



Introduction

Staphylococcus aureus can cause microbial keratitis [1], conjunctivitis [2] and non-infectious contact lens related corneal infiltrative events [3]. S. aureus infections can be difficult to treat because of its ability to acquire resistant to multiple antibiotics [4]. Contact lens multipurpose disinfectant solutions (MPDS) are used to disinfect contact lenses when they are not being worn. MPDSs are disinfectant used in contact lens cases in order to avoid contamination and microbial growth, but these disinfectants also lose their efficacy when become contaminated by improper use or poor handling and thus bacteria grow and develop resistance against these disinfectants.

Due to limited information available on antimicrobial and MPDS susceptibility patterns of clinical isolates of S. aureus from Australia in comparison to other countries. The purpose of this study was to investigate the antibiotic and MPDS sensitives of S. aureus isolates from different ocular surface conditions isolated in Australia and the USA.

Materials and Methods

63 S. aureus strains: 23 microbial keratitis [MK], 26 conjunctivitis and 14 strains from non-infectious contact lens-related corneal infiltrative [niCIE] were selected. The minimum inhibitory concentration (MIC) was established for the isolates using the broth microdilution method in 96-well plates to give final concentration ranging from 5120 µg/ml to 0.25 µg/ml. The strains were classified as sensitive or resistant by using CLSI and EUCAST breakpoints.

The antimicrobials tested were four multi-purpose disinfecting solutions and eight antibiotics. All the dilutions for antimicrobials were made in phosphate buffer saline. The bacterial inoculum was prepared in Mueller-Hinton broth.

Statistical analysis: Differences in the frequency of antibiotic susceptibility between infectious and non-infectious groups from Australia and the USA, were compared using Fisher's exact test. For statistical analysis, p values of ≤ 0.05 were considered significant, and p values ≤0.1 were considered to show a trend for differences.

Results

Table.1: Frequency of antibiotic susceptibility of S. aureus isolates from MK or conjunctivitis from USA and Australia.

Table with 5 columns: Ocular Condition, Antibiotics, USA (% susceptible), Australia (% susceptible), P-value. Rows include MK and Infectious Conjunctivitis for various antibiotics like Ciprofloxacin, Ceftazidime, Oxacillin, etc.

Fig.1: MIC of S. aureus niCIE strains to MPDS

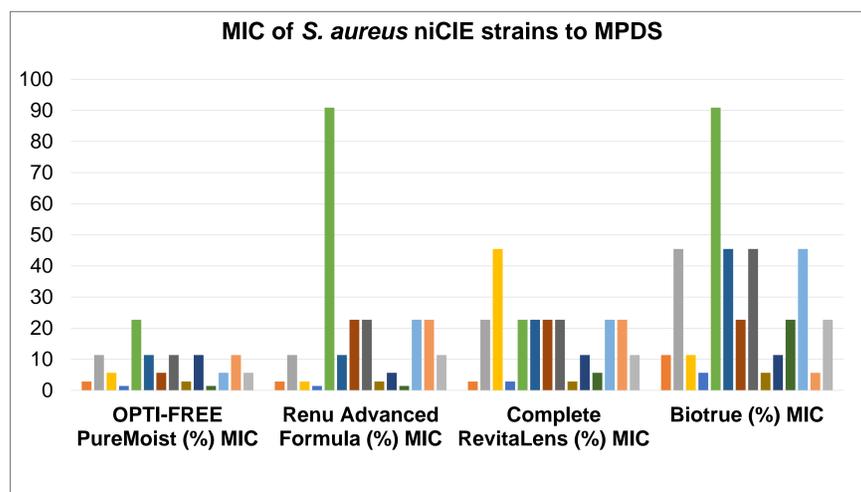


Table.3: Relative susceptibilities of contact lens related niCIE isolates to antibiotics and MPDS

Table with 13 columns: S. aureus Strains, CIP, CEFT, OXA, GEN, VAN, CHL, AZI, P-B, OPTI, RENU, REV, BIO. Rows list strain numbers (12, 20, 24, 25, 27, 28, 32, 33, 48, 117, 26, 29, 31, 41) and shading indicates susceptibility/resistance.

No shading indicates that strains were susceptible, and grey indicates they were resistant. CIP, Ciprofloxacin; CEFT, Ceftazidime; OXA, Oxacillin; GEN, Gentamicin; VAN, Vancomycin; CHL, Chloramphenicol; AZI, Azithromycin; P-B, Polymyxin B; OPTI, OPTI-FREE PureMoist; RENU, Renu Advanced Formula; REV, Complete RevitaLens OcuTec; BIO, Biotrue.

Discussion and Conclusion

- All strains were susceptible to vancomycin (100%) and gentamicin (98%).
The frequent use of ciprofloxacin in the USA and chloramphenicol in Australia could explain low sensitivity of strains to these antibiotics.
OPTI-FREE PureMoist was the most effective and Biotrue was the least effective multi-purpose solutions against S. aureus niCIE isolates.
There was no concordance between antibiotic and MPDS sensitivity, so antibiotic sensitivity was not a good predictor of resistance to MPDS.
Further whole genome sequence will help to better understand resistance mechanisms of S. aureus from different ocular conditions.

References

1. O'Callaghan, R.J. The Pathogenesis of Staphylococcus aureus Eye Infections. Pathogens (Basel, Switzerland) 2018, 7, 9, doi:10.3390/pathogens7010009.
2. Solberg, R.; Meberg, A.; Schøyen, R. [Neonatal conjunctivitis in a nursery and a neonatal unit]. Tidsskr Nor Laegeforen 1991, 111, 1230-1232.
3. Sweeney, D.F.; Jalbert, I.; Covey, M.; Sankaridurg, P.R.; Vajdic, C.; Holden, B.A.; Sharma, S.; Ramachandran, L.; Willcox, M.D.; Rao, G.N. Clinical characterization of corneal infiltrative events observed with soft contact lens wear. Cornea 2003, 22, 435-442, doi:10.1097/00003226-200307000-00009.
4. Chambers, H.F.; Deleo, F.R. Waves of resistance: Staphylococcus aureus in the antibiotic era. Nat Rev Microbiol 2009, 7, 629-641, doi:10.1038/nrmicro2200.
5. Otto, M. Staphylococcus aureus toxins. Curr Opin Microbiol 2014, 17, 32-37, doi:https://doi.org/10.1016/j.mib.2013.11.004.

Table.2: Frequency of antibiotic susceptibility of Australian S. aureus isolates from infectious and non-infectious ocular conditions.

Table with 4 columns: Antibiotics, Ocular Condition (Infectious, Non-infectious Corneal Infiltrative events), P-value. Rows include Ciprofloxacin, Ceftazidime, Oxacillin, Gentamicin, Vancomycin, Chloramphenicol, Azithromycin, Polymyxin B.