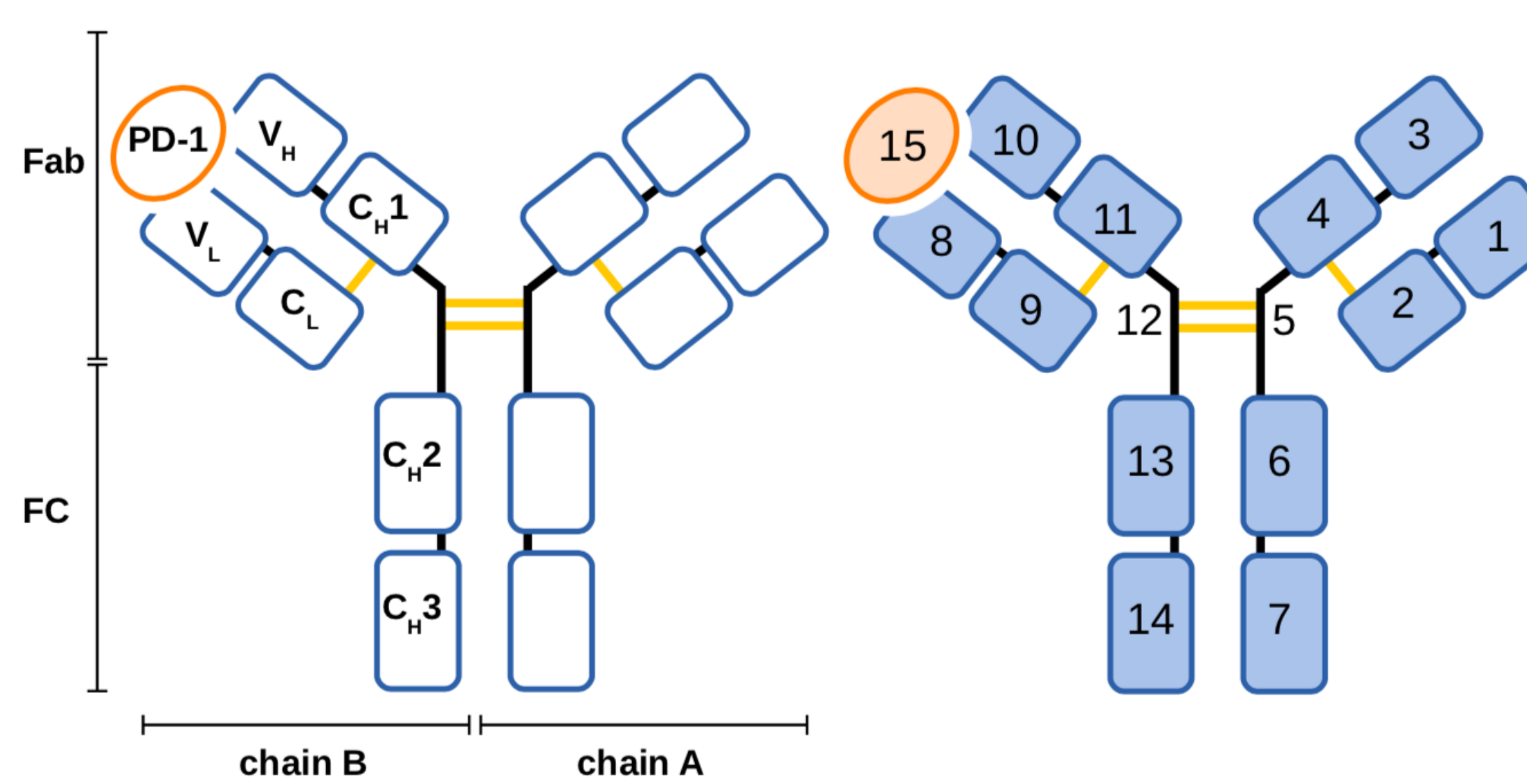
We simulate with plain MD (4 $\mu$ s) the therapeutic antibody Pembrolizumab [1], an immunoglobulin of the IgG4 family, in either **apo** or **holo** form. Here we compare show the results for both systems.

### The Antibody



**Figure 1** Ribbon representations of the therapeutic antibody Pembrolizumab and its antigen PD-1 (PDB code: 5DK3, 5GGS) and of the apo form.

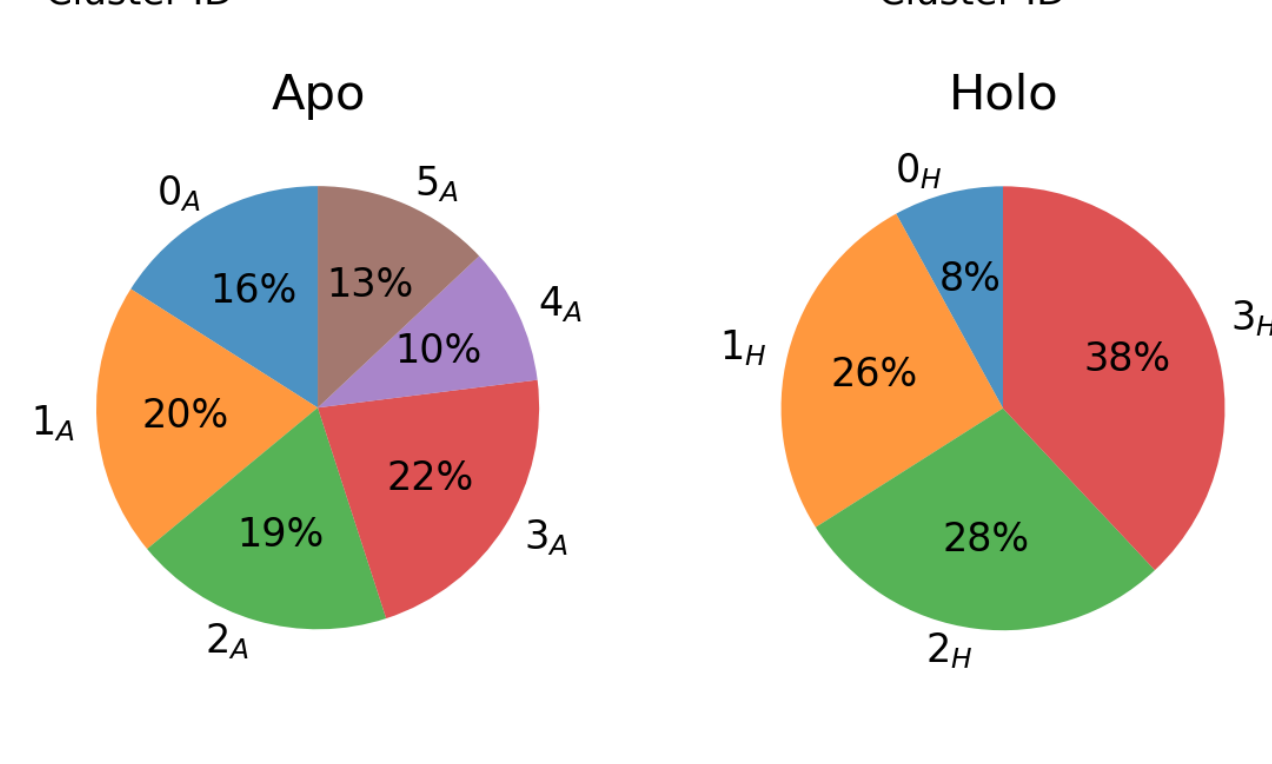
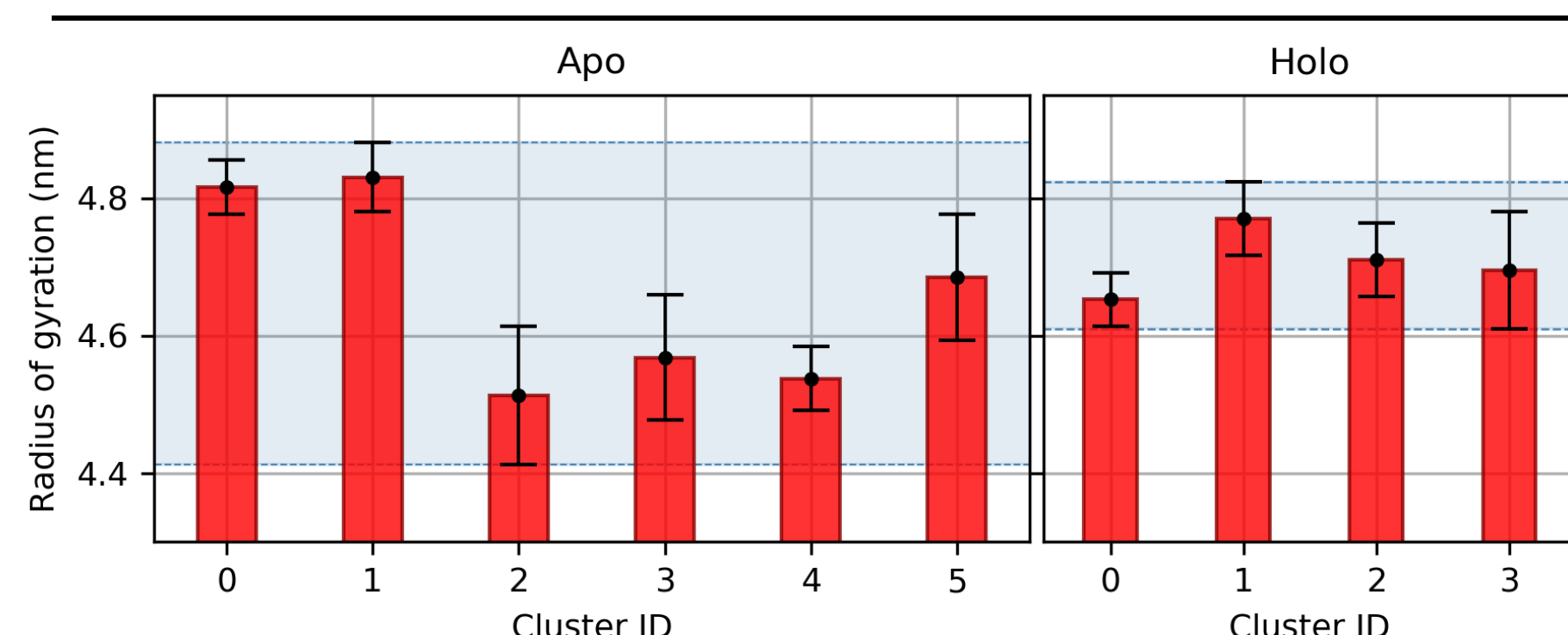
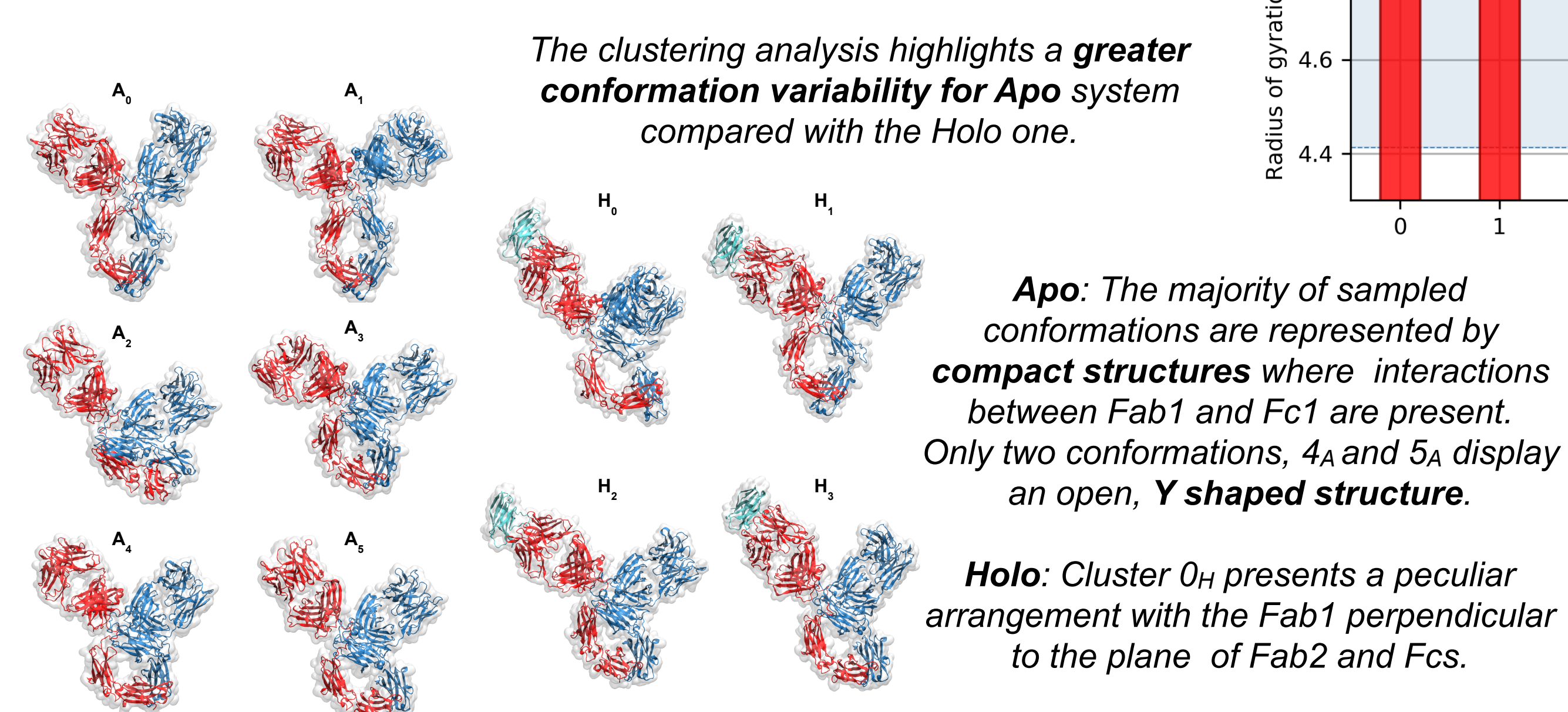
IgG antibodies are composed by **2 heavy** and **2 light** chains, each of them has constant domains and variable domains which represent the binding region of the antigens. The whole structure can be divided in **Fab**, **Fc** and **hinge** regions.



**Figure 2** Schematic representation of the antibody structure, in yellow the disulfide bonds are depicted.

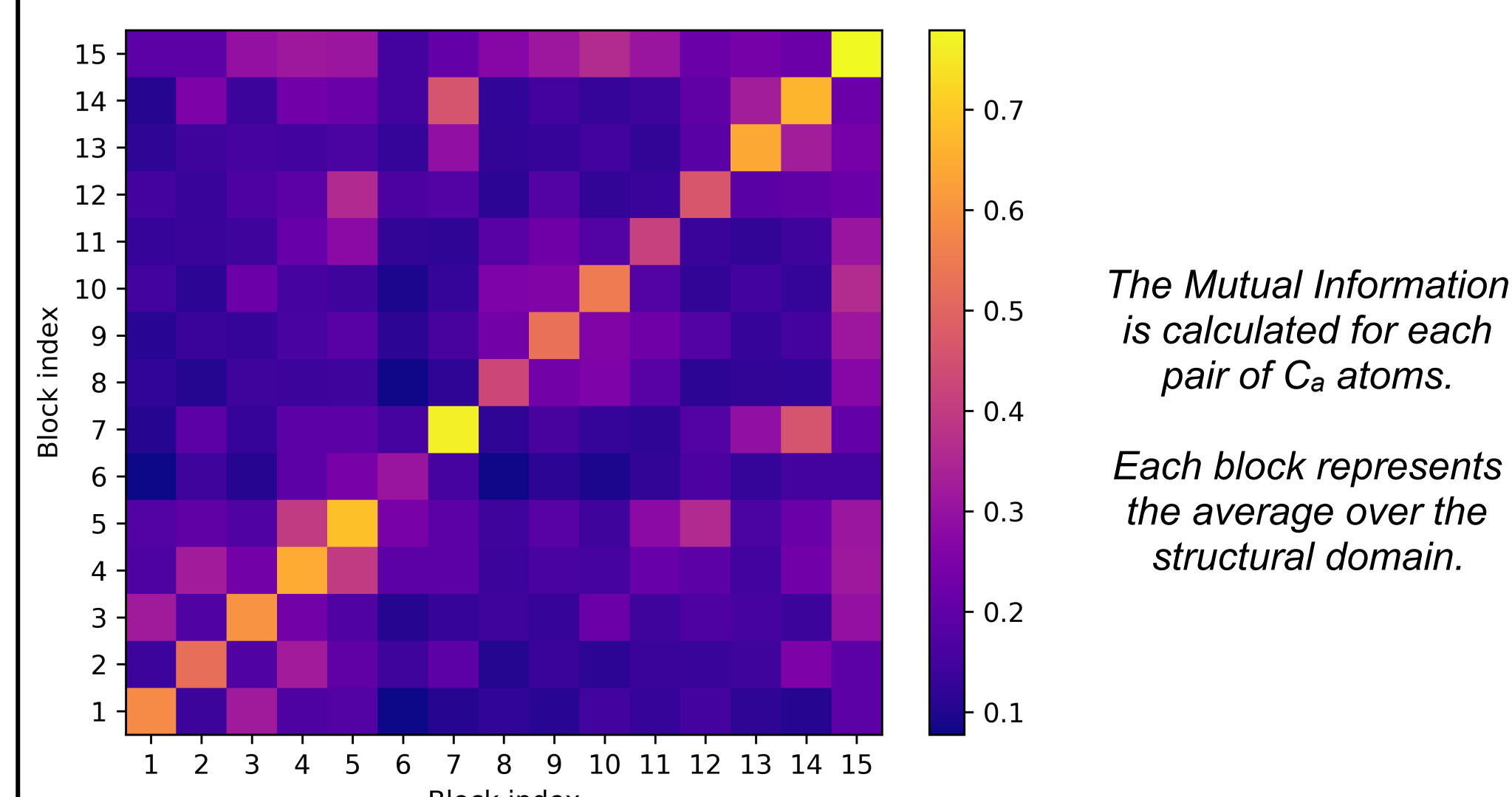
The role of monoclonal antibodies (**mAbs**) for therapeutic applications dramatically increased in recent years. mAbs have been developed to treat a large variety of conditions, including **cancer**, autoimmune diseases and, very recently, **COVID-19**. Engineering of the antibody sequence is habitually performed to **optimize its therapeutic efficacy**. MD is used as a valuable tool to assess the effects of these modifications on the antibody structure and to predict the **stability** and the **conformation variability** of the molecule.

### Apo and Holo forms display different conformation variability

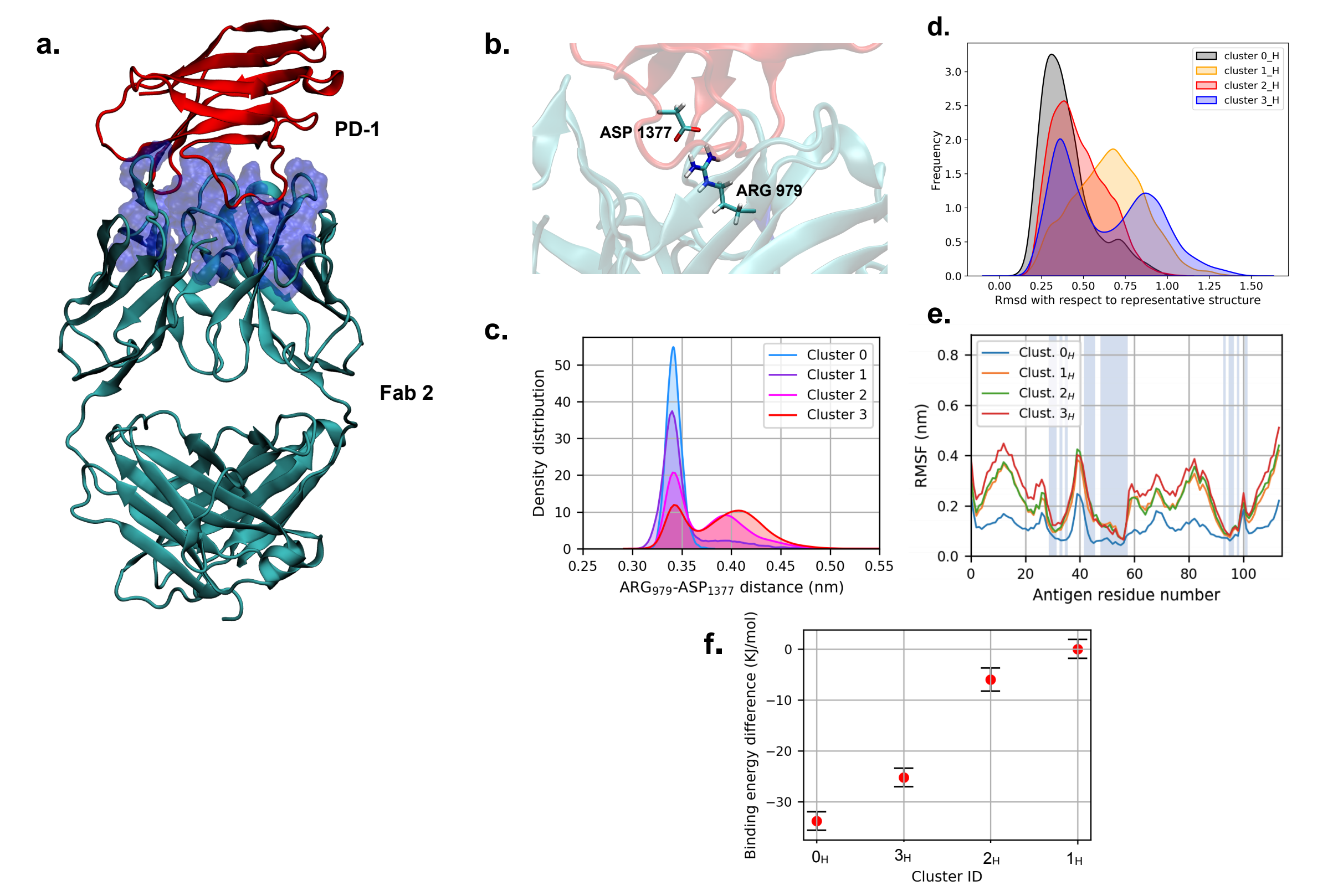


### Mutual Information

$$I_{i,j} = \iint p(x_i, x_j) \log \left( \frac{p(x_i, x_j)}{p(x_i)p(x_j)} \right) dx_i dx_j$$



### Binding Modes



The binding is strengthened when the antibody is in cluster 0<sub>H</sub> resulting in large and stable values of the contact surface area between pembrolizumab and PD-1 (a. b. c.)

The RMSF of the antigen molecule reflects the strength of the interactions, showing the lowest values for cluster 0<sub>H</sub> (e.)

The calculation of RMSD in each cluster, with respect to the respective representative structure reveals a sharper distribution in cluster 0<sub>H</sub> compared to the others, corroborating the hypothesis that the latter is the most stable conformation (d.)

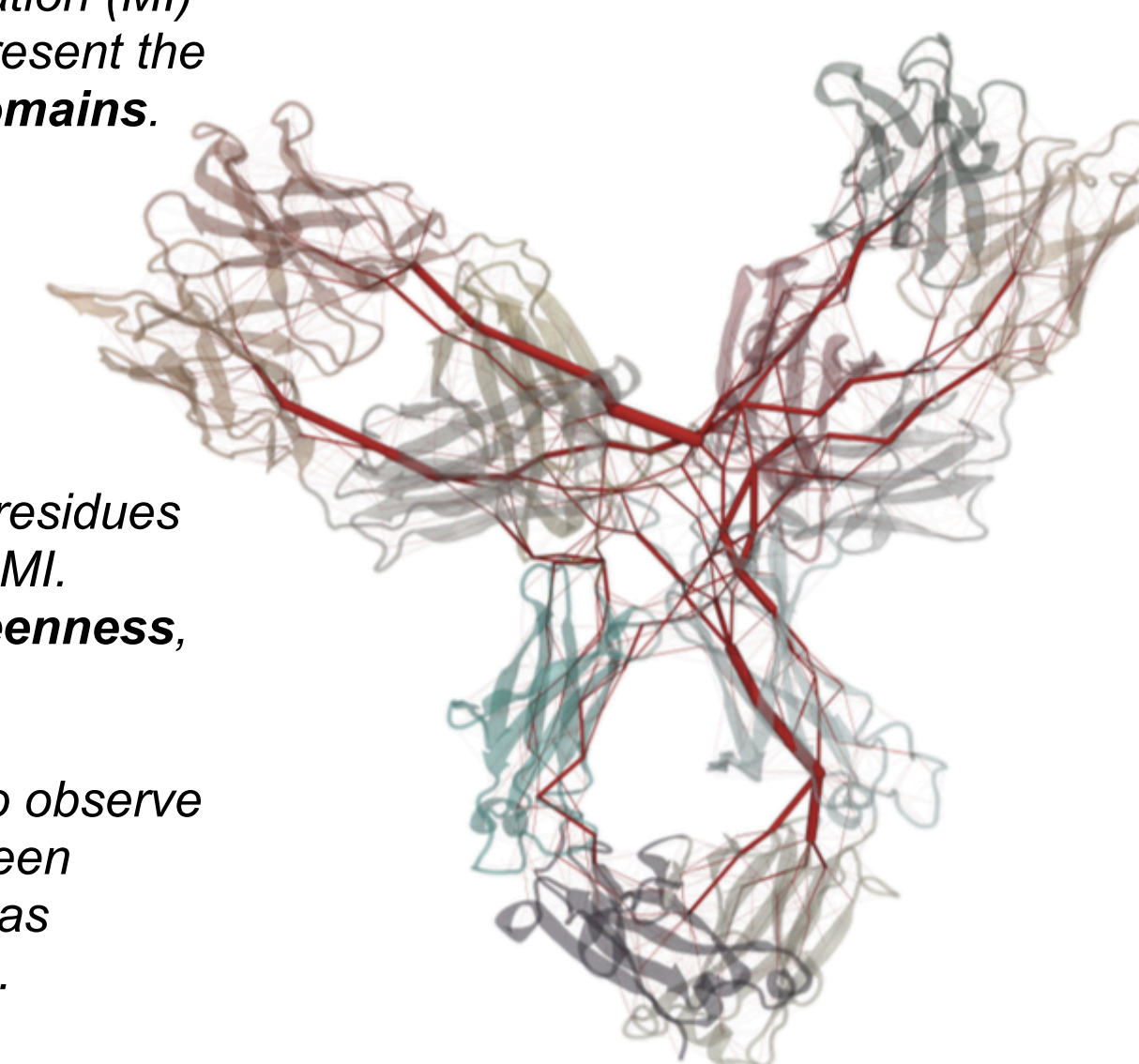
In the bottom figure (f.) the values of binding enthalpies, obtained with the MM/PBSA approach, are plotted [3].

### Network Analysis

On the basis of Mutual Information (MI) we built a network [2] that represent the connection between the domains.

The network is composed by residues and it is weighted by the MI. We computed the edge betweenness, represented in figure.

From this graph it is possible to observe the communication between Fab1 and Fab2 as well as between Fab1 and Fc.



### Conclusions

- We observe a large conformational variability, especially in the apo form. The holo system exhibits a more uniform behaviour than the apo one, suggesting a correlation with the antigen binding.
- The network analysis based on information theory gives information about the communication between the protein domains.
- Cluster 0<sub>H</sub> shows the highest structural uniformity, therefore the most stable binding mode.

### Perspectives

- Simulation of the glycosylated antibody to investigate the differences, in terms of dynamical properties, with respect to the unglycosylated systems.

### References

[1] Scapin G., Yang X., Prosdise W. W., McCoy M., Reichert P., Johnston J. M., Kashi R. S. & Strickland C. (2015). Structure of full-length human anti-PD1 therapeutic IgG4 antibody pembrolizumab. *Nat Struct Mol Biol*, 22(12):953-8.  
 [2] M. C. Melo, R. C. Bernardi, C. De La Fuente-nunez, and Z. Luthey-Schulten, "Gen-eralized correlation-based dynamical network analysis: a new high-performance approach for identifying allosteric communications in molecular dynamics trajectories," *The Journal of Chemical Physics*, vol. 153, no. 13, p. 134104, 2020.  
 [3] R. Kumari, R. Kumar, O. S. D. D. Consortium, and A. Lynn, "g\_mmpbsa: A gromacs tool for high-throughput mm-pbsa calculations," *Journal of chemical information and modeling*, vol. 54, no. 7, pp. 1951-1962, 2014.